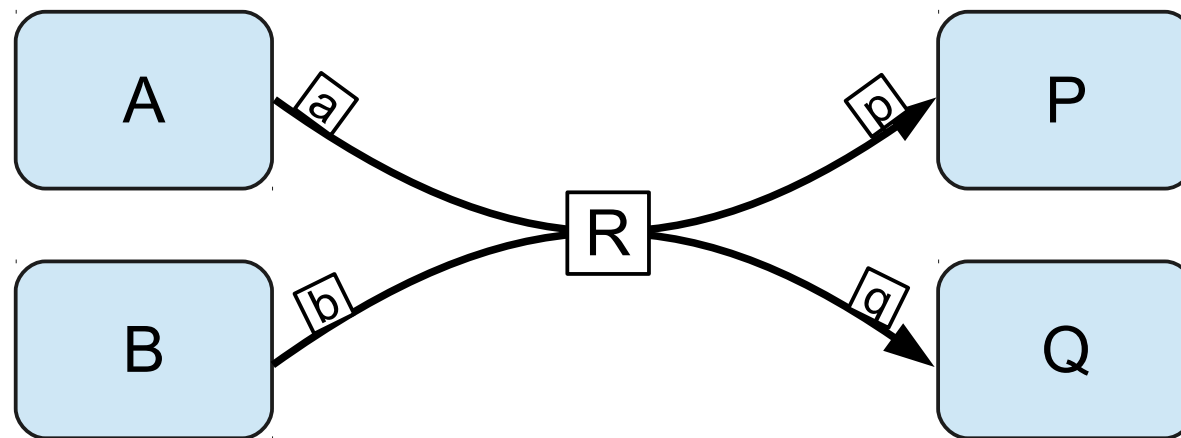


# Modelling chemical kinetics

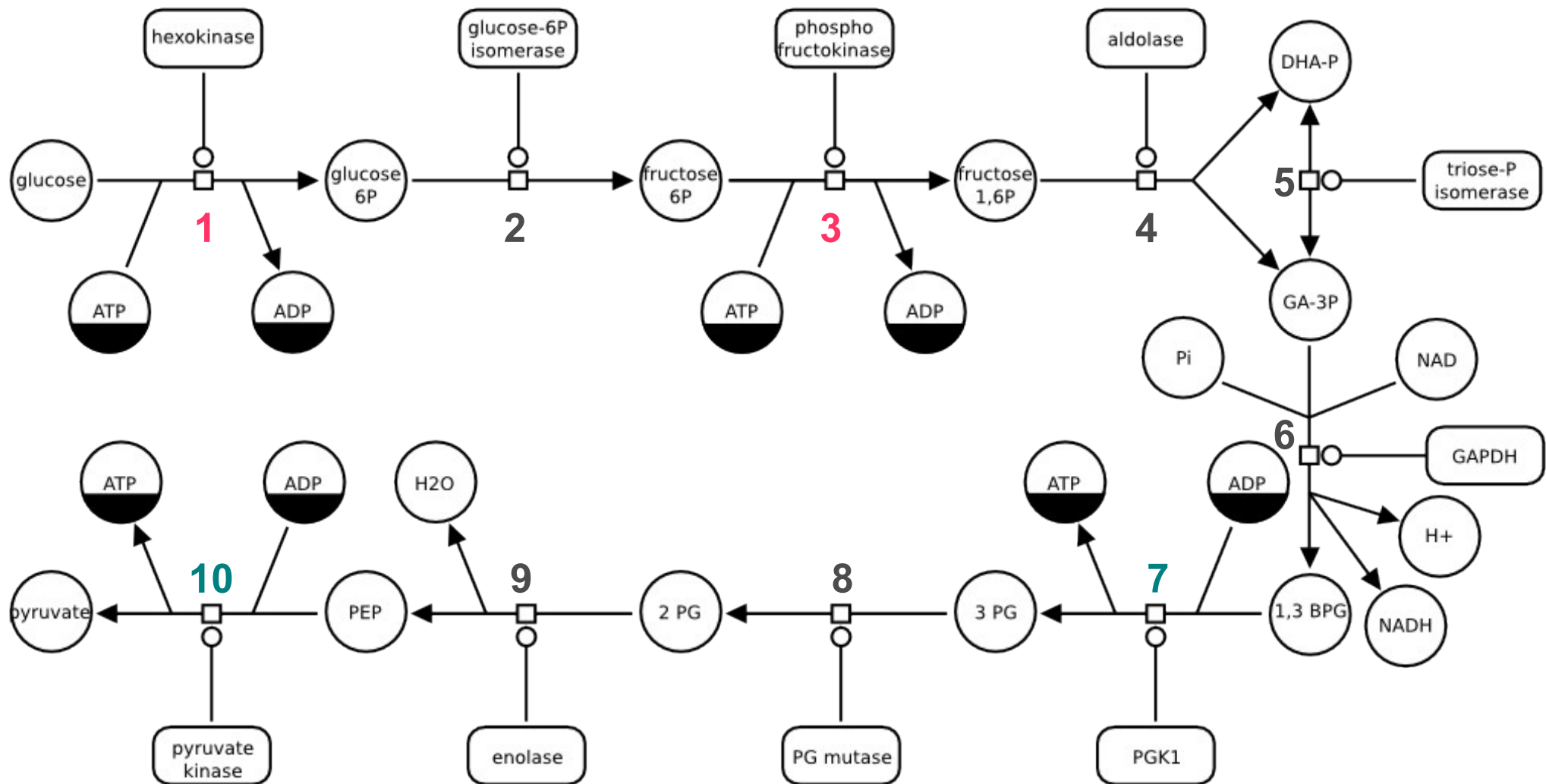
Nicolas Le Novère, Babraham Institute, EMBL-EBI

# Systems Biology models $\neq$ ODE models

- Reconstruction of state variable evolution from process descriptions:
- Processes can be combined in ODEs (for deterministic simulations); transformed in propensities (for stochastic simulations)
- Systems can be reconfigured quickly by adding or removing a process

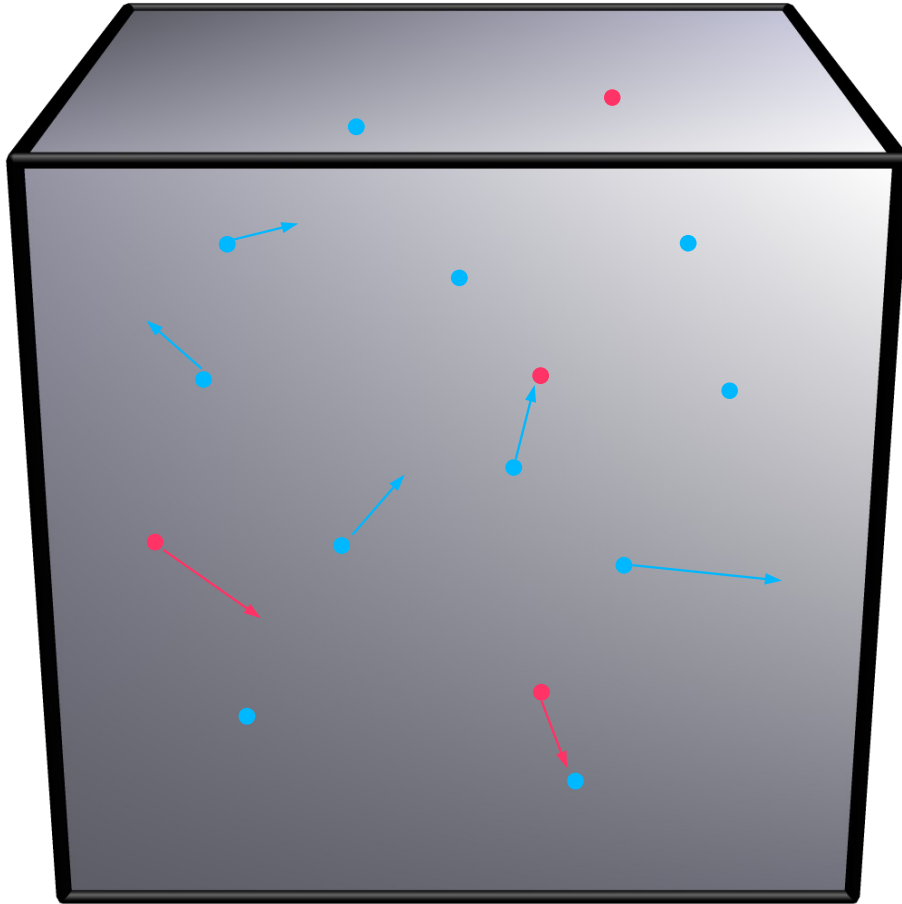


substances A and B are consumed by reaction R that produces substances P and Q

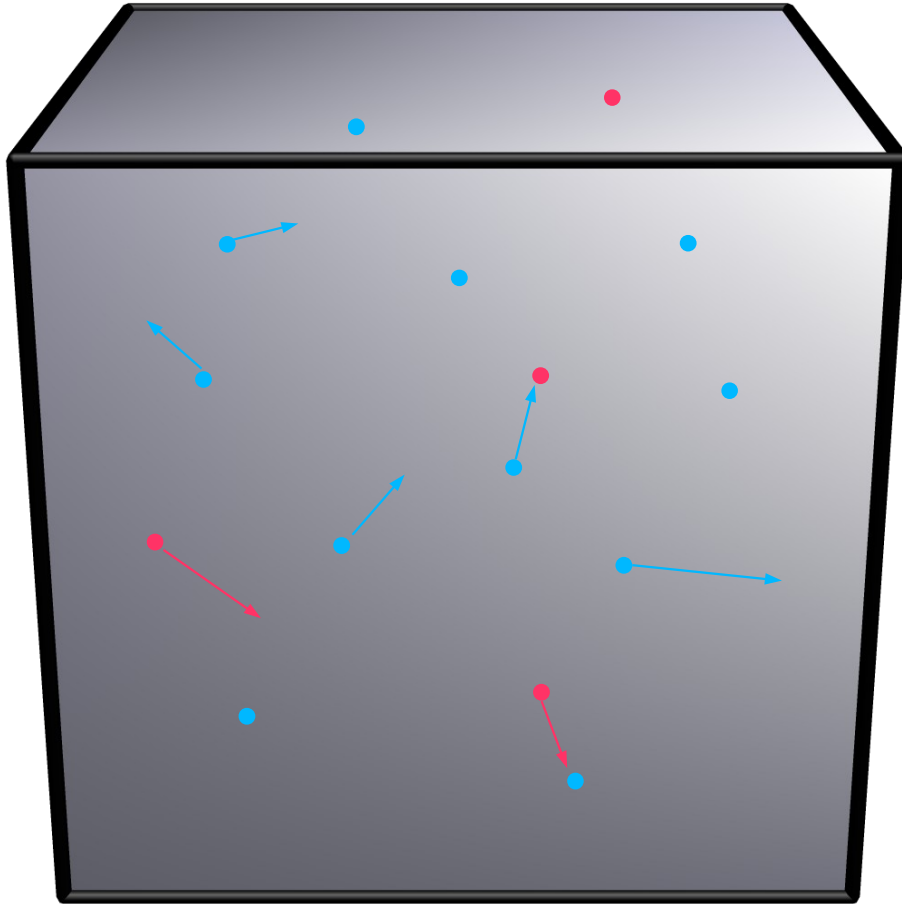


ATP is consumed by processes **1** and **3**, and produced by processes **7** and **10**  
(for 1 reactions **1** and **3**, there are 2 reactions **7** and **10**)

# Statistical physics and chemical reaction



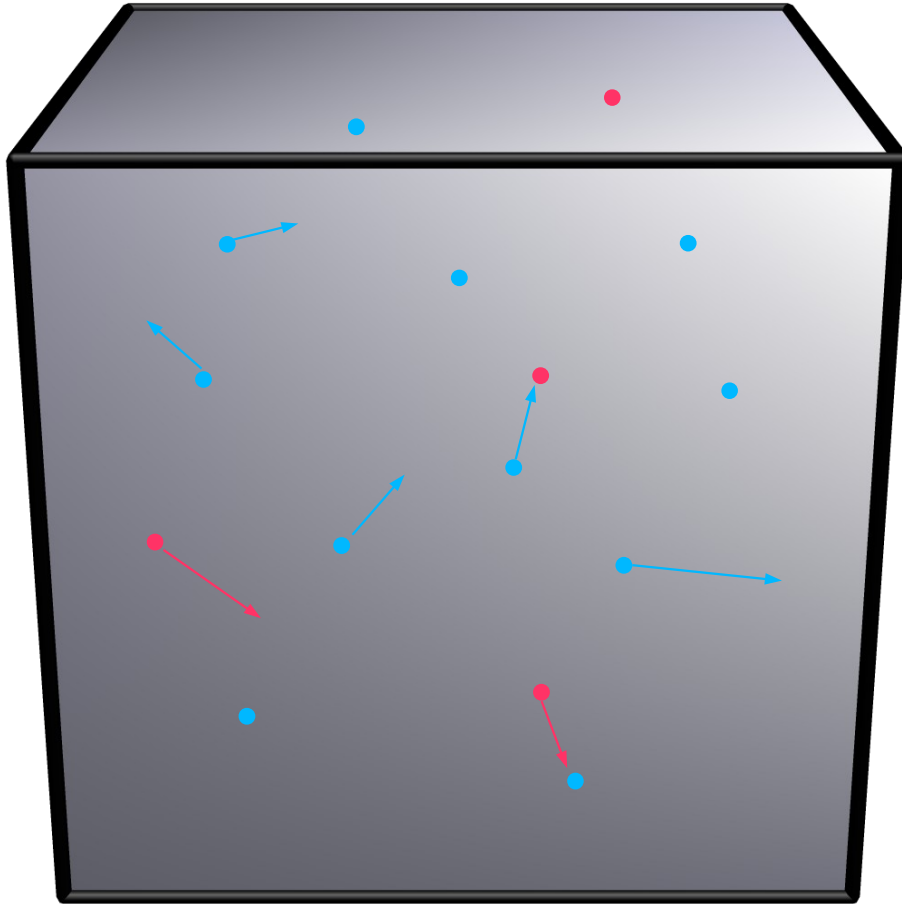
# Statistical physics and chemical reaction



Probability to find an object in a container within an interval of time

$$P(\bullet) \propto \frac{n(\bullet)}{V} = [\bullet]$$

# Statistical physics and chemical reaction



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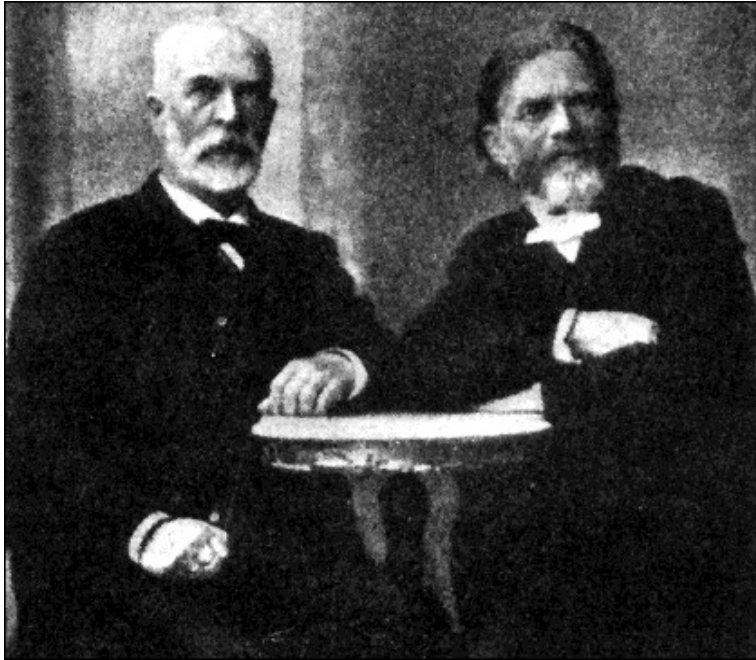
$$P(\text{reaction } \bullet + \bullet) = P(\bullet) \times P(\bullet) \times P(\bullet \text{ reacts with } \bullet)$$

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# Law of Mass Action

Waage and Guldberg (1864)



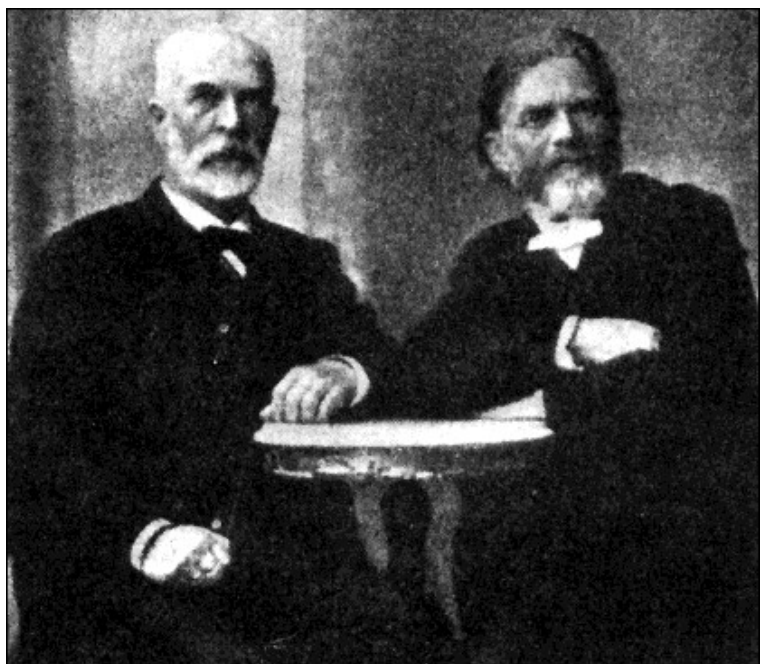
$$v = k \cdot \prod_i a_i^{n_i}$$

Diagram illustrating the Law of Mass Action equation:

- $v$ : velocity
- $k$ : rate-constant
- $a_i$ : activity
- $n_i$ : stoichiometry

# Law of Mass Action

Waage and Guldberg (1864)



$$v = k \cdot \prod_i a_i^{n_i}$$

Diagram illustrating the Law of Mass Action equation:  $v = k \cdot \prod_i a_i^{n_i}$ . The variables are labeled as follows:

- $v$ : velocity (indicated by an arrow from the label "velocity" to  $v$ )
- $k$ : rate-constant (indicated by an arrow from the label "rate-constant" to  $k$ )
- $a_i$ : activity (indicated by an arrow from the label "activity" to  $a_i$ )
- $n_i$ : stoichiometry (indicated by an arrow from the label "stoichiometry" to  $n_i$ )

$$v = k \cdot \prod_i P_i^{n_i} \quad \text{gas}$$

$$v = k \cdot \prod_i [X_i]^{n_i} \quad \text{solution}$$

# Evolution of a reactant

- Velocity multiplied by stoichiometry
- negative if consumption, positive if production
- Example of a unimolecular reaction  $x \xrightarrow{k} y$

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$$\frac{d[x]}{dt} = -1 \cdot v = -1 \cdot k \cdot [x]$$

$$\frac{d[y]}{dt} = +1 \cdot v = +1 \cdot k \cdot [x]$$

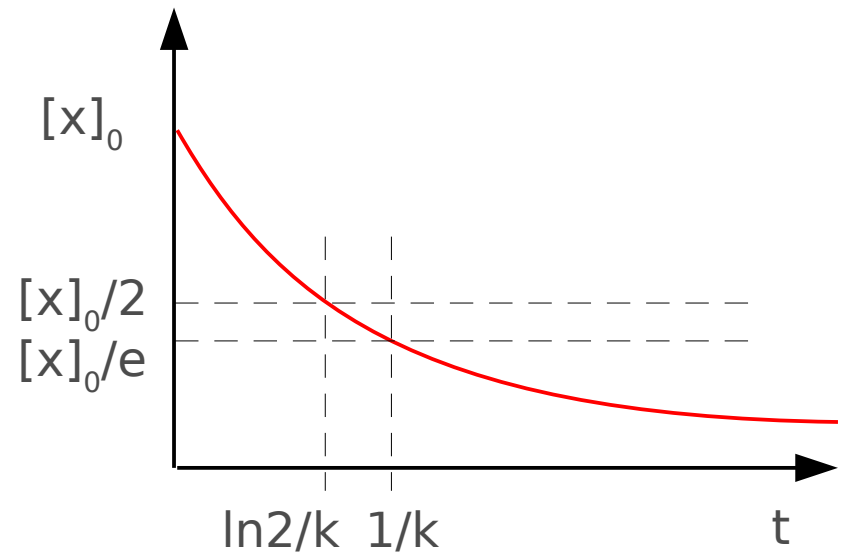
# Evolution of a reactant

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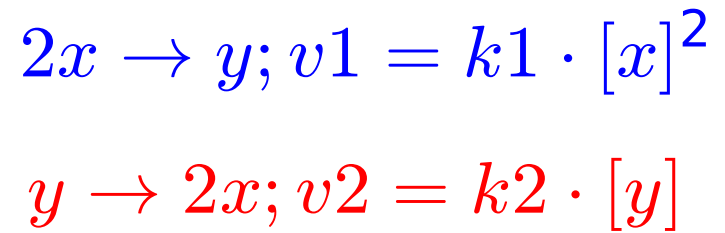
$$\frac{d[x]}{dt} = -1 \cdot v = -1 \cdot k \cdot [x]$$

$$\frac{d[y]}{dt} = +1 \cdot v = +1 \cdot k \cdot [x]$$

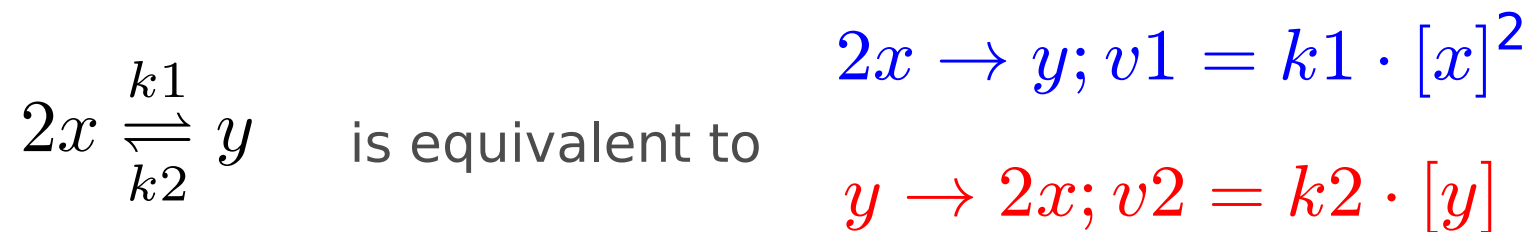
$$x(t) = [x]_0 \cdot e^{-kt}$$



# Reversible reaction



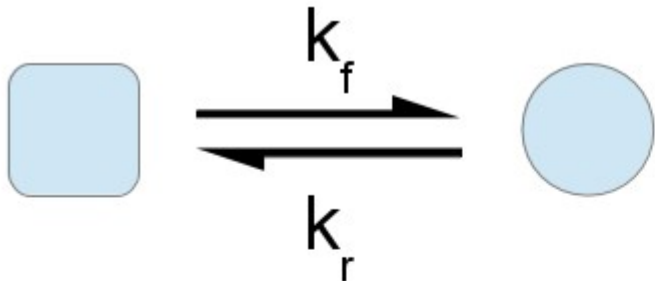
# Reversible reaction



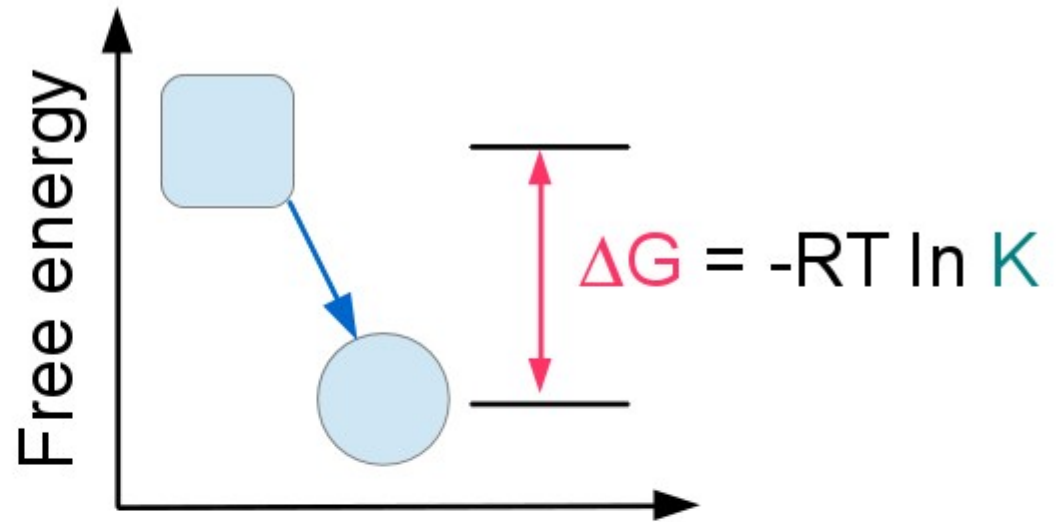
$$\frac{d[x]}{dt} = -2 \cdot v_1 + 2 \cdot v_2 = -2 \cdot k_1 \cdot [x]^2 + 2 \cdot k_2 \cdot [y]$$

$$\frac{d[y]}{dt} = +1 \cdot v_1 - 1 \cdot v_2 = +1 \cdot k_1 \cdot [x]^2 - 1 \cdot k_2 \cdot [y]$$

# Conformational equilibrium



$$K = \frac{[\text{square}]}{[\text{circle}]} = \frac{k_r}{k_f}$$



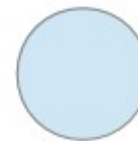
$$K < 1$$



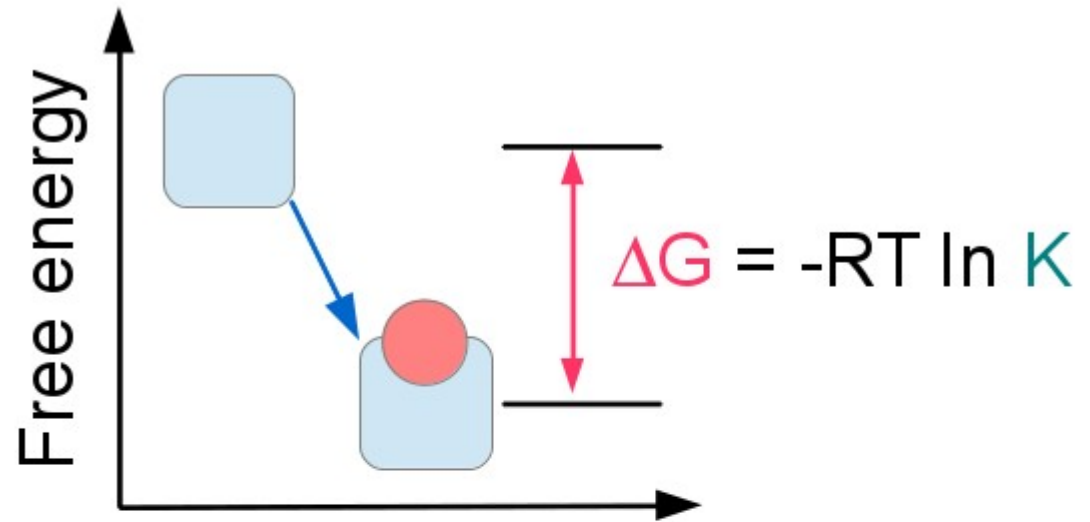
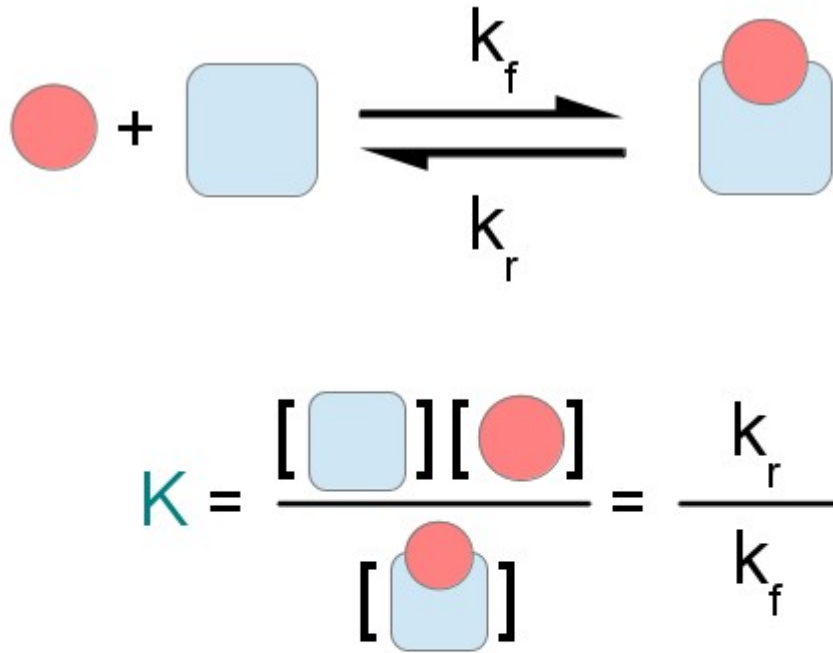
$$k_f > k_r$$

$$\Delta G > 0$$

Shift towards



# Binding equilibrium

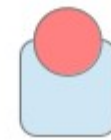


$$K < 1 \longrightarrow$$

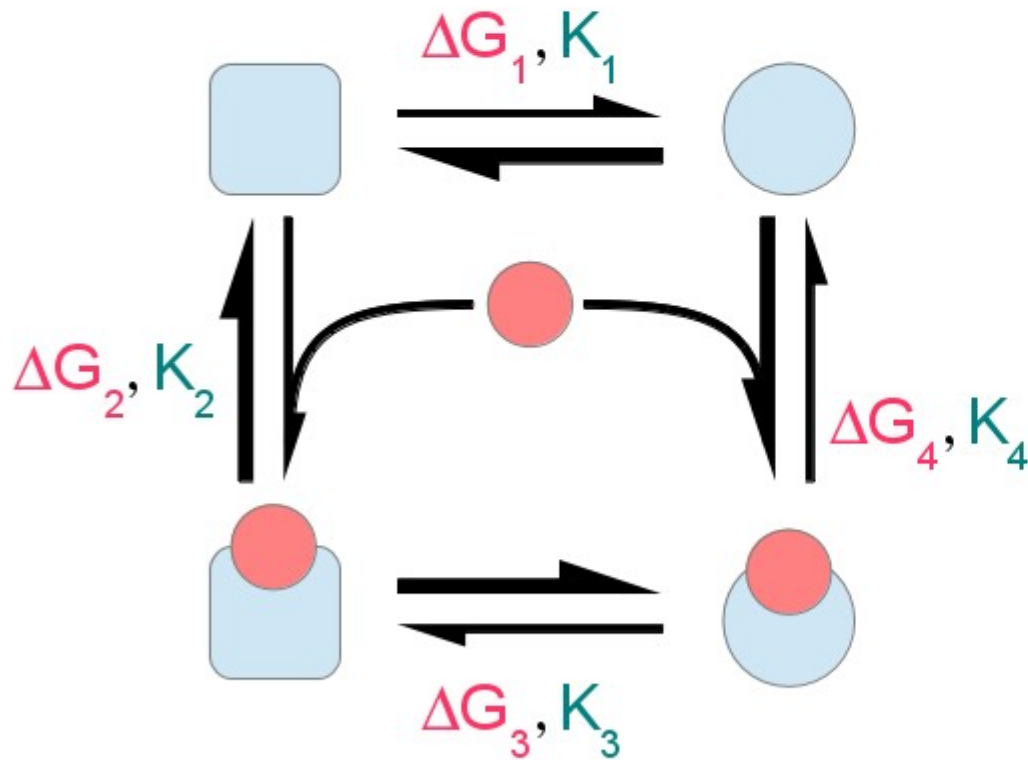
$$k_f > k_r$$

$$\Delta G > 0$$

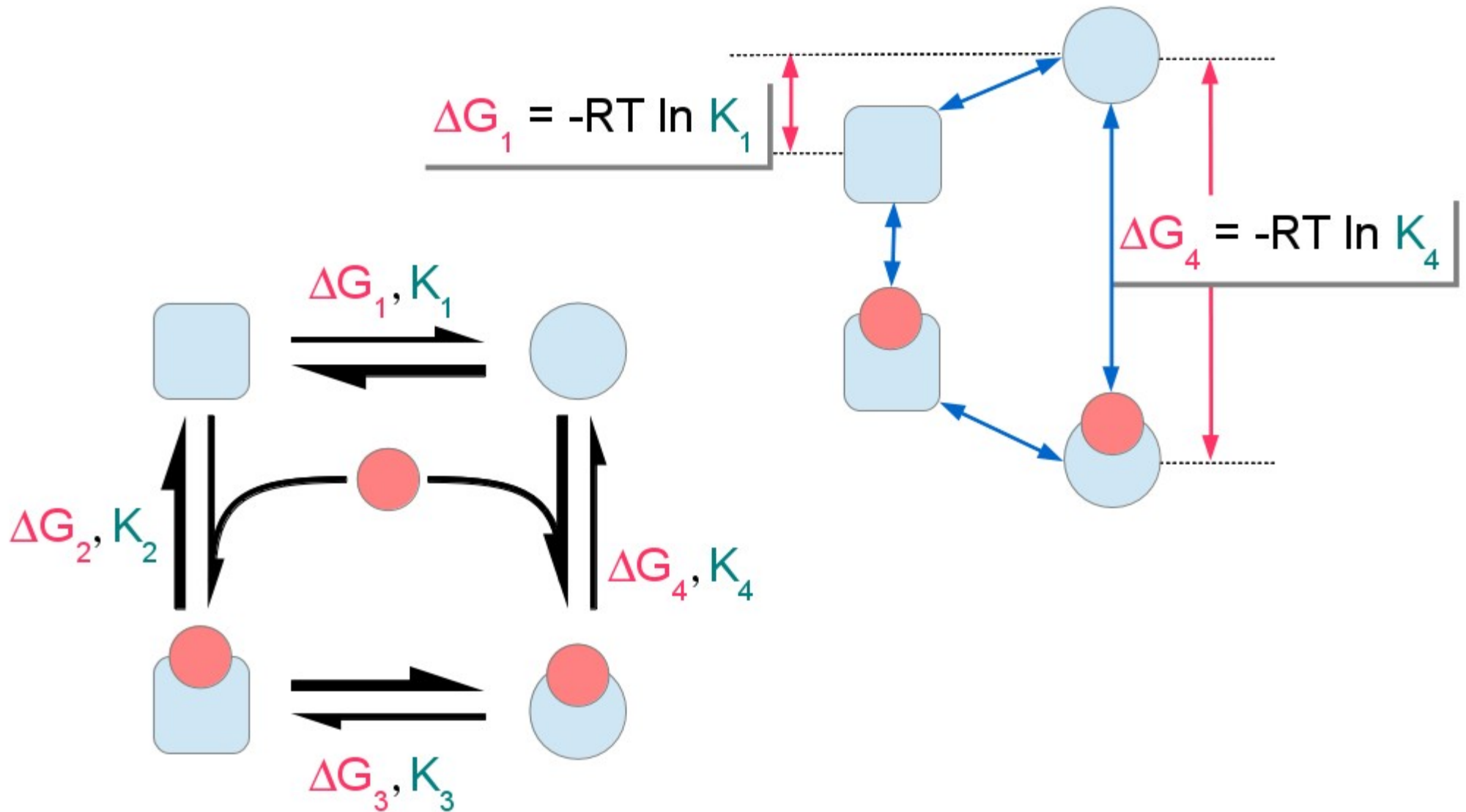
Shift towards



# How does a ligand activate its target?

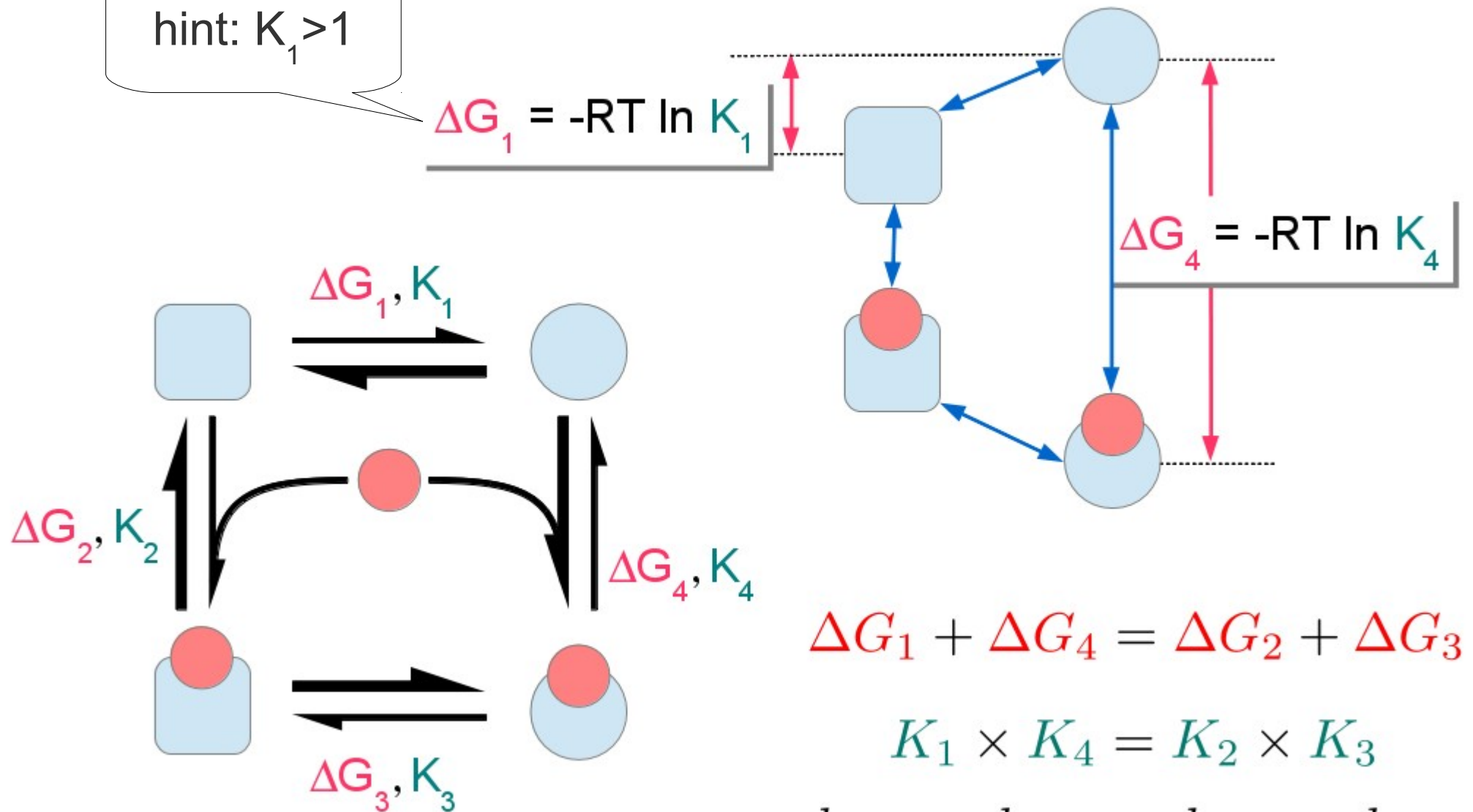


# How does a ligand activate its target?



# How does a ligand activate its target?

hint:  $K_1 > 1$



$$\Delta G_1 + \Delta G_4 = \Delta G_2 + \Delta G_3$$

$$K_1 \times K_4 = K_2 \times K_3$$

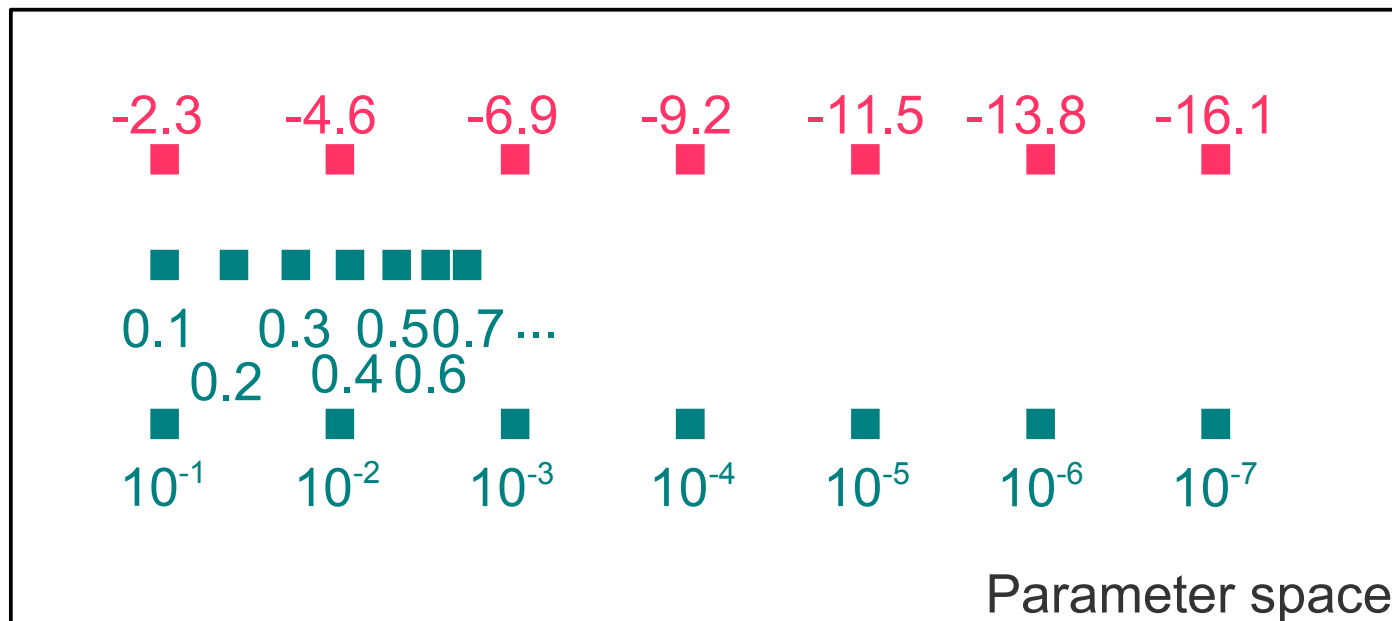
$$\frac{k_{\text{off}1}}{k_{\text{on}1}} \times \frac{k_{\text{off}4}}{k_{\text{on}4}} = \frac{k_{\text{off}2}}{k_{\text{on}2}} \times \frac{k_{\text{off}3}}{k_{\text{on}3}}$$

Add energies

Multiply constants

+1 quantum energy = constant divided by 10

Explore constants exponentially:



# Example of an enzymatic reaction



# Example of an enzymatic reaction

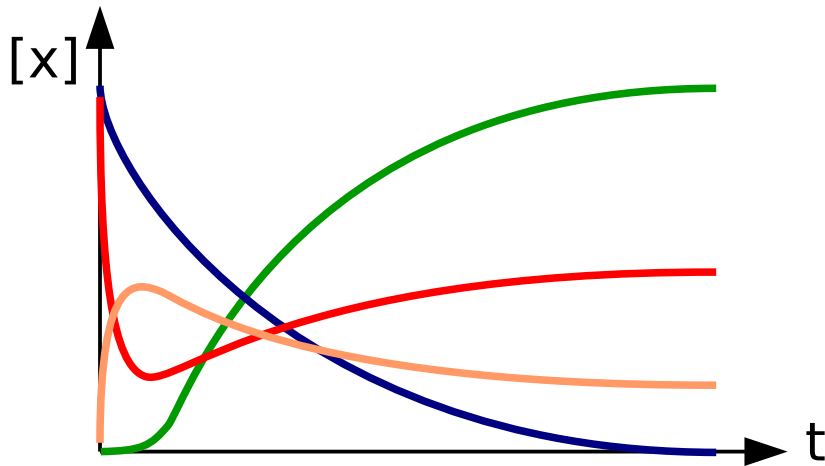


$$\begin{aligned} d[E]/dt &= -k_1[E][S] + k_2[ES] + k_3[ES] \\ d[S]/dt &= -k_1[E][S] + k_2[ES] \\ d[ES]/dt &= +k_1[E][S] - k_2[ES] - k_3[ES] \\ d[P]/dt &= +k_3[ES] \end{aligned}$$

# Example of an enzymatic reaction



$$\begin{aligned} d[E]/dt &= -k_1[E][S] + k_2[ES] + k_3[ES] \\ d[S]/dt &= -k_1[E][S] + k_2[ES] \\ d[ES]/dt &= +k_1[E][S] - k_2[ES] - k_3[ES] \\ d[P]/dt &= +k_3[ES] \end{aligned}$$



Not feasible in general



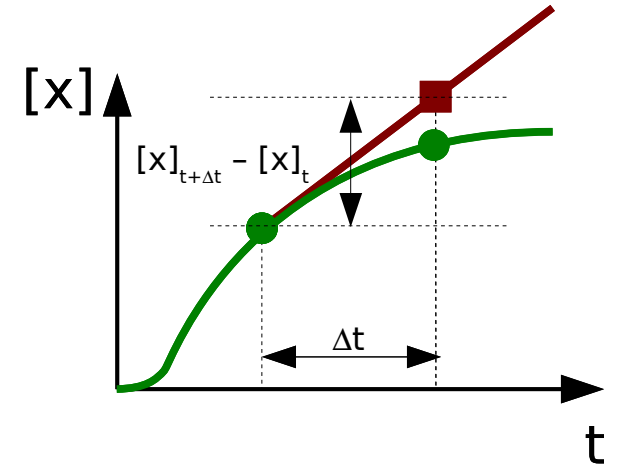
Numerical integration

# Numerical integration

Euler method:

$$d[x]/dt \approx ([x]_{t+\Delta t} - [x]_t)/\Delta t$$

$$[x]_{t+\Delta t} \approx [x]_t + d[x]/dt \cdot \Delta t$$

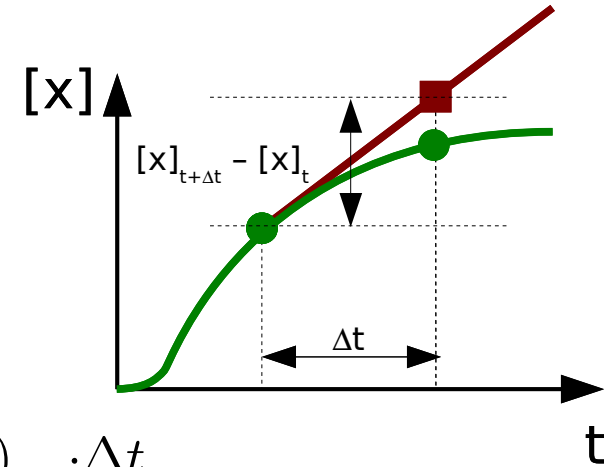


# Numerical integration

Euler method:

$$d[x]/dt \approx ([x]_{t+\Delta t} - [x]_t) / \Delta t$$

$$[x]_{t+\Delta t} \approx [x]_t + d[x]/dt \cdot \Delta t$$



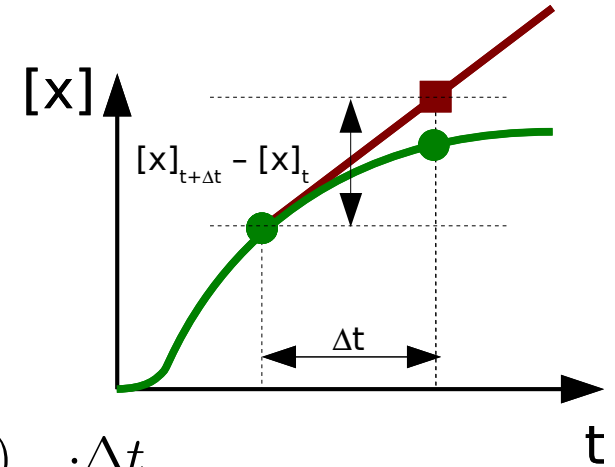
$$\begin{aligned}
 [E]_{t+\Delta t} &= [E]_t + ((k_2 + k_3)[ES]_t - k_1[E]_t[S]_t) \cdot \Delta t \\
 [S]_{t+\Delta t} &= [S]_t + (k_2[ES]_t - k_1[E]_t[S]_t) \cdot \Delta t \\
 [ES]_{t+\Delta t} &= [ES]_t + (k_1[E]_t[S]_t - (k_2 + k_3)[ES]_t) \cdot \Delta t \\
 [P]_{t+\Delta t} &= [P]_t + k_3[ES]_t \cdot \Delta t
 \end{aligned}$$

# Numerical integration

Euler method:

$$d[x]/dt \approx ([x]_{t+\Delta t} - [x]_t) / \Delta t$$

$$[x]_{t+\Delta t} \approx [x]_t + d[x]/dt \cdot \Delta t$$

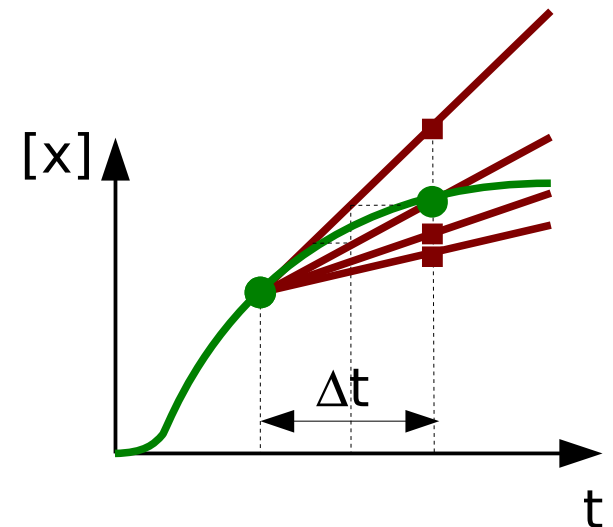


$$\begin{aligned} [E]_{t+\Delta t} &= [E]_t + ((k_2 + k_3)[ES]_t - k_1[E]_t[S]_t) \cdot \Delta t \\ [S]_{t+\Delta t} &= [S]_t + (k_2[ES]_t - k_1[E]_t[S]_t) \cdot \Delta t \\ [ES]_{t+\Delta t} &= [ES]_t + (k_1[E]_t[S]_t - (k_2 + k_3)[ES]_t) \cdot \Delta t \\ [P]_{t+\Delta t} &= [P]_t + k_3[ES]_t \cdot \Delta t \end{aligned}$$

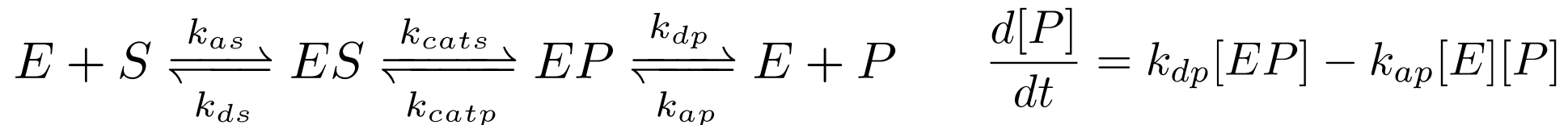
4<sup>th</sup> order Runge-Kutta:

$$[x]_{t+\Delta t} \approx [x]_t + (F_1 + 2F_2 + 2F_3 + F_4) / 6 \cdot \Delta t$$

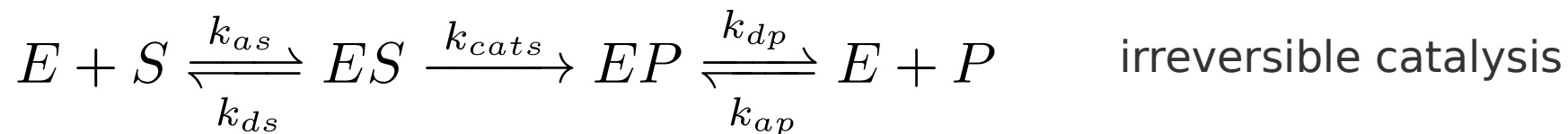
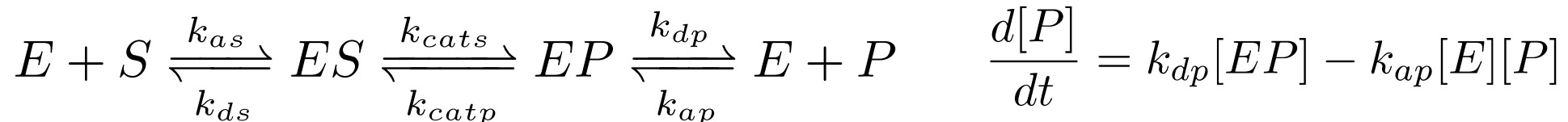
$$\begin{aligned} \text{with } F_1 &= d[x]/dt = f([x]_t, t) \\ F_2 &= f([x]_t + \Delta t/2 \cdot F_1, t + \Delta t/2) \\ F_3 &= f([x]_t + \Delta t/2 \cdot F_2, t + \Delta t/2) \\ F_4 &= f([x]_t + \Delta t \cdot F_3, t + \Delta t) \end{aligned}$$



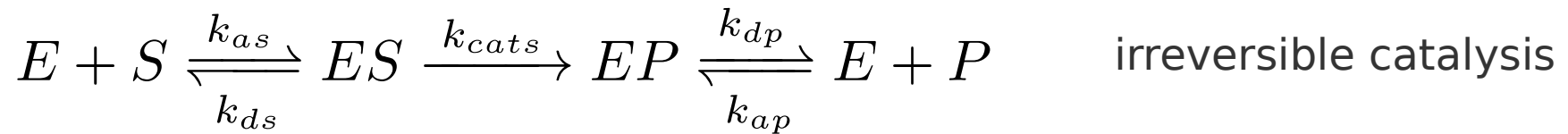
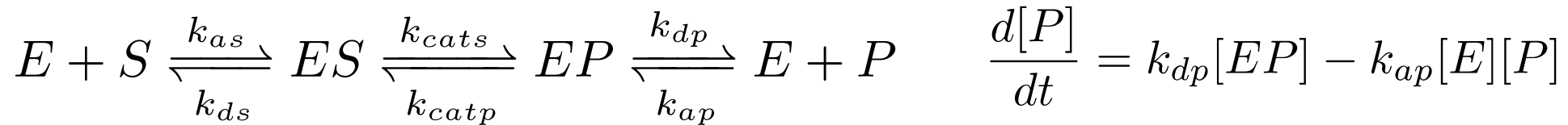
# Choose the right formalism



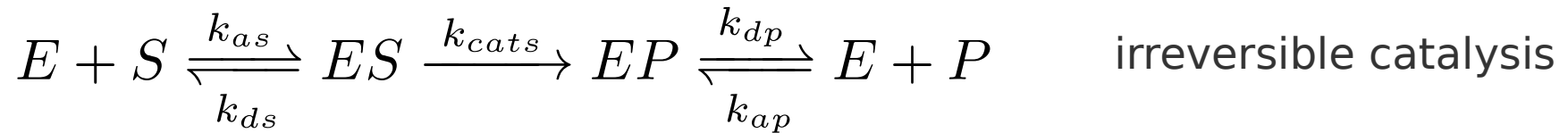
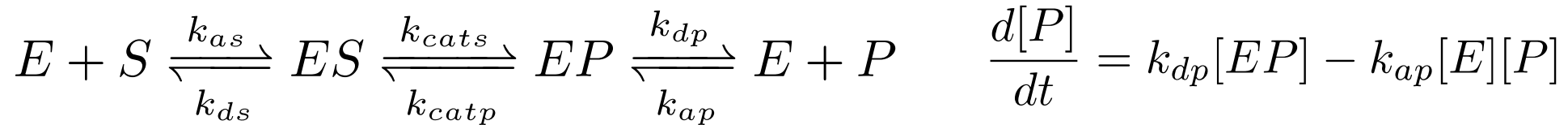
# Choose the right formalism



# Choose the right formalism



# Choose the right formalism



$$\frac{d[P]}{dt} = [E]k_{\text{cat}} \frac{[S]}{K_m + [S]}$$

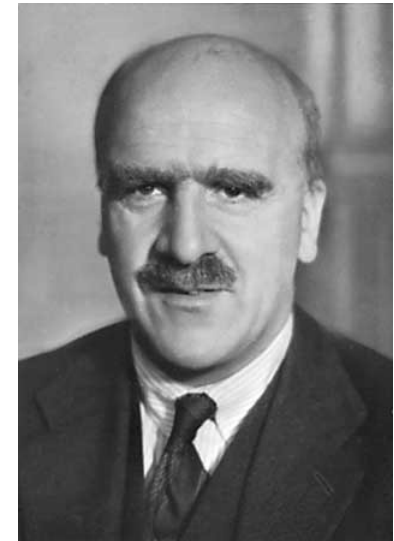
# Enzyme kinetics

Victor Henri (1903) *Lois Générales de l'Action des Diastases*. Paris, Hermann.

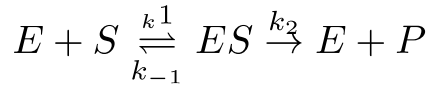


Leonor Michaelis, Maud Menten (1913). *Die Kinetik der Invertinwirkung*, *Biochem. Z.* 49:333-369

George Edward Briggs and John Burdon Sanderson Haldane (1925) *A note on the kinetics of enzyme action*, *Biochem. J.*, 19: 338-339



# Briggs-Haldane on Henri-Michaelis-Menten



$$\frac{d[P]}{dt} = k_2[ES]$$

$$[E] = [E_0] - [ES]$$

$$\frac{d[ES]}{dt} = k_1[E][S] - k_{-1}[ES] - k_2[ES] = 0$$

$$[ES] \frac{K_m}{[S]} = [E_0] - [ES]$$

$$[ES] = \frac{k_1[E][S]}{k_{-1} + k_2}$$

$$[ES] \left(1 + \frac{K_m}{[S]}\right) = [E_0]$$

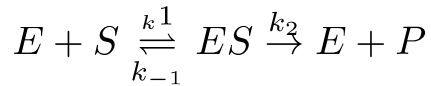
$$K_m = \frac{k_{-1} + k_2}{k_1}$$

$$[ES] = [E_0] \frac{1}{1 + \frac{K_m}{[S]}}$$

$$[ES] = \frac{[E][S]}{K_m}$$

$$\frac{d[P]}{dt} = k_2[E_0] \frac{[S]}{K_m + [S]} = V_{max} \frac{[S]}{K_m + [S]}$$

# Briggs-Haldane on Henri-Michaelis-Menten



$$\frac{d[ES]}{dt} = k_1[E][S] - k_{-1}[ES] - k_2[ES] = 0$$

steady-state!!!

$$[ES] = \frac{k_1[E][S]}{k_{-1} + k_2}$$

$$K_m = \frac{k_{-1} + k_2}{k_1}$$

$$[ES] = \frac{[E][S]}{K_m}$$

$$\frac{d[P]}{dt} = k_2[ES]$$

$$[E] = [E_0] - [ES]$$

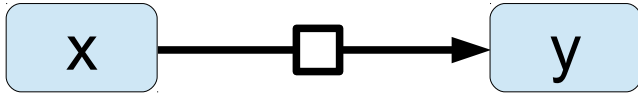
$$[ES] \frac{K_m}{[S]} = [E_0] - [ES]$$

$$[ES] \left(1 + \frac{K_m}{[S]}\right) = [E_0]$$

$$[ES] = [E_0] \frac{1}{1 + \frac{K_m}{[S]}}$$

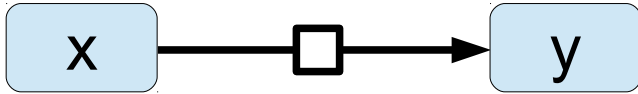
$$\frac{d[P]}{dt} = k_2[E_0] \frac{[S]}{K_m + [S]} = V_{max} \frac{[S]}{K_m + [S]}$$

# Generalisation: activators

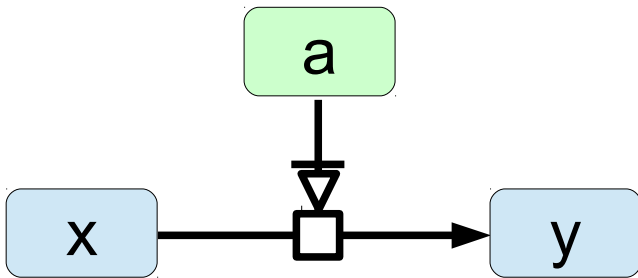
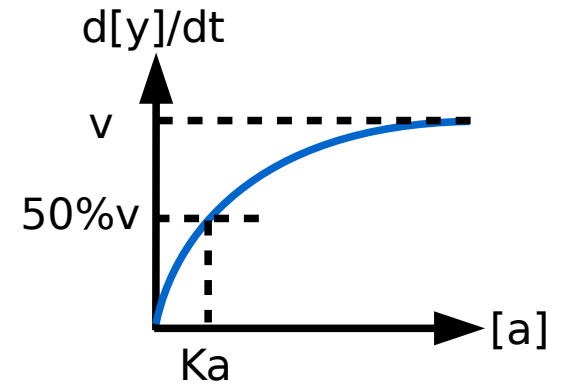


$$\frac{d[y]}{dt} = v(= k \cdot [x])$$

# Generalisation: activators



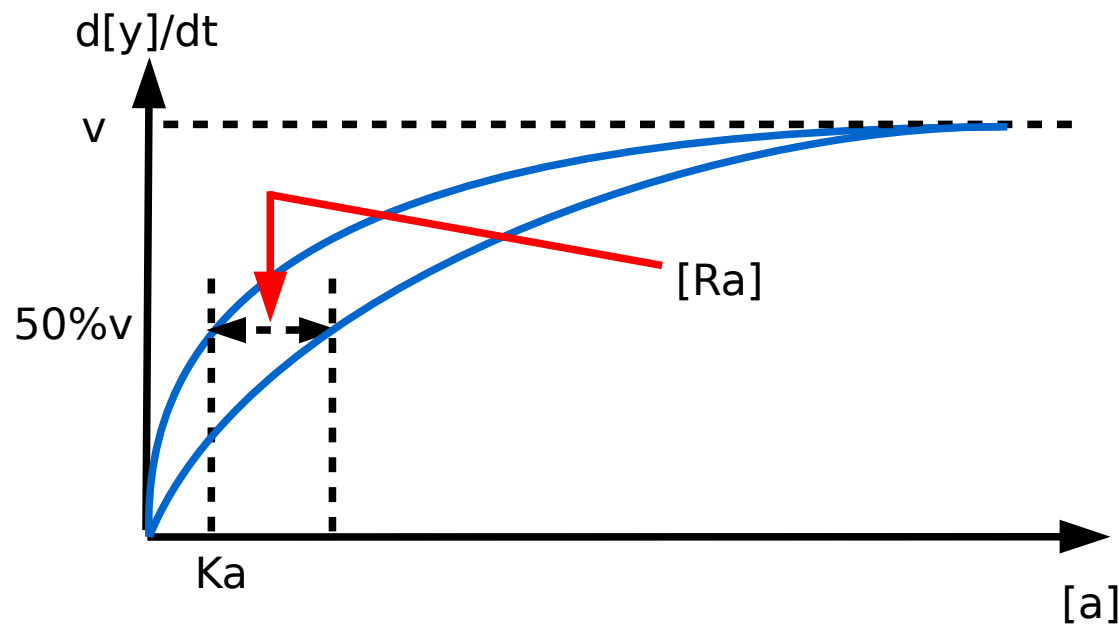
$$\frac{d[y]}{dt} = v (= k \cdot [x])$$



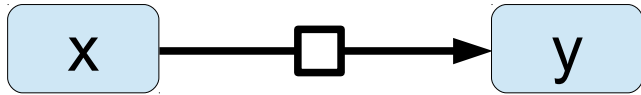
$$\frac{d[y]}{dt} = v \cdot \frac{[a]}{Ka + [a]}$$

# Beware of the ligand depletion!

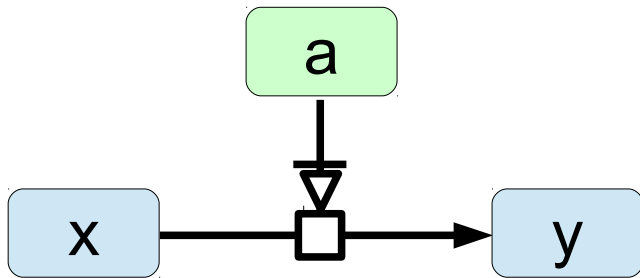
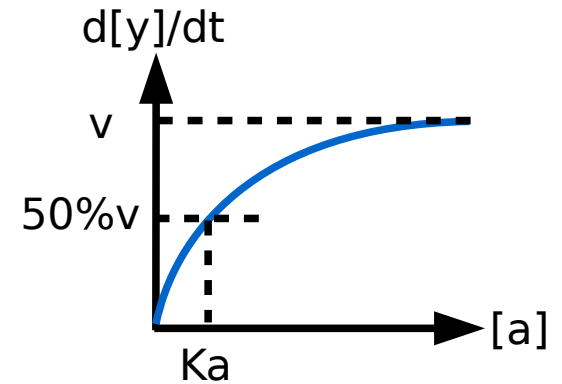
[a] is the concentration of **FREE** activator. In dose-response experiments, the x-axis most often represents the **TOTAL** concentration (initial concentration). The two are equal only if the concentration of sensor (receptor, enzyme etc.) is much lower than the  $K_a$ .



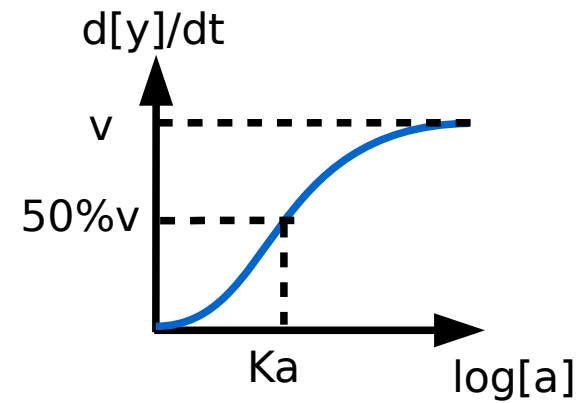
# Generalisation: activators



$$\frac{d[y]}{dt} = v(= k \cdot [x])$$



$$\frac{d[y]}{dt} = v \cdot \frac{[a]}{Ka + [a]}$$

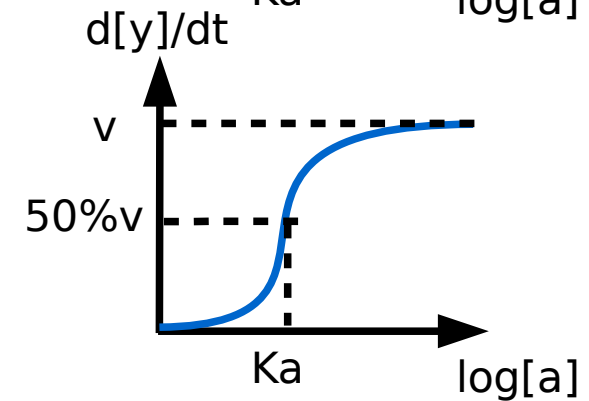
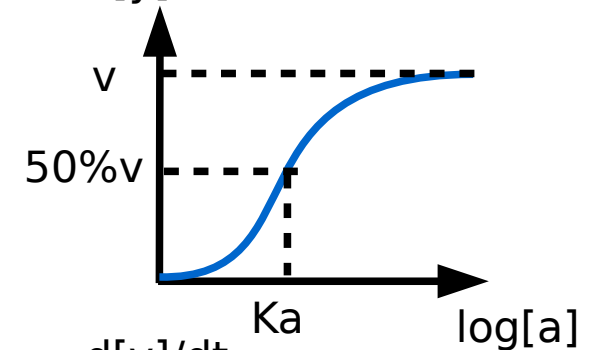
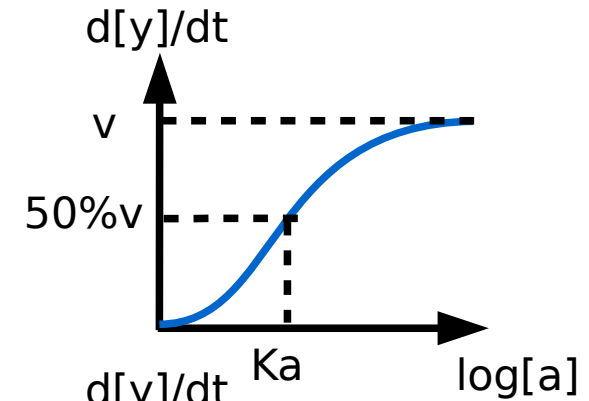


# Phenomenological ultrasensitivity

$$\frac{d[y]}{dt} = v \cdot \frac{[a]}{Ka + [a]}$$

$$\frac{d[y]}{dt} = v \cdot \frac{[a]^2}{Ka^2 + [a]^2}$$

$$\frac{d[y]}{dt} = v \cdot \frac{[a]^n}{Ka^n + [a]^n}$$



# Origins of cooperativity: Hill

iv PROCEEDINGS OF THE PHYSIOLOGICAL

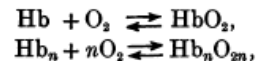
**The possible effects of the aggregation of the molecules of hæmoglobin on its dissociation curves.** By A. V. HILL.

In a previous communication Barcroft and I gave evidence which seemed to us to prove conclusively that dialysed hæmoglobin consists simply of molecules containing each one atom of iron. The molecular weight is therefore  $Hb = 16,660$ . These experiments have not been published yet, but I shall assume the results.

Other observers (Reid, Roaf, Hüfner and Gansser) working on different solutions have obtained divergent results. The method used by all of them was the direct estimation of the osmotic pressure, by means of a membrane permeable to salts, but not to hæmoglobin. The method involves a relatively large error, because the quantity measured is small. It is doubtful however whether this can explain the discordant results.

Our work led me to believe that the divergence between the results of different observers was due to an aggregation of the hæmoglobin molecules by the salts present in the solution, a consequent lowering of the number of molecules, and an increase in the average molecular weight as observed by the osmotic pressure method. To test this hypothesis I have applied it to several of the dissociation curves obtained by Barcroft and Camis with hæmoglobin in solutions of various salts, and with hæmoglobin prepared by Bohr's method.

The equation for the reaction would be



where  $Hb_n$  represents the aggregate of  $n$  molecules of  $Hb$ . I have supposed that in every solution there are many different sized aggregates, corresponding to many values of  $n$ .

If there were in the solution only  $Hb$  and  $Hb_2$  the dissociation curve would be

$$y = \lambda \frac{K'x^2}{1 + K'x^2} + (100 - \lambda) \frac{Kx}{1 + Kx} \dots\dots\dots (A),$$

where  $\lambda\%$  is as  $Hb_2$ ,  $(100 - \lambda)\%$  as  $Hb$ ,  $K'$  is the equilibrium constant of the reaction  $Hb_2 + 2O_2 \rightleftharpoons Hb_2O_4$  and  $K$  that of  $Hb + O_2 \rightleftharpoons HbO_2$ :  $K$  has the value .125 (Barcroft and Roberts).

Hill (1910) *J Physiol* 40: iv-vii.



# Origins of cooperativity: Hill

iv PROCEEDINGS OF THE PHYSIOLOGICAL

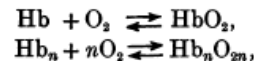
**The possible effects of the aggregation of the molecules of hæmoglobin on its dissociation curves.** By A. V. HILL.

In a previous communication Barcroft and I gave evidence seemed to us to prove conclusively that dialysed hæmoglobin simply of molecules containing each one atom of iron. The weight is therefore  $\text{Hb} = 16,660$ . These experiments have been published yet, but I shall assume the results.

Other observers (Reid, Roaf, Hüfner and Gansser) have obtained different solutions have obtained divergent results. The reason by all of them was the direct estimation of the osmotic pressure means of a membrane permeable to salts, but not to hæmoglobin. This method involves a relatively large error, because the quantity of hæmoglobin is small. It is doubtful however whether this can explain the discordant results.

Our work led me to believe that the divergence between the results of different observers was due to an aggregation of the hæmoglobin molecules by the salts present in the solution, a consequent lowering of the number of molecules, and an increase in the average molecular weight as observed by the osmotic pressure method. To test this hypothesis I have applied it to several of the dissociation curves obtained by Barcroft and Camis with hæmoglobin in solutions of various salts, and with hæmoglobin prepared by Bohr's method.

The equation for the reaction would be



where  $\text{Hb}_n$  represents the aggregate of  $n$  molecules of Hb. I have supposed that in every solution there are many different sized aggregates, corresponding to many values of  $n$ .

If there were in the solution only Hb and  $\text{Hb}_2$  the dissociation curve would be

$$y = \lambda \frac{K'x^2}{1 + K'x^2} + (100 - \lambda) \frac{Kx}{1 + Kx} \dots\dots\dots(\text{A}),$$

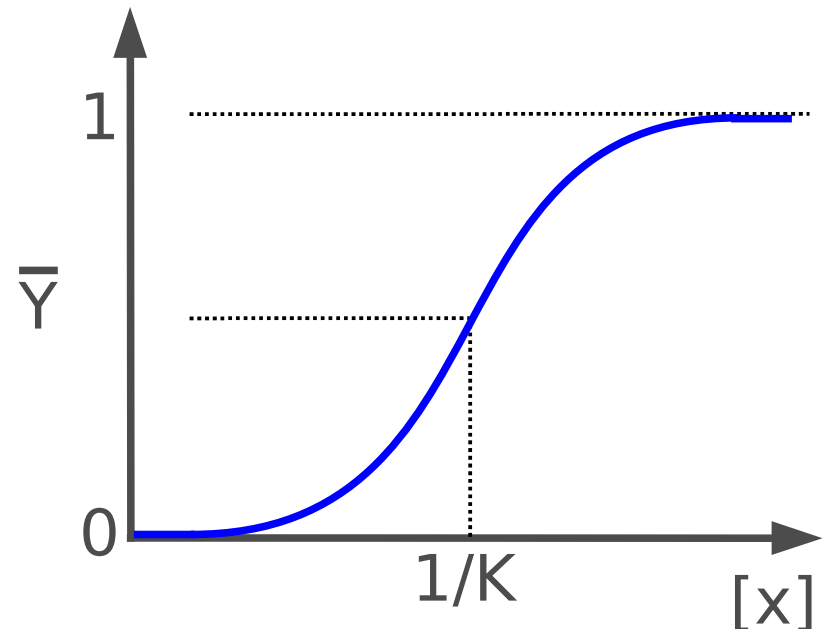
where  $\lambda\%$  is as  $\text{Hb}_2$ ,  $(100 - \lambda)\%$  as Hb,  $K'$  is the equilibrium constant of the reaction  $\text{Hb}_2 + 2\text{O}_2 \rightleftharpoons \text{Hb}_2\text{O}_4$  and  $K$  that of  $\text{Hb} + \text{O}_2 \rightleftharpoons \text{HbO}_2$ :  $K$  has the value .125 (Barcroft and Roberts).

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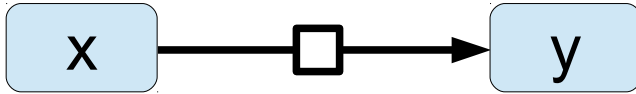
Now it is unlikely that in either of these cases there is only Hb and  $\text{Hb}_2$ : and as the calculation of the constants in these equations is very tedious I decided to try whether the equation

$$y = 100 \frac{Kx^n}{1 + Kx^n} \dots\dots\dots(\text{B})$$

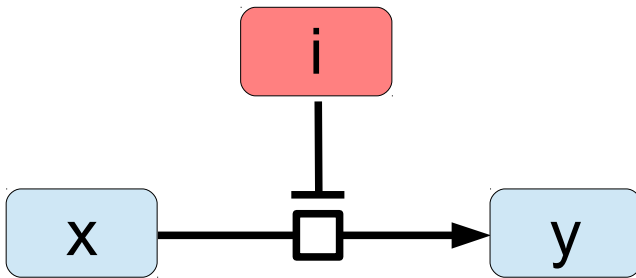
would satisfy the observations.



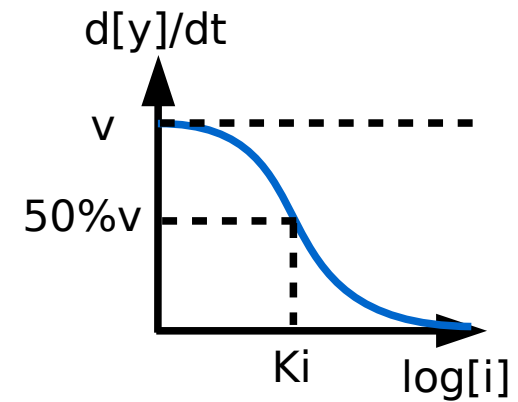
# Generalisation: inhibitors



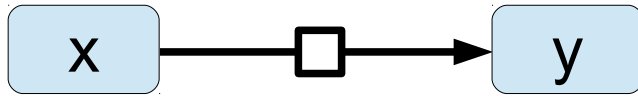
$$\frac{d[y]}{dt} = v (= k \cdot [x])$$



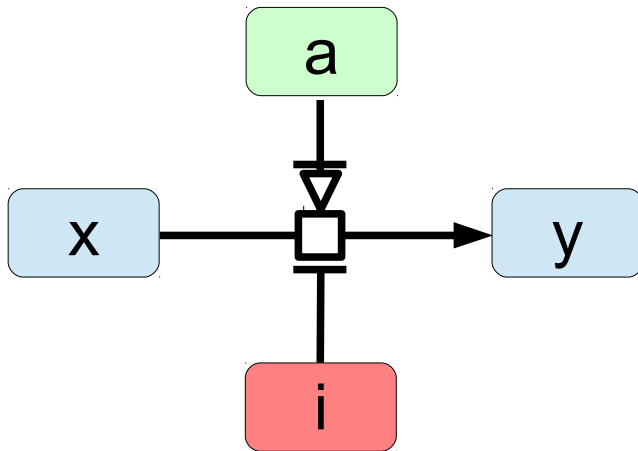
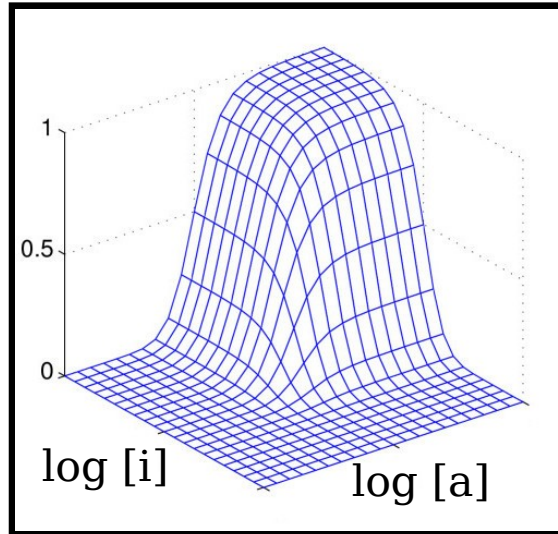
$$\frac{d[y]}{dt} = v \cdot \frac{K i^m}{K i^m + [i]^m}$$



# Generalisation: activators and inhibitors

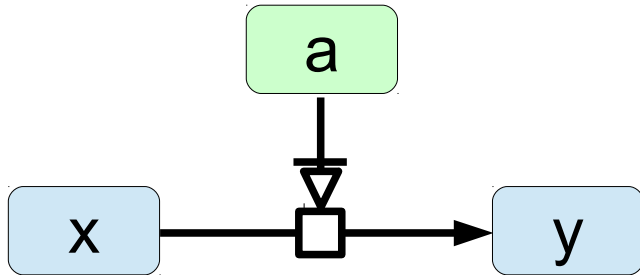


$$\frac{d[y]}{dt} = v(= k \cdot [x])$$

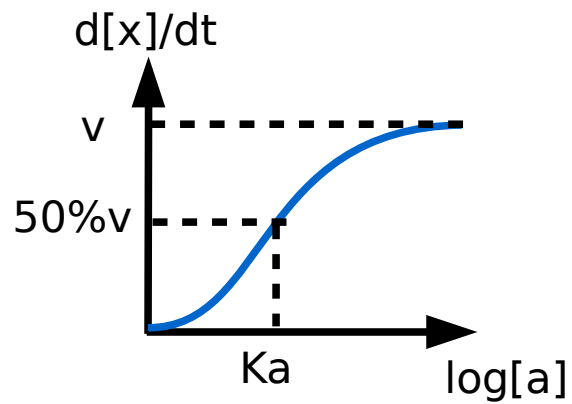


$$\frac{d[y]}{dt} = v \cdot \frac{[a]^n}{K a^n + [a]^n} \cdot \frac{K i^m}{K i^m + [i]^m}$$

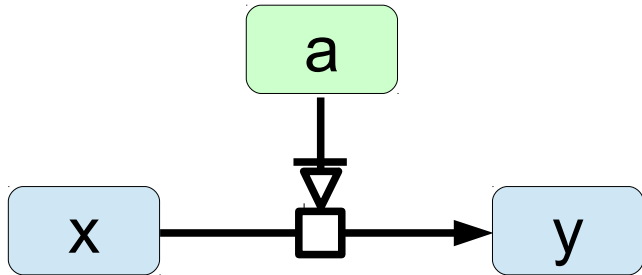
# absolute Vs relative activators



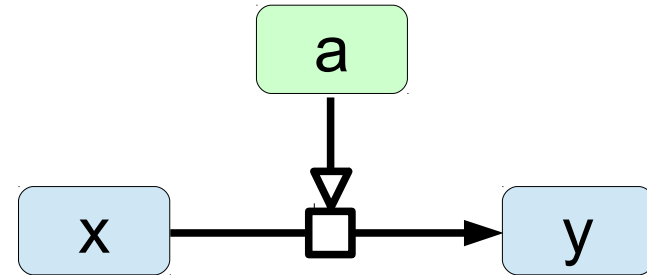
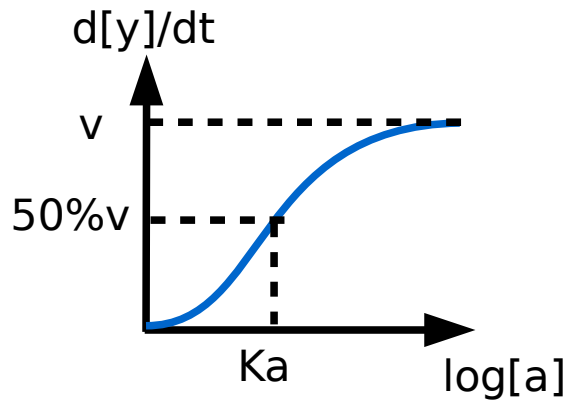
$$\frac{d[y]}{dt} = v \cdot \frac{[a]}{K_a + [a]}$$



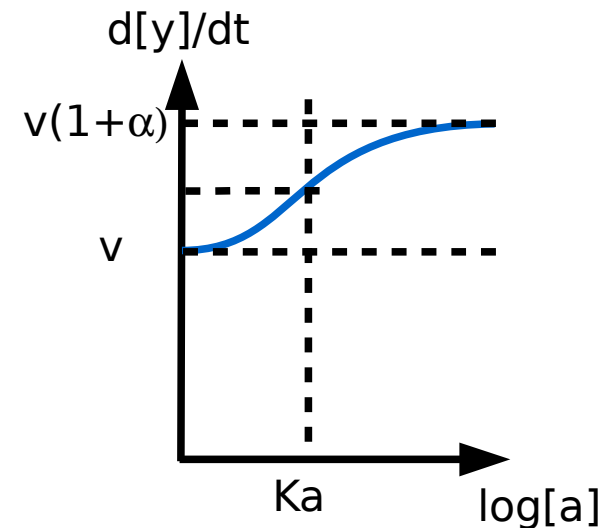
# absolute Vs relative activators



$$\frac{d[y]}{dt} = v \cdot \frac{[a]}{K_a + [a]}$$

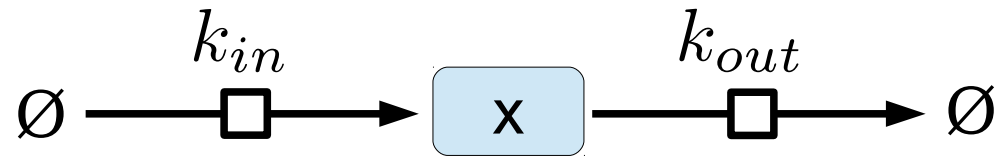


$$\frac{d[y]}{dt} = v \cdot \left(1 + \alpha \cdot \frac{[a]}{K_a + [a]}\right)$$



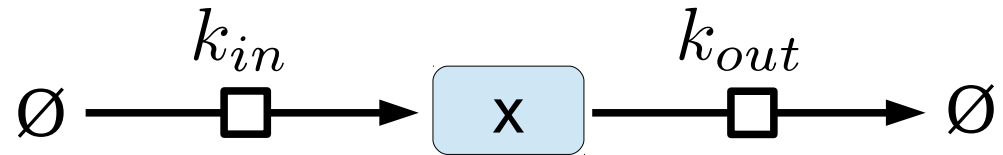
# Homeostasis

How can we maintain a stable level with a dynamic system?



# Homeostasis

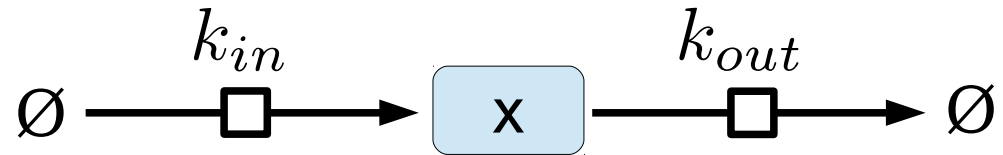
How can we maintain a stable level with a dynamic system?



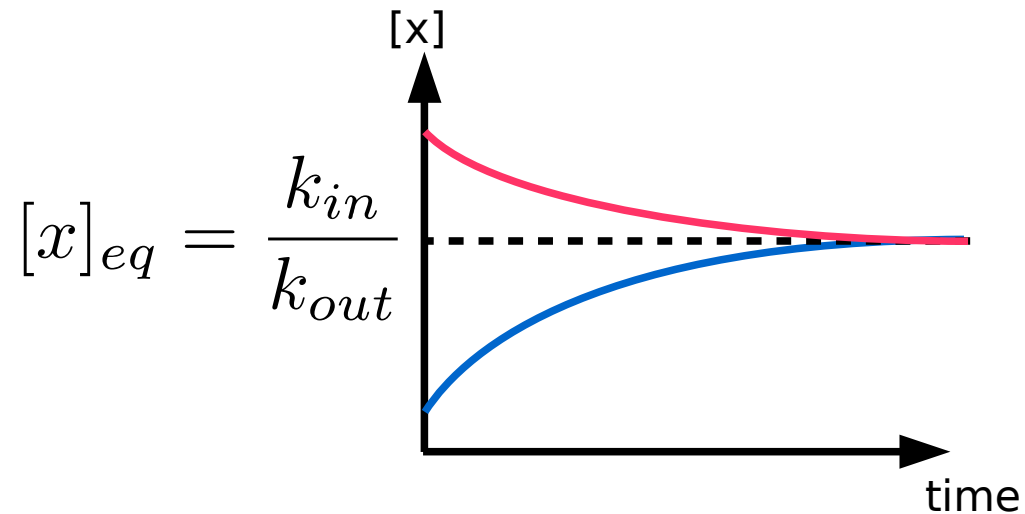
$$\frac{d[x]}{dt} = k_{in} - k_{out} \cdot [x]$$

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How can we maintain a stable level with a dynamic system?

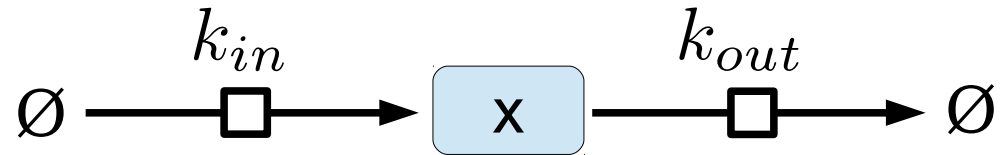


$$\frac{d[x]}{dt} = k_{in} - k_{out} \cdot [x]$$

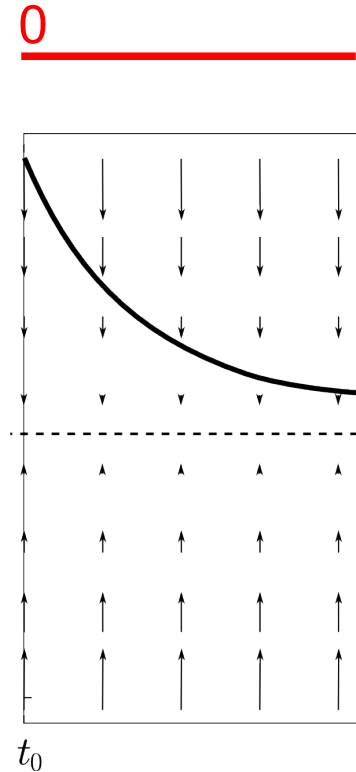
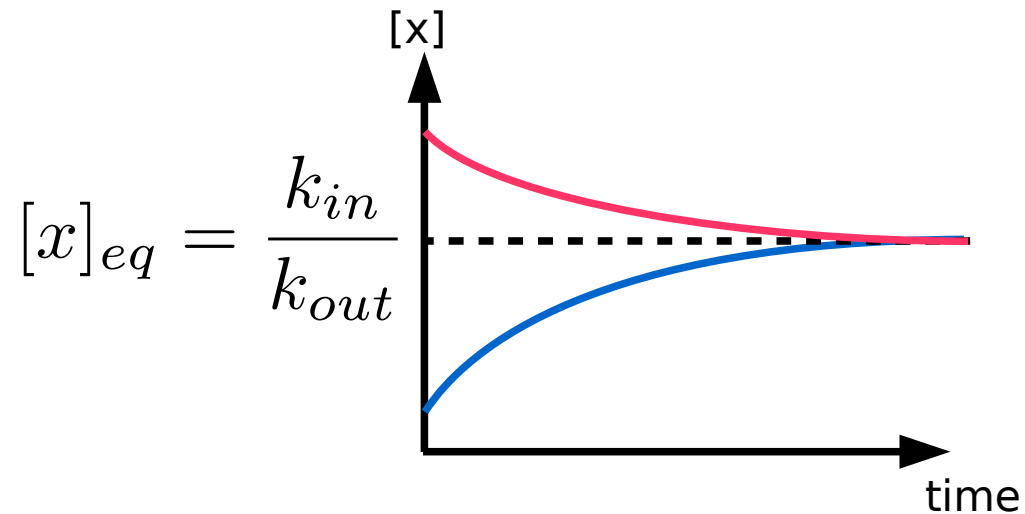


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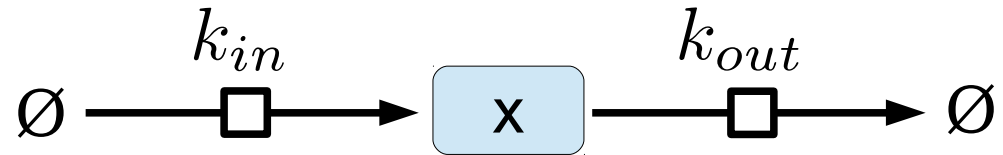


$$\frac{d[x]}{dt} = k_{in} - k_{out} \cdot [x]$$

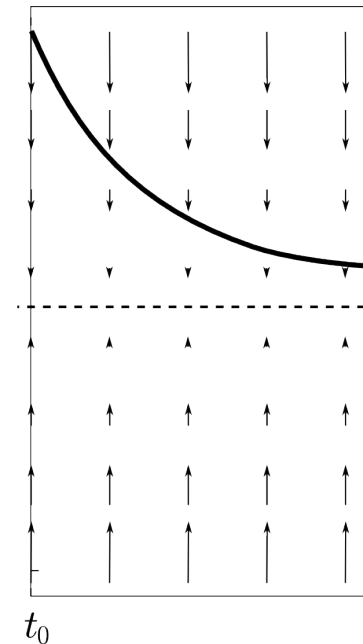
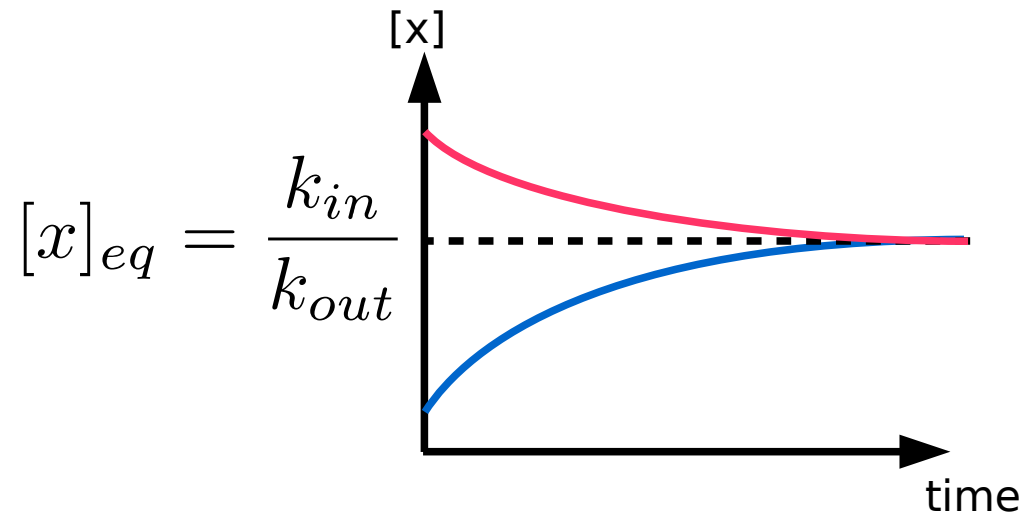
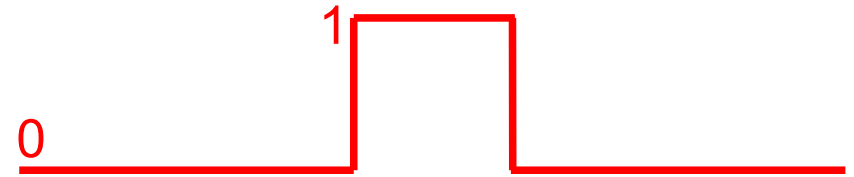


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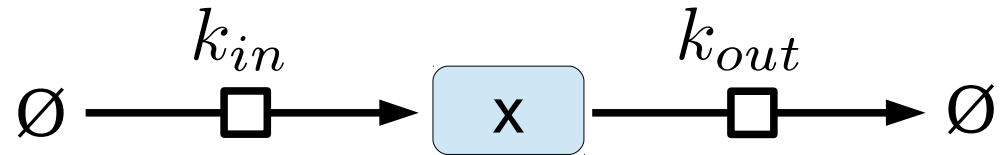


$$\frac{d[x]}{dt} = k_{in} - k_{out} \cdot [x] - k_{pert} \cdot [x]$$

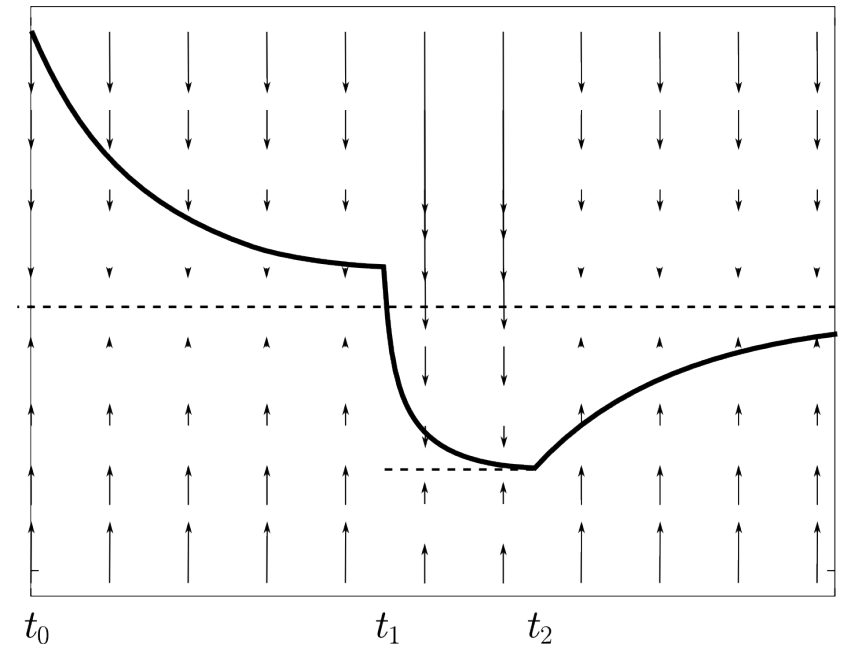
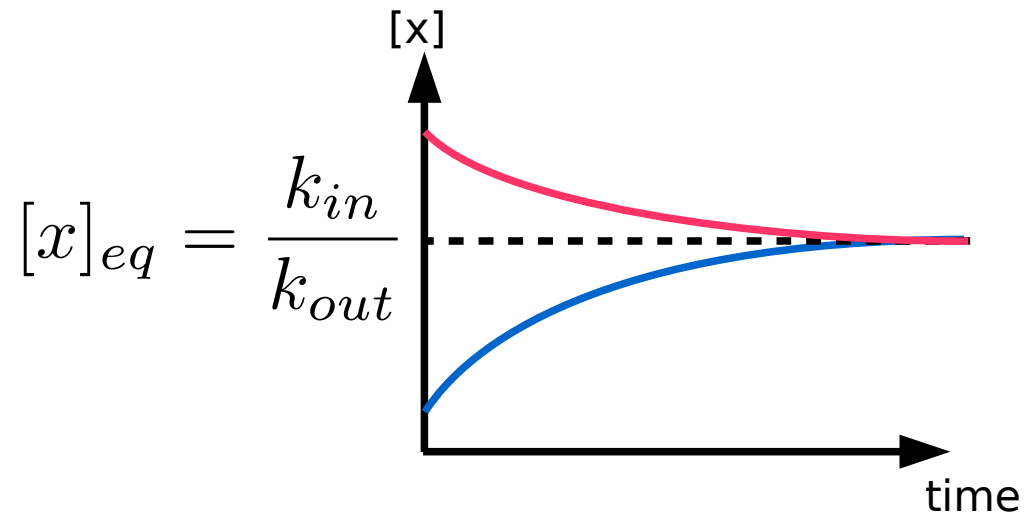
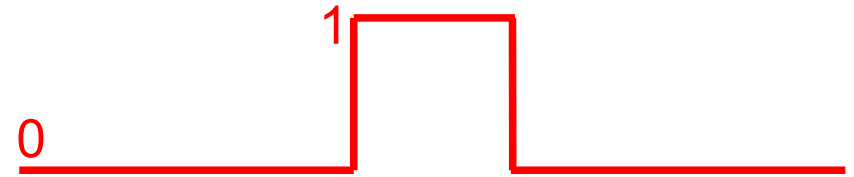


# Homeostasis

How can we maintain a stable level with a dynamic system?



$$\frac{d[x]}{dt} = k_{in} - k_{out} \cdot [x] - k_{pert} \cdot [x]$$



Questions?