

Multi-scale modelling of neuronal signalling

Nicolas Le Novère, EMBL-EBI

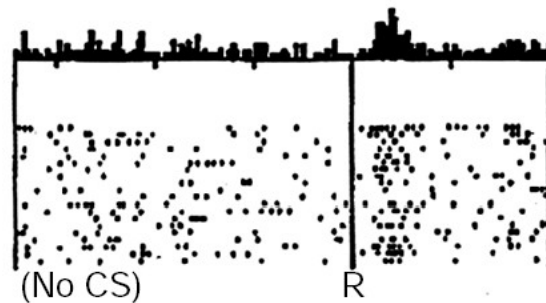


Modelling of neuronal signalling at multiple scales

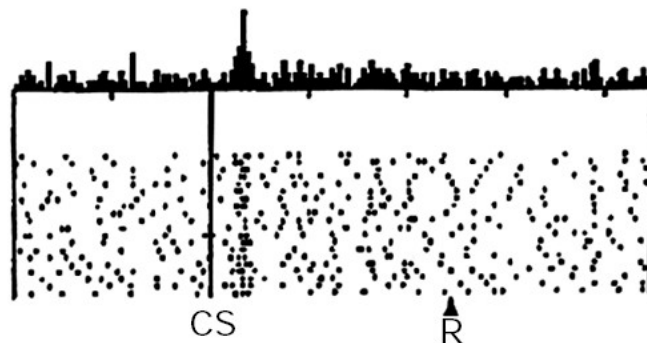
Nicolas Le Novère, EMBL-EBI



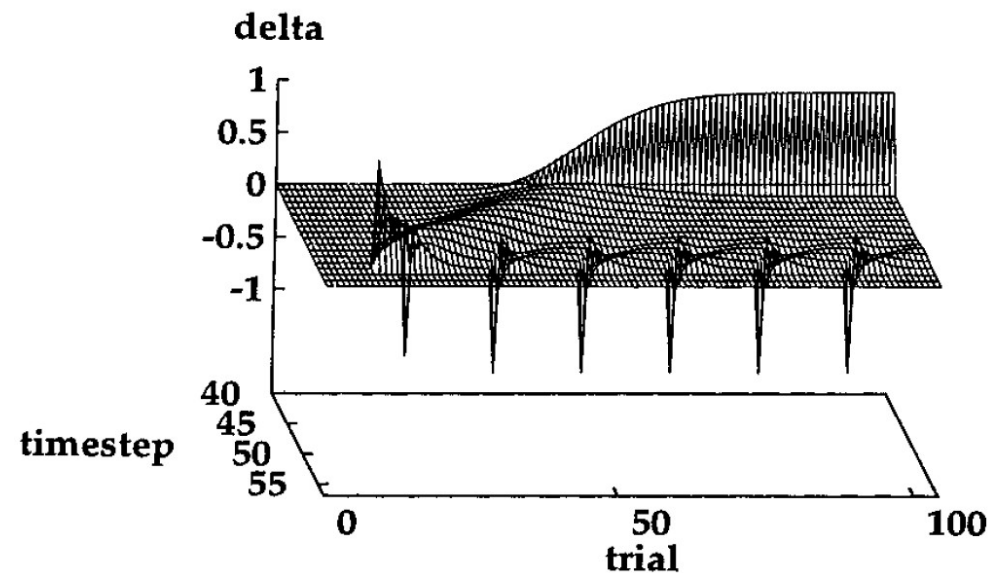
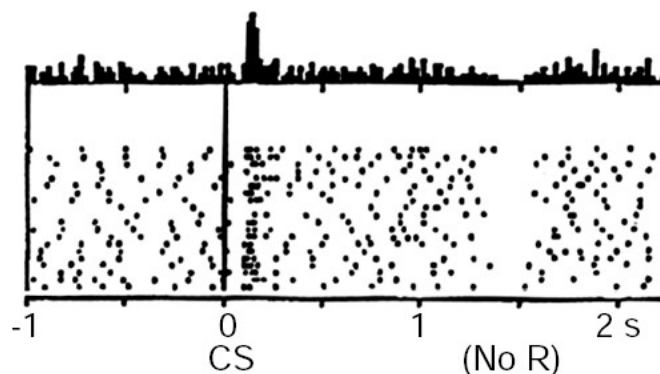
No prediction
Reward occurs



Reward predicted
Reward occurs



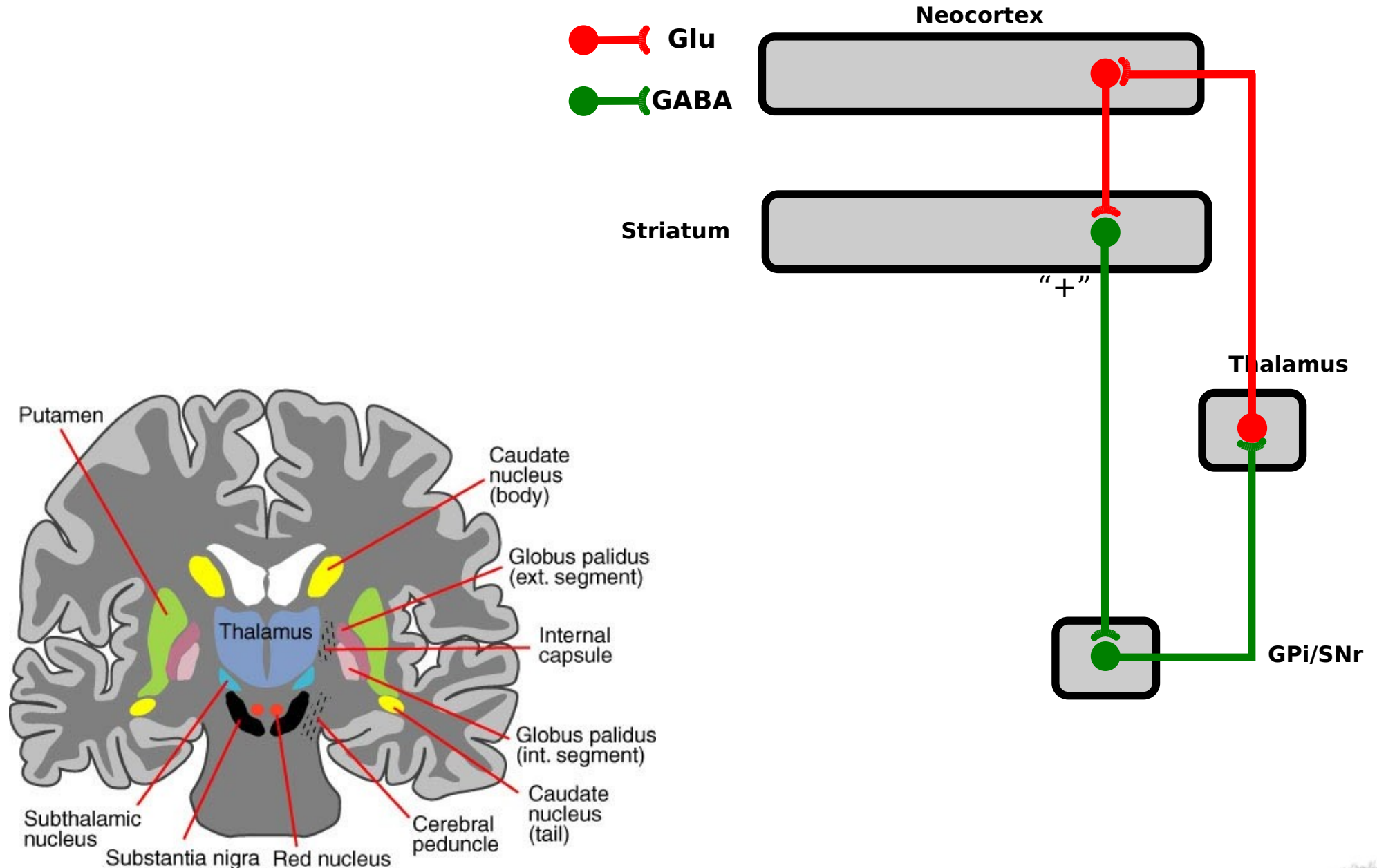
Reward predicted
No reward occurs

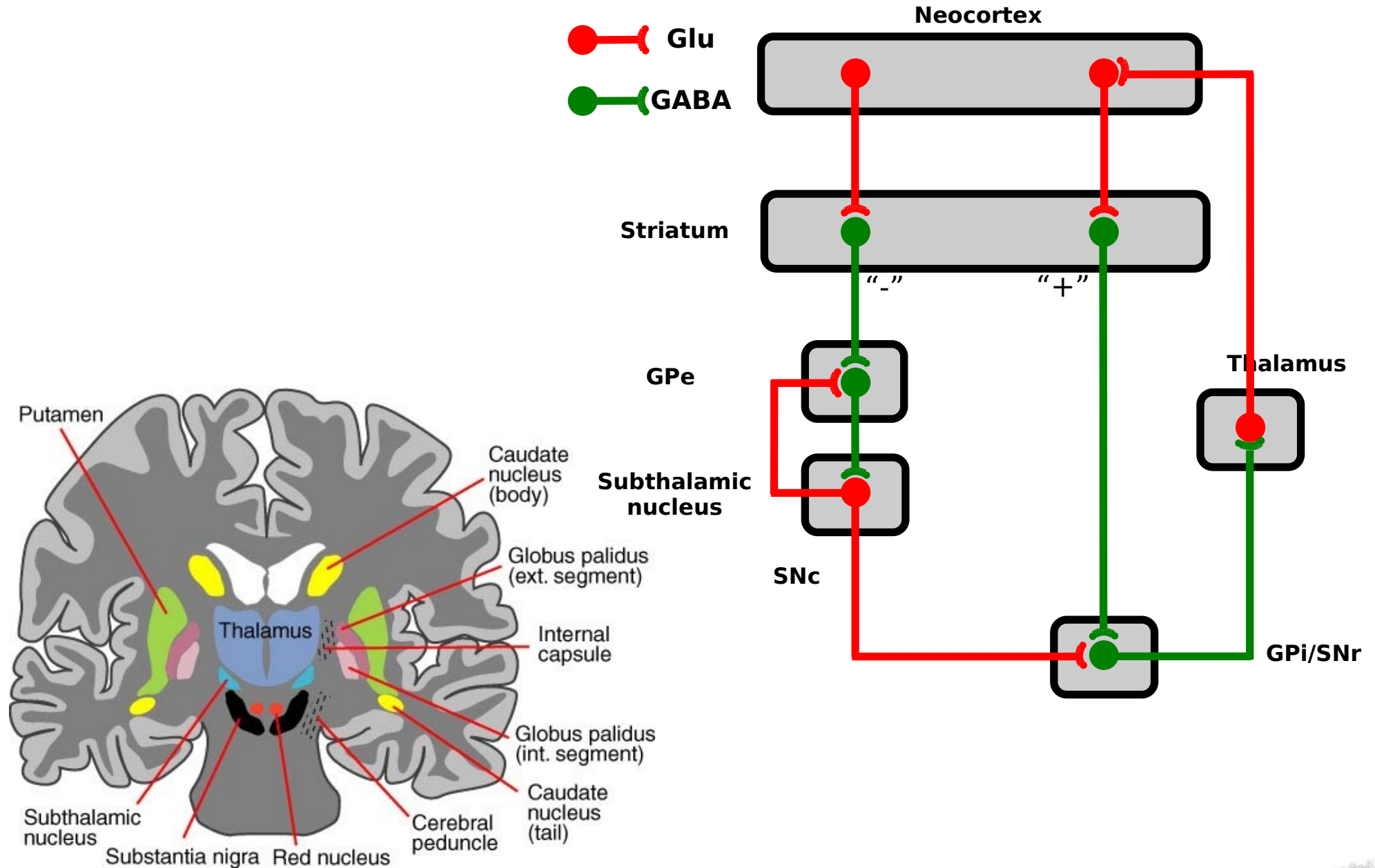


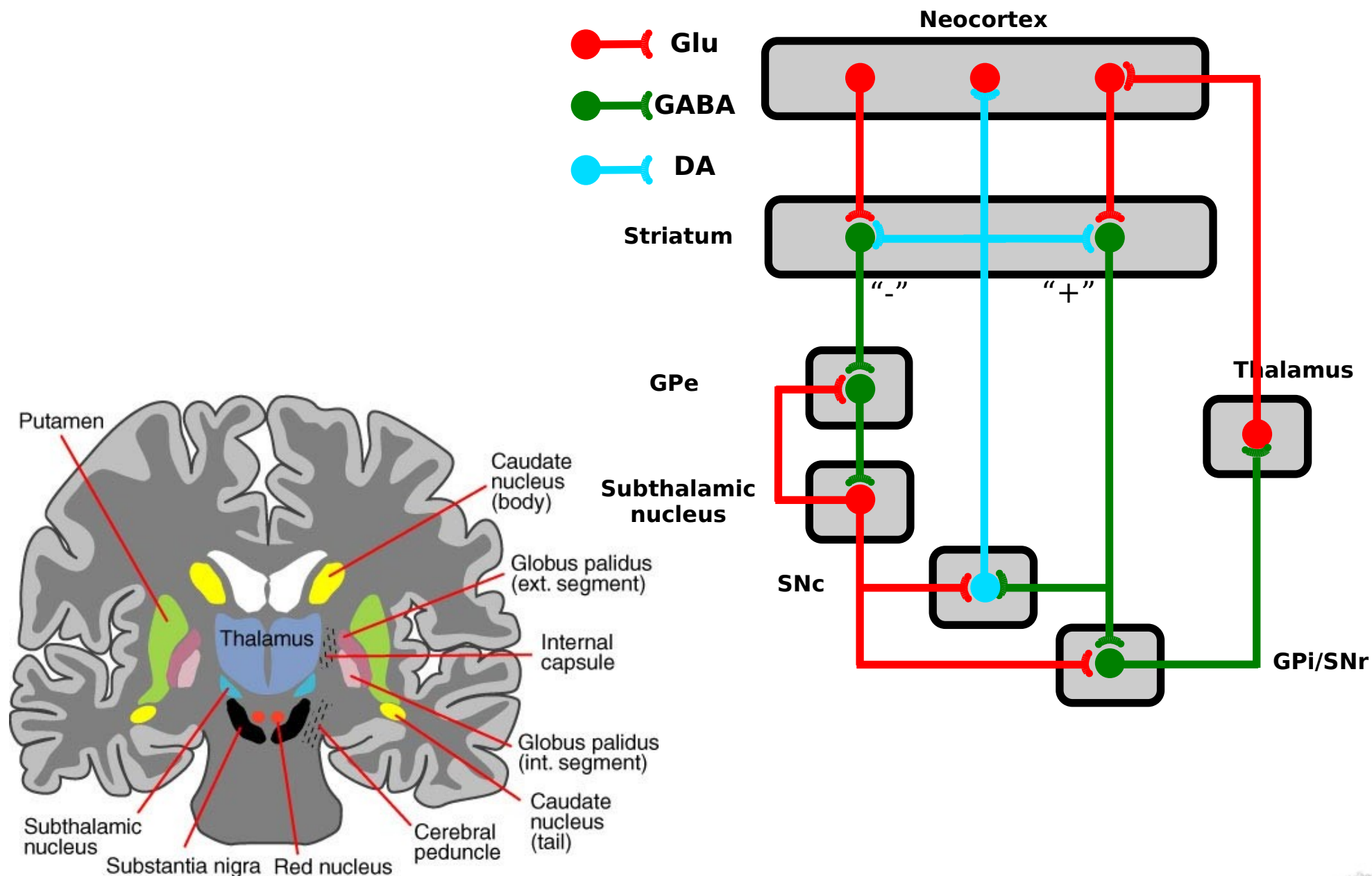
Schultz W, Dayan P, Montague PR (1997)
A neural substrate of prediction and reward.
Science 275:1593-1599.

Montague PR, Dayan P, Sejnowski TJ (1996)
A framework for mesencephalic dopamine
systems based on predictive Hebbian learning.
J Neurosci 16: 1936-1947

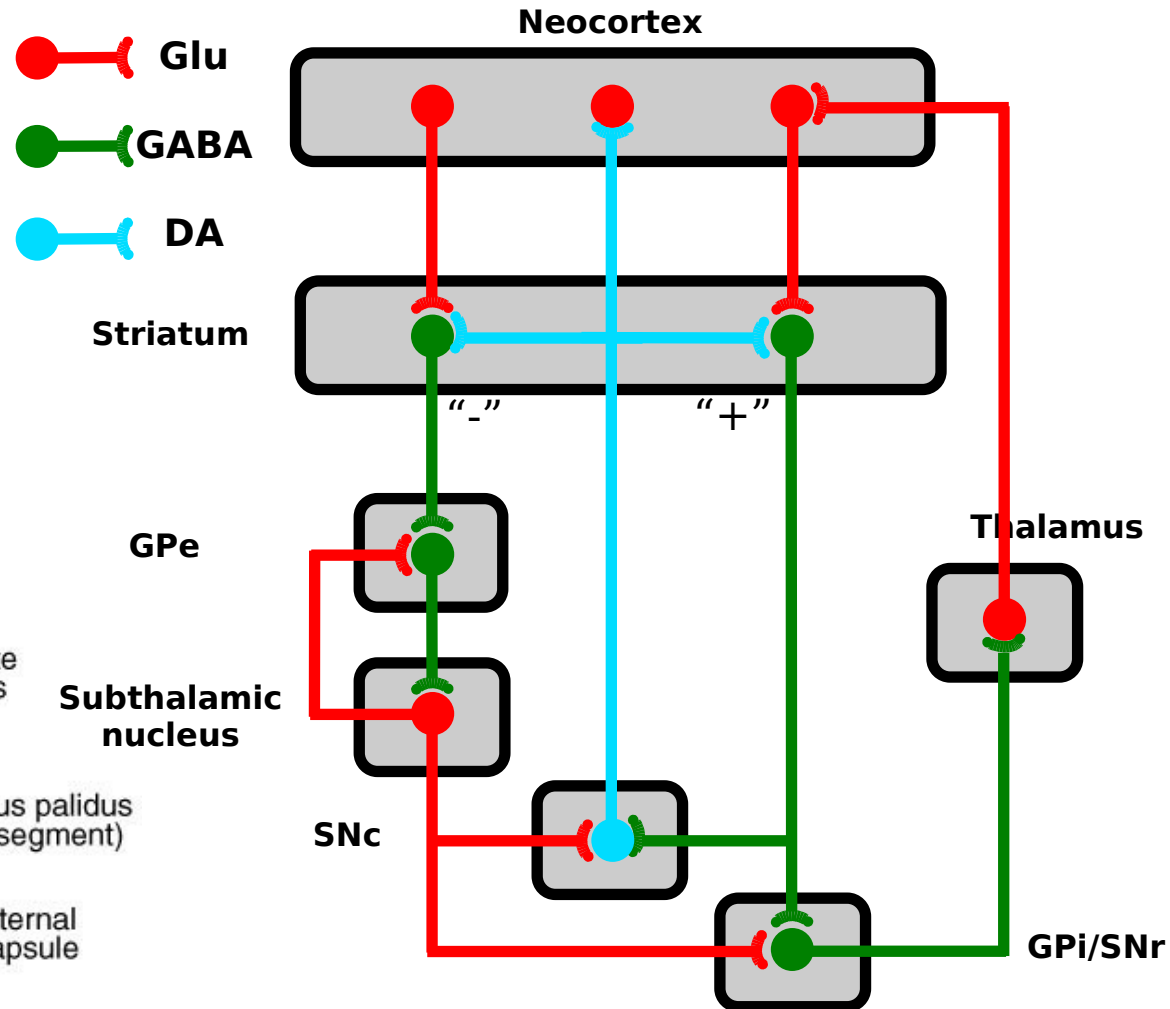
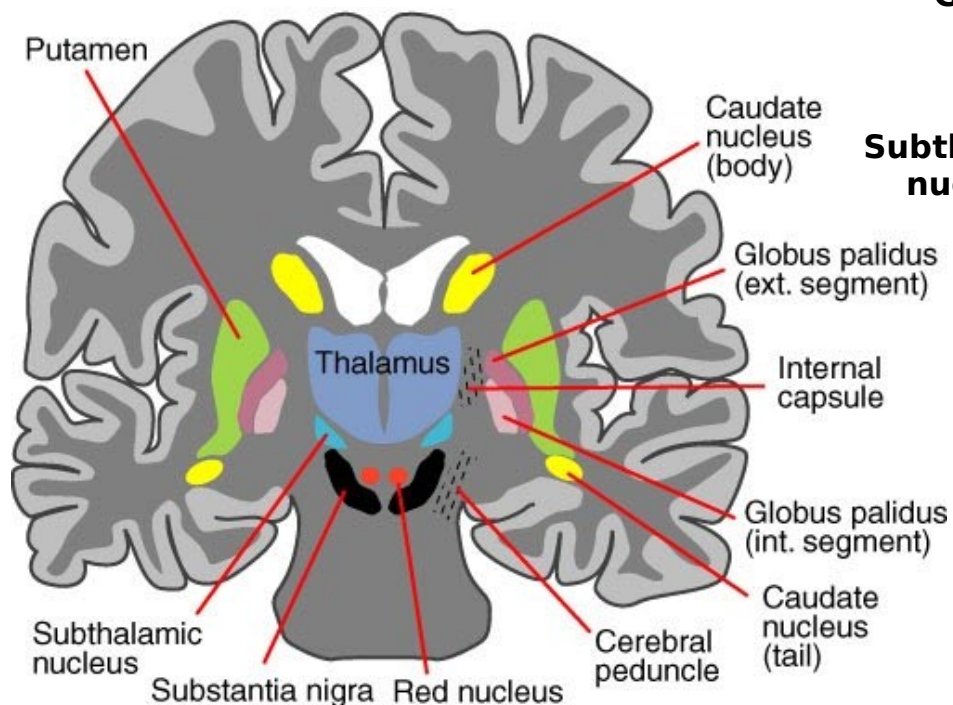






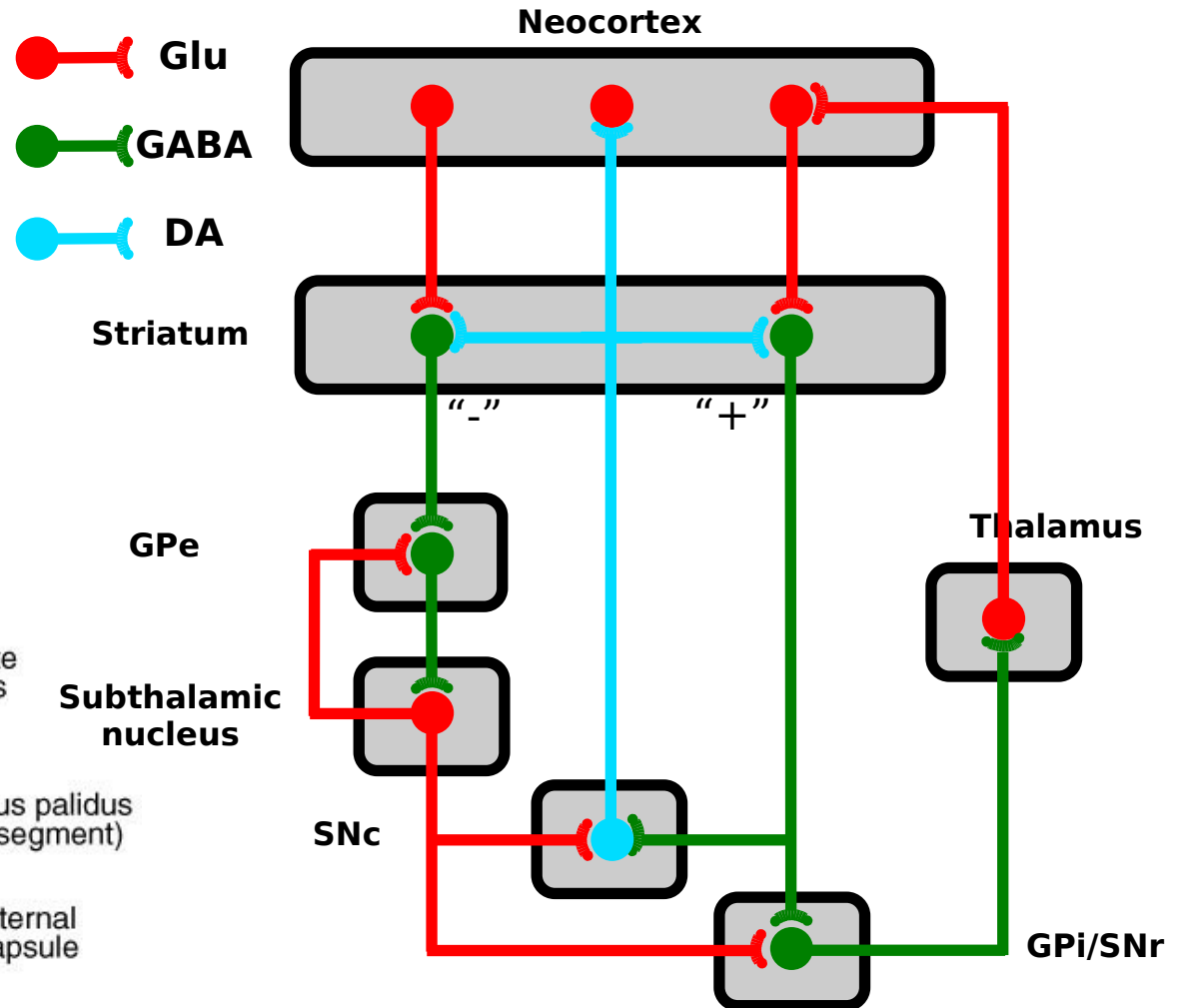
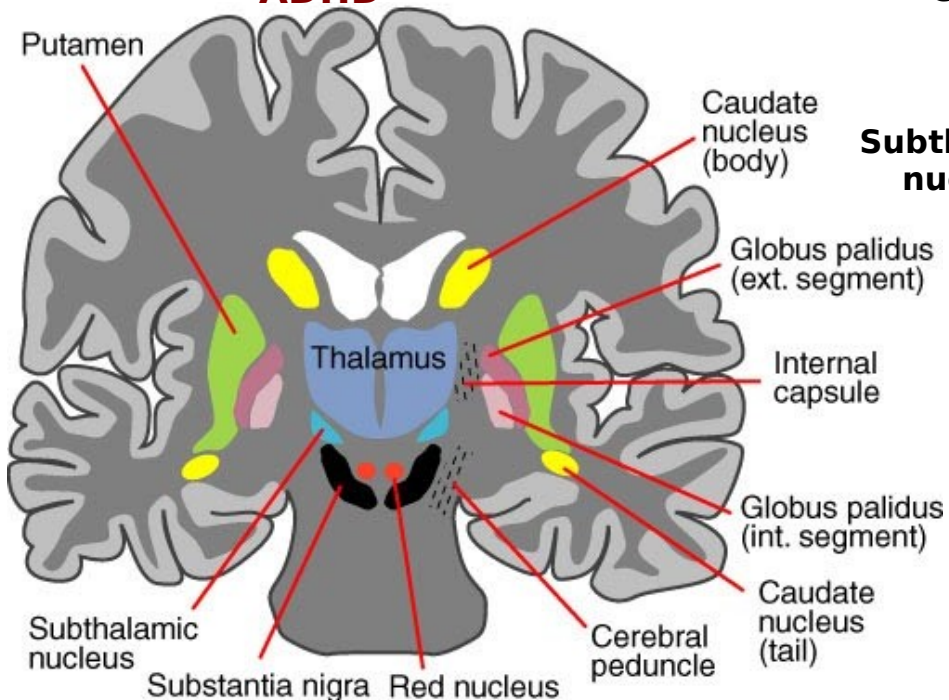


Voluntary motion
Emotional control
Reward
Learning



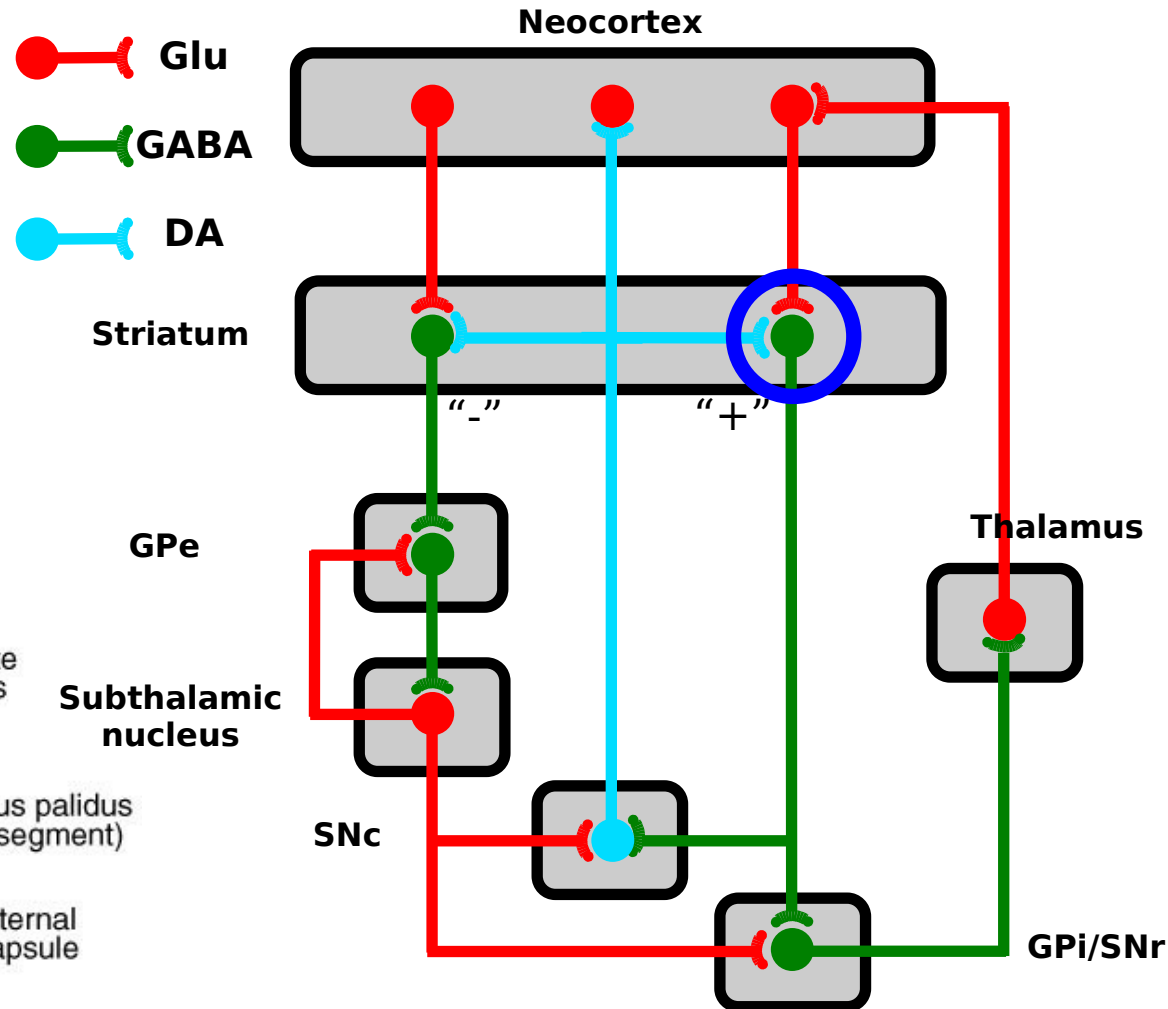
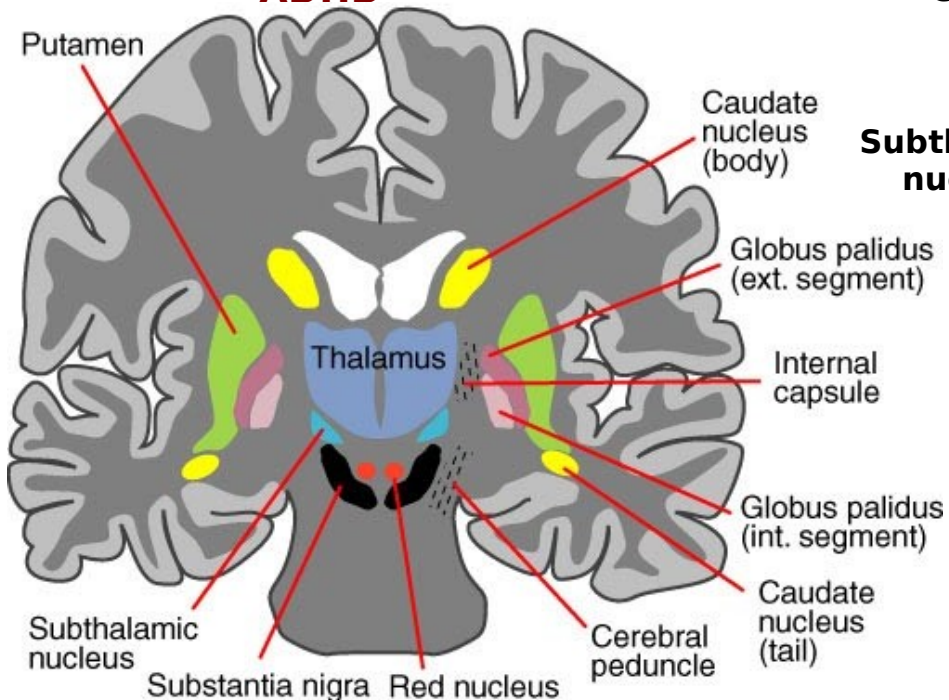
Voluntary motion
Emotional control
Reward
Learning

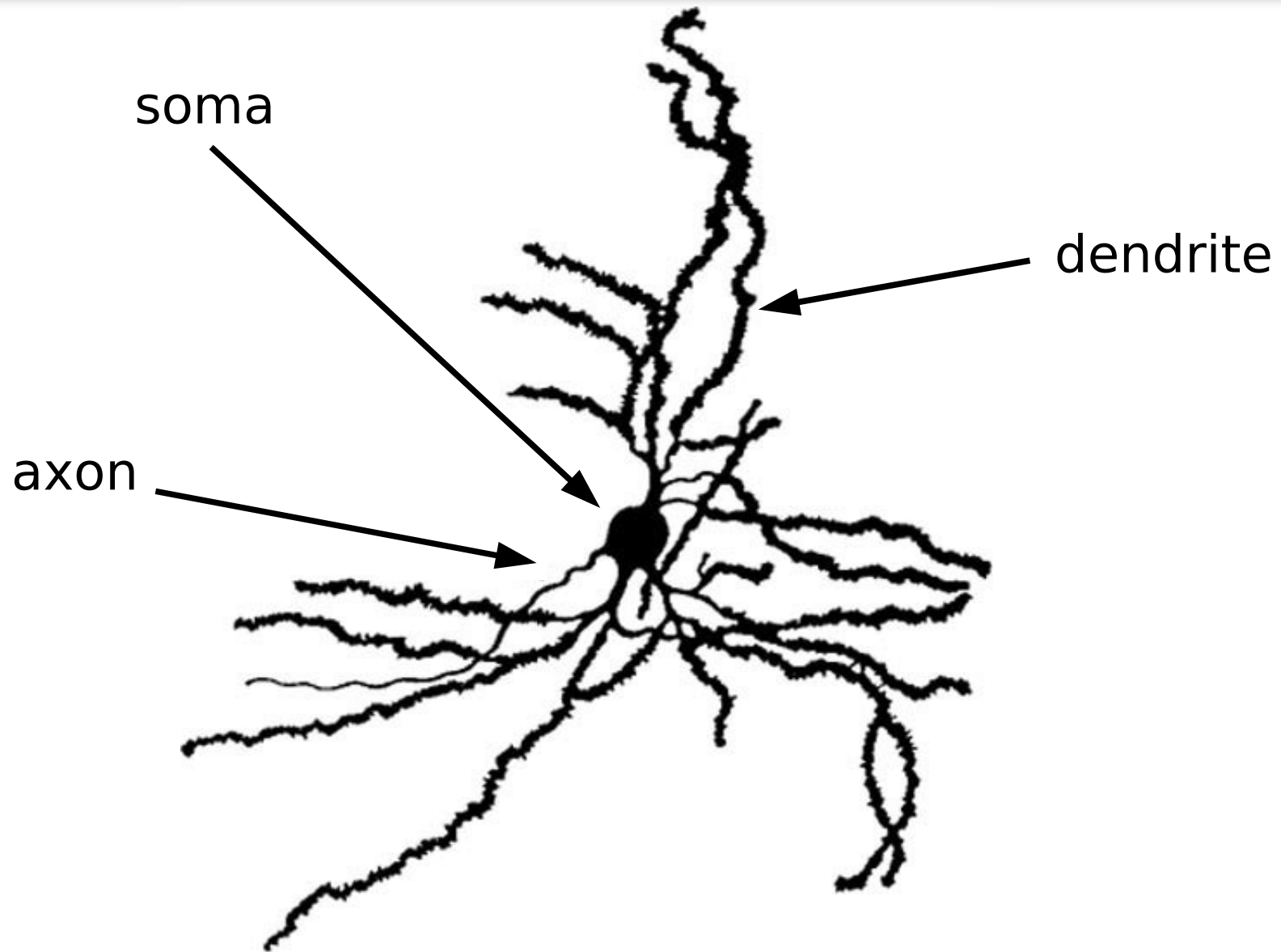
Parkinson's disease
Huntington's chorea
Hemibalism
Schizophrenia
Drug addiction
Depression
Obsessive-Compulsive Disorder
Tourette syndrome
ADHD



Voluntary motion
Emotional control
Reward
Learning

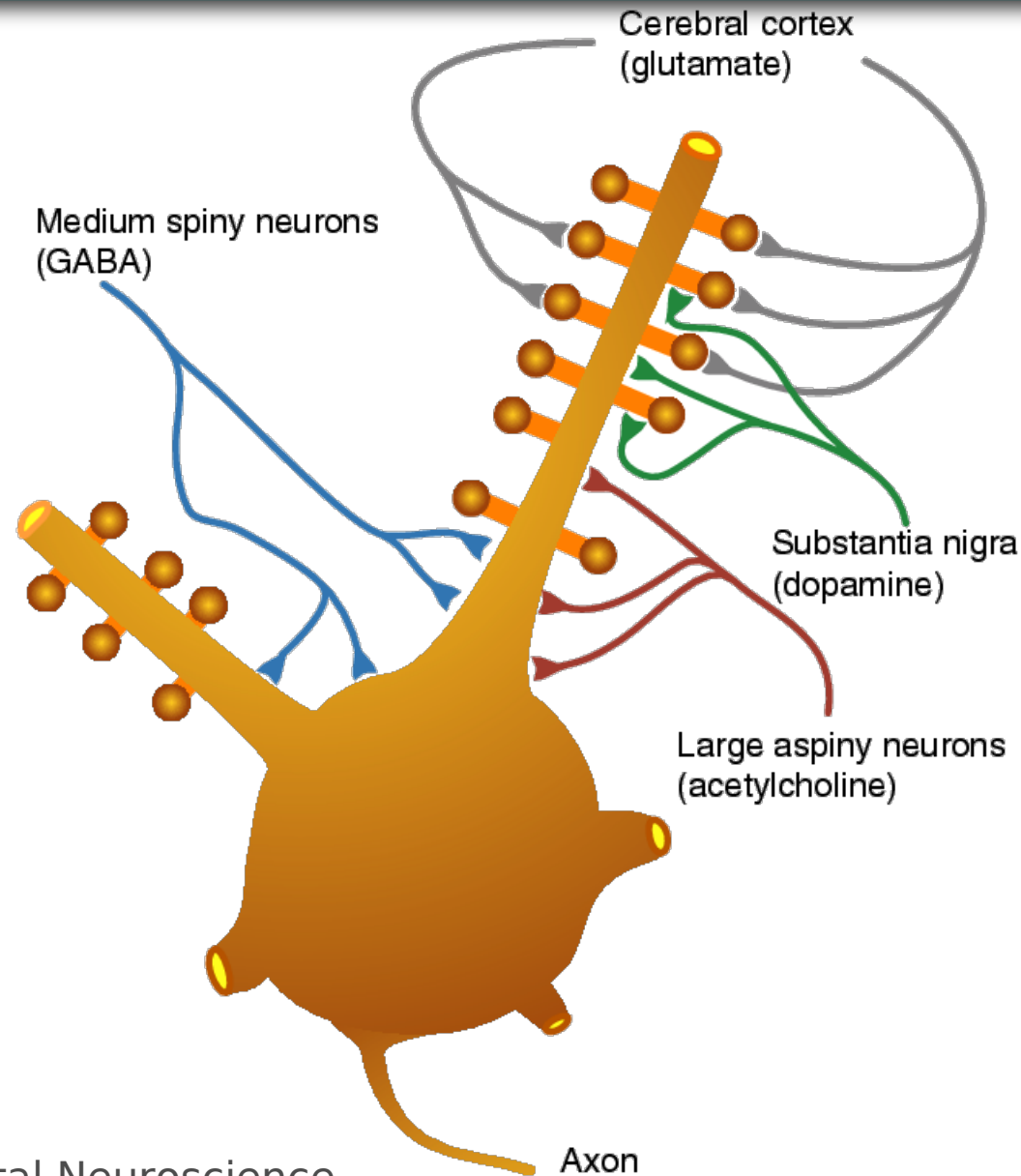
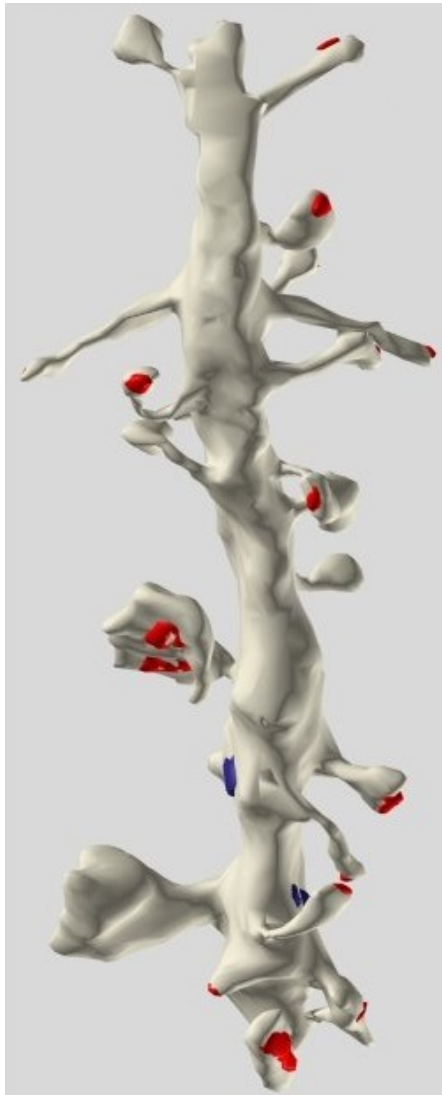
Parkinson's disease
Huntington's chorea
Hemibalism
Schizophrenia
Drug addiction
Depression
Obsessive-Compulsive
Disorder
Tourette syndrome
ADHD





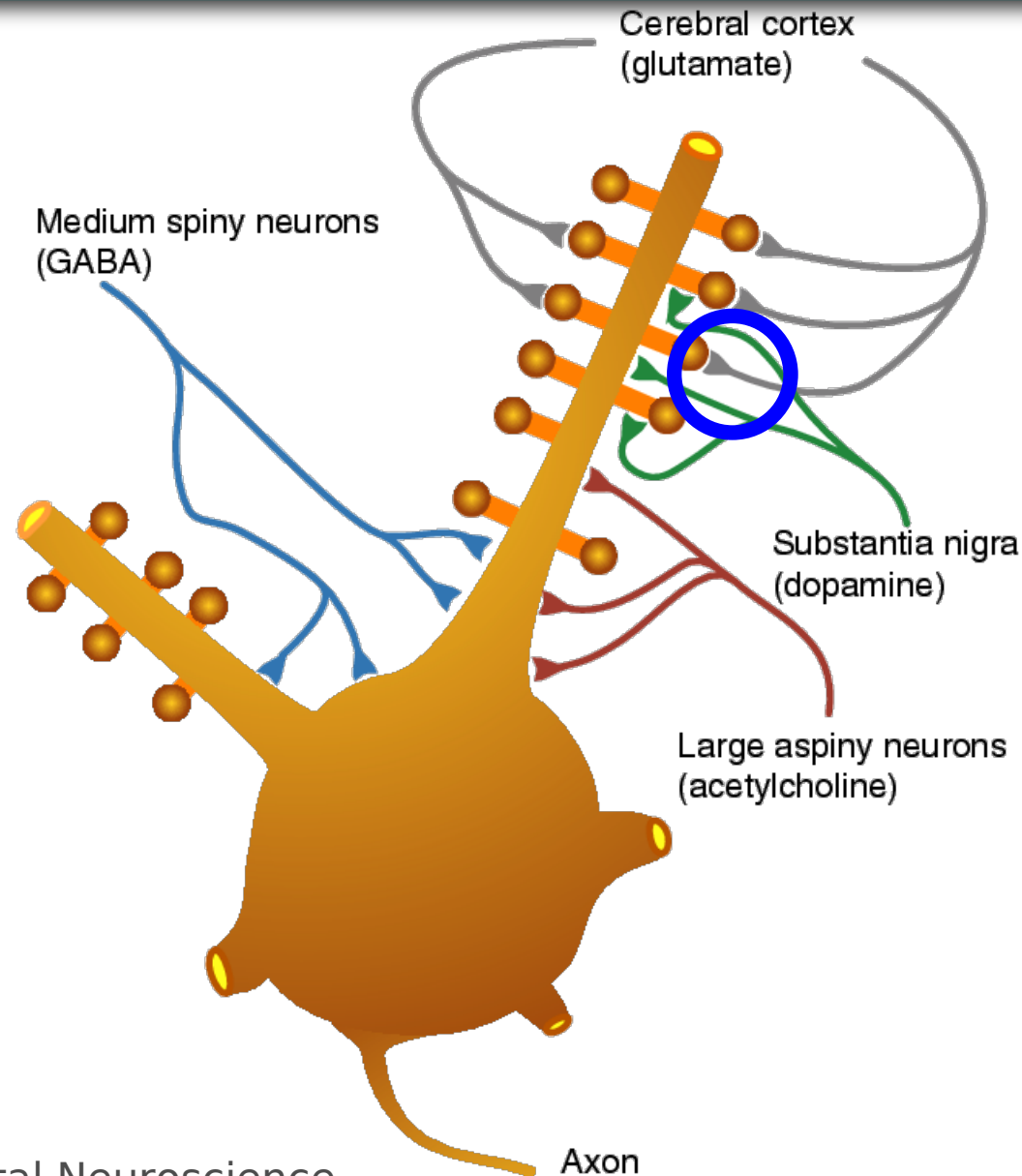
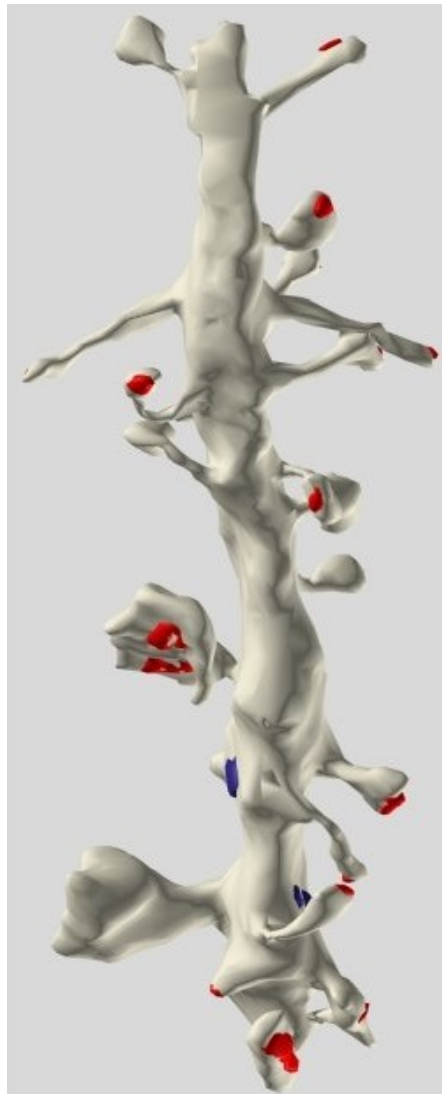
Copyright © 2002, Elsevier Science (USA). All rights reserved.





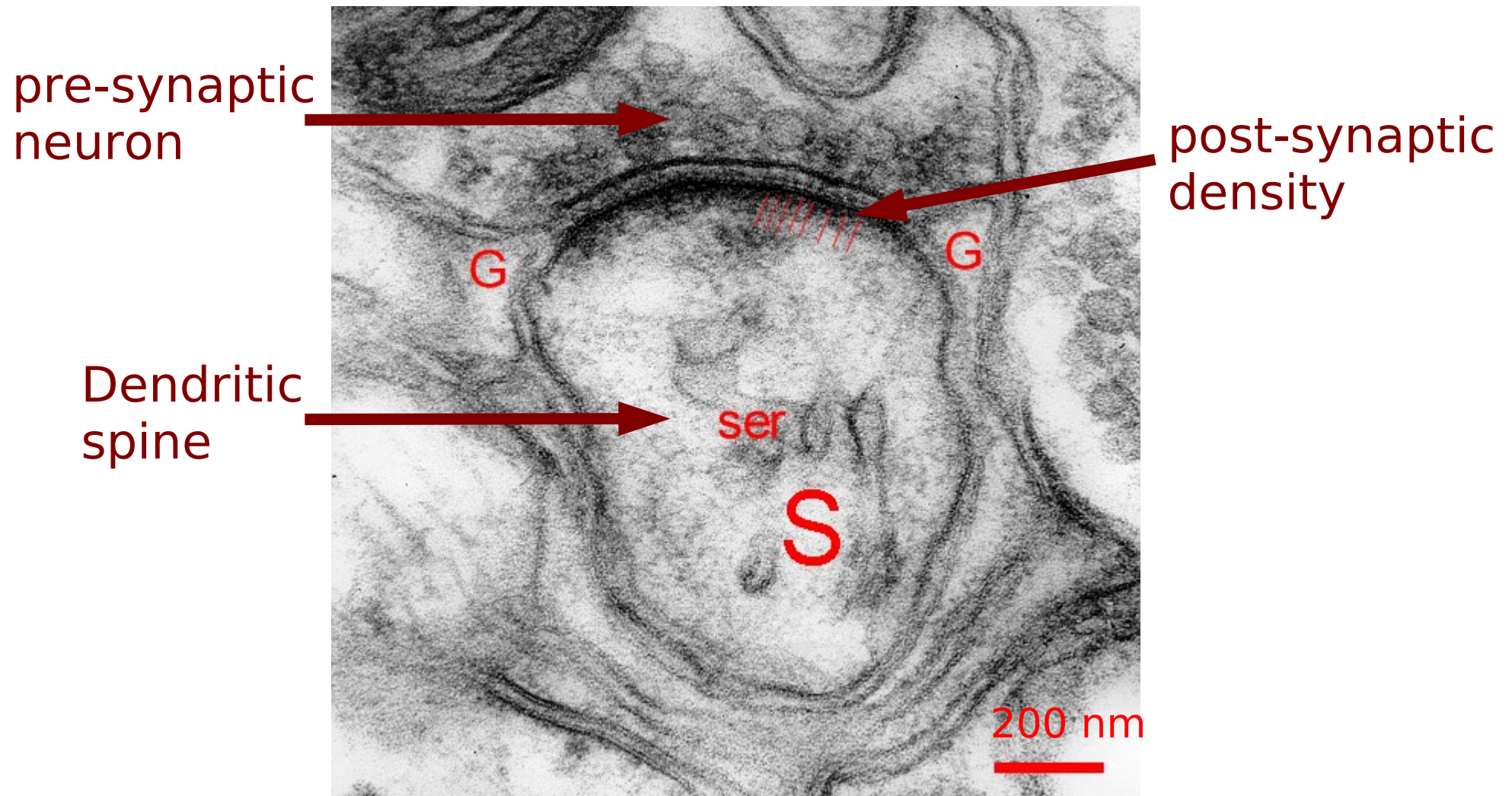
Fundamental Neuroscience
Squire et al. 2nd ed(2003)

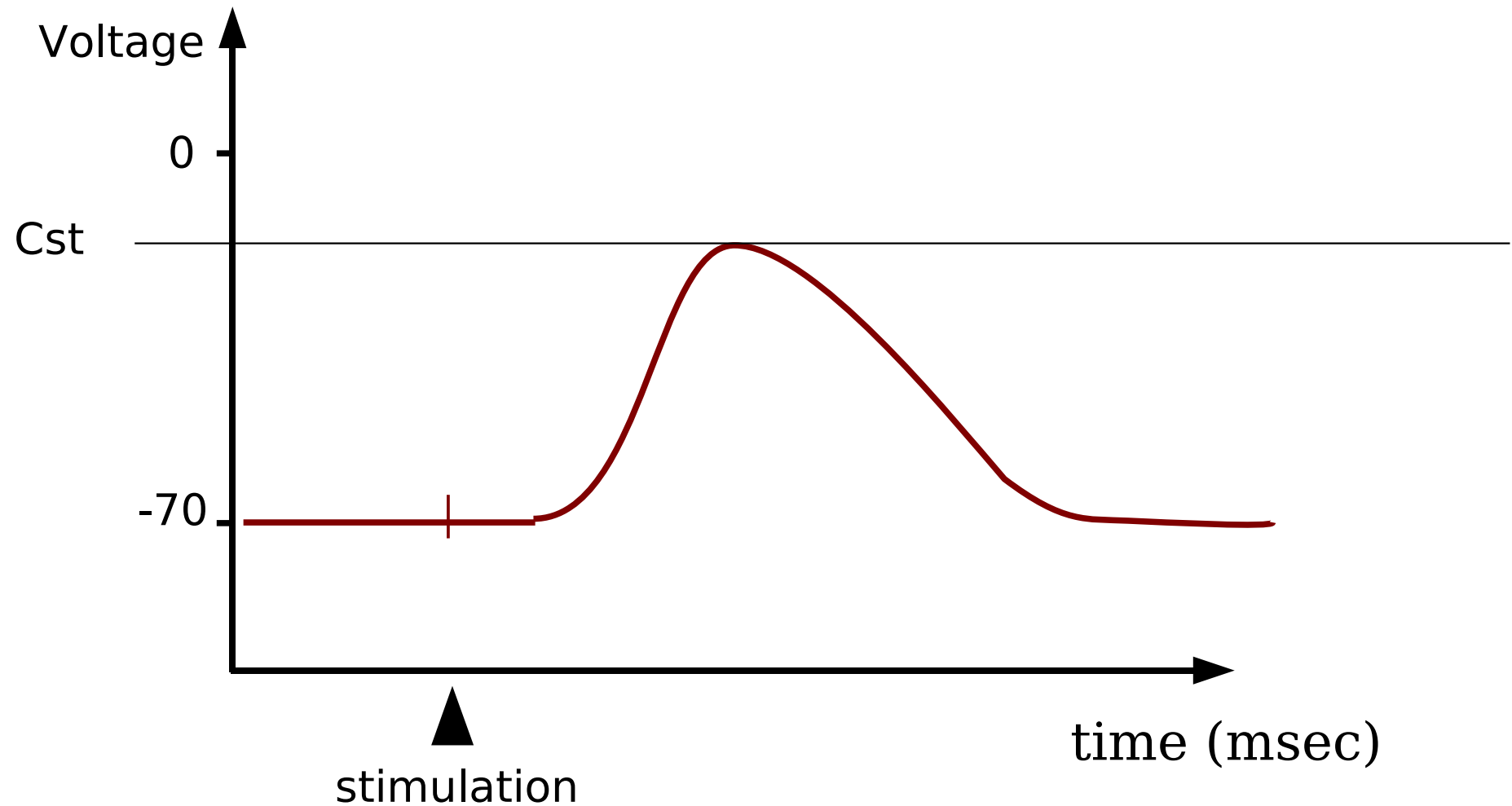


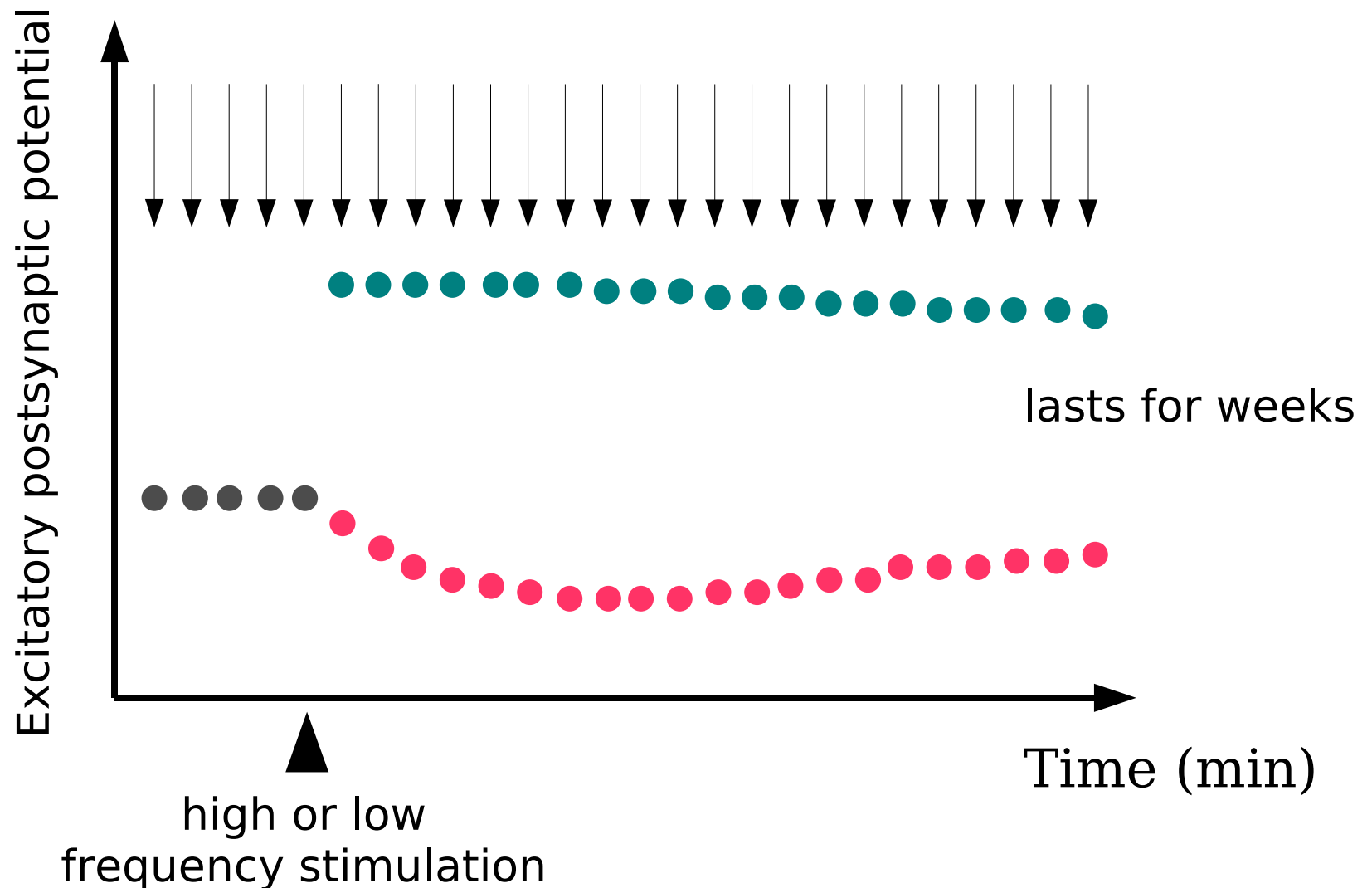


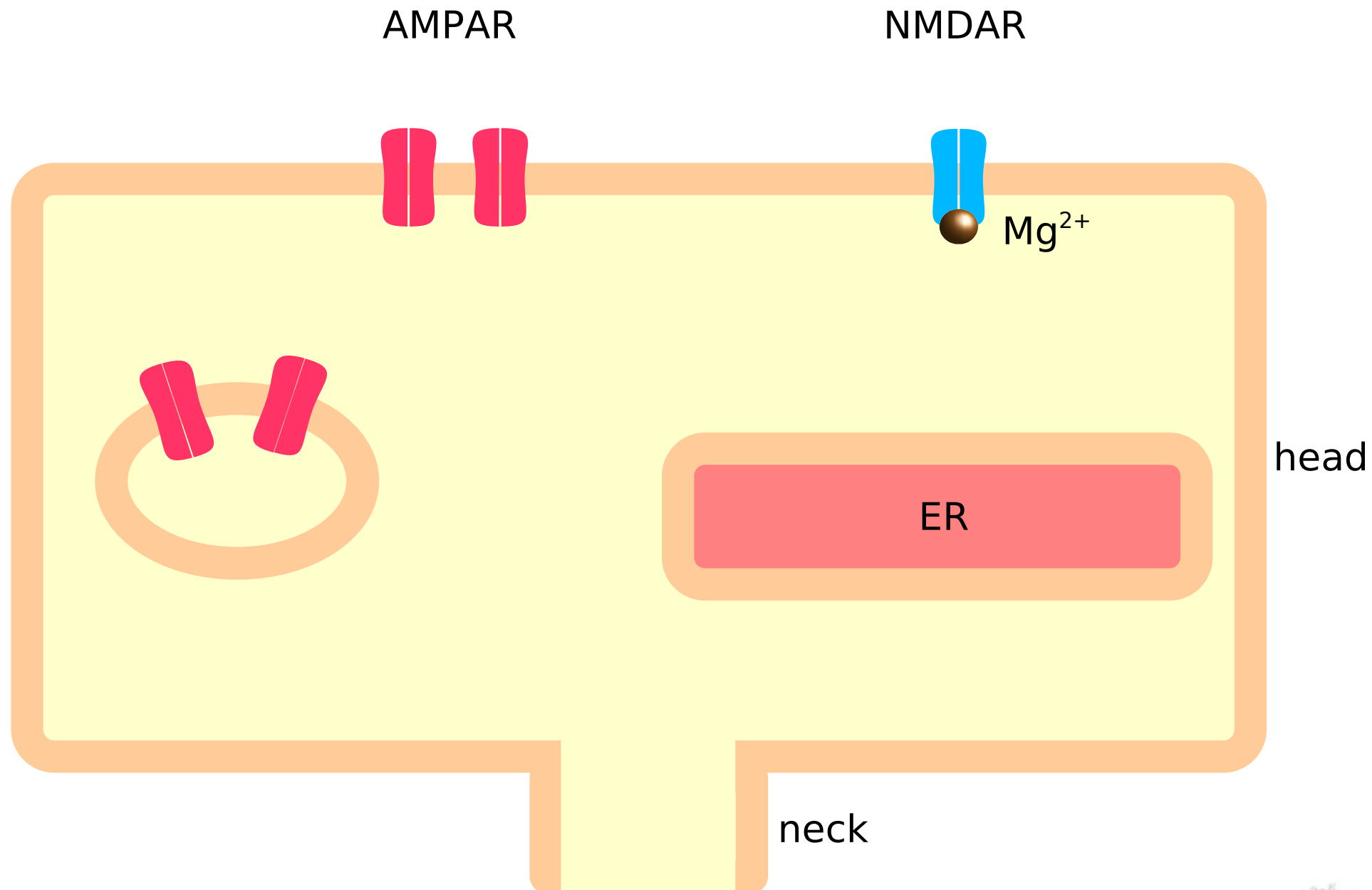
Fundamental Neuroscience
Squire et al. 2nd ed(2003)



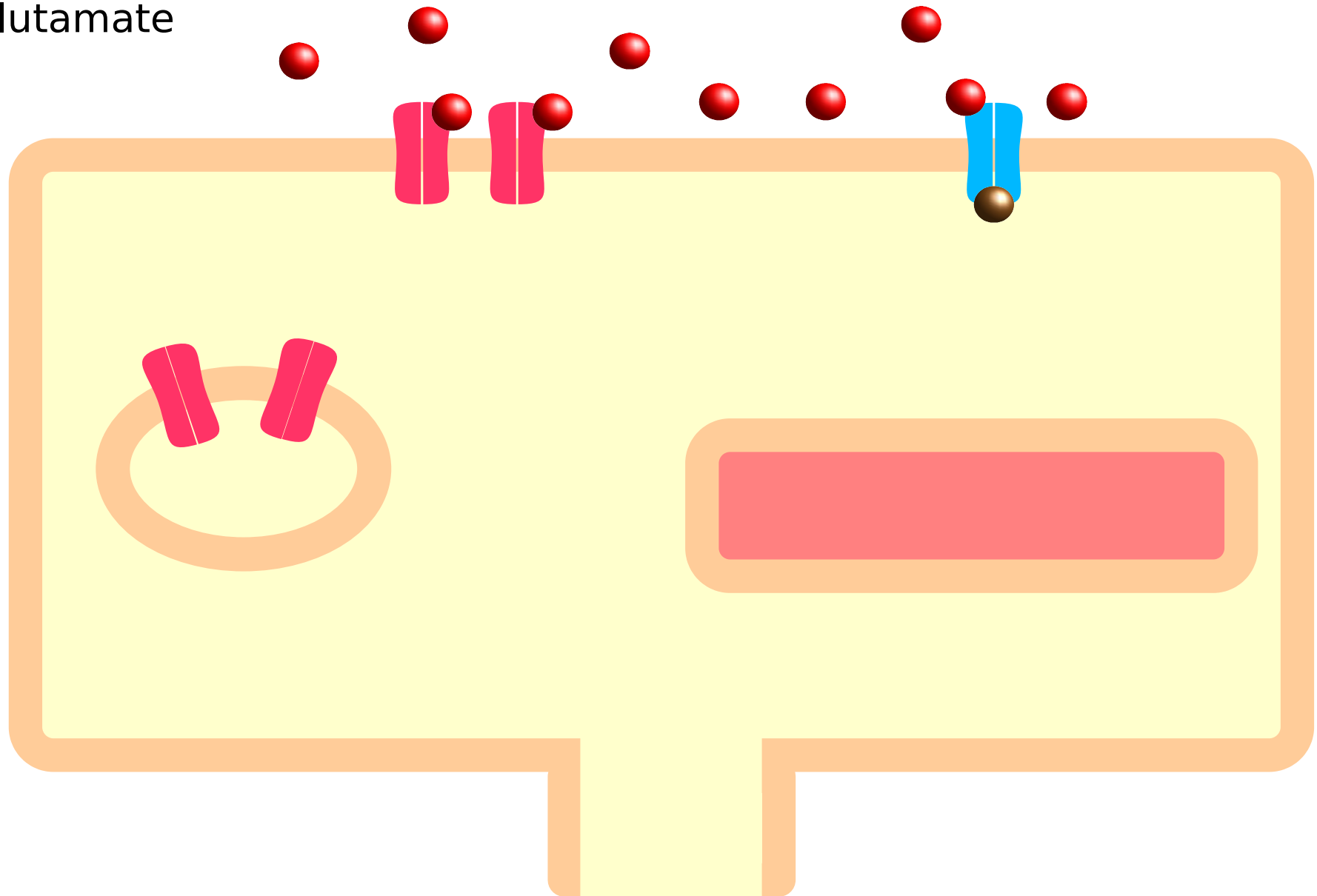


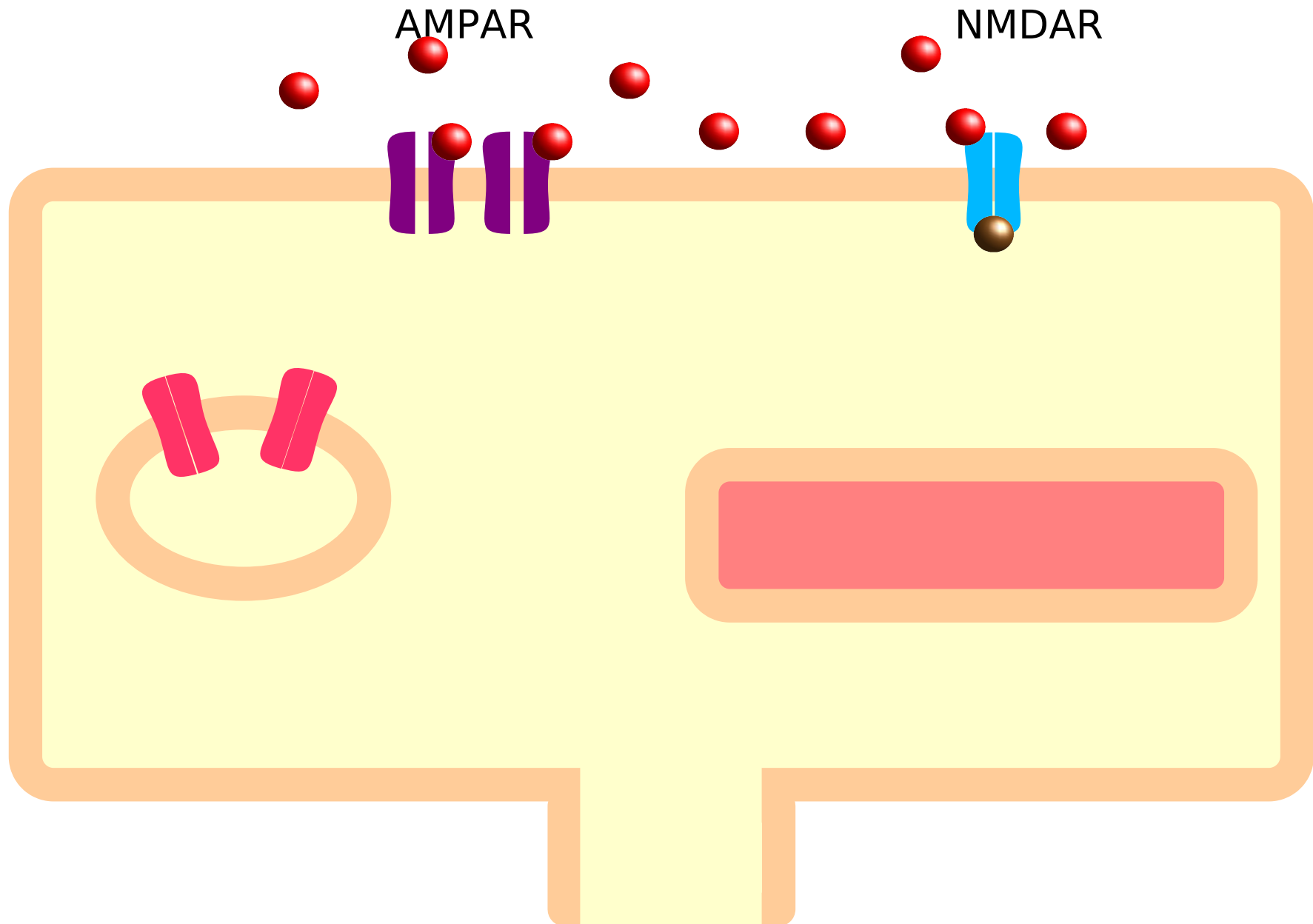


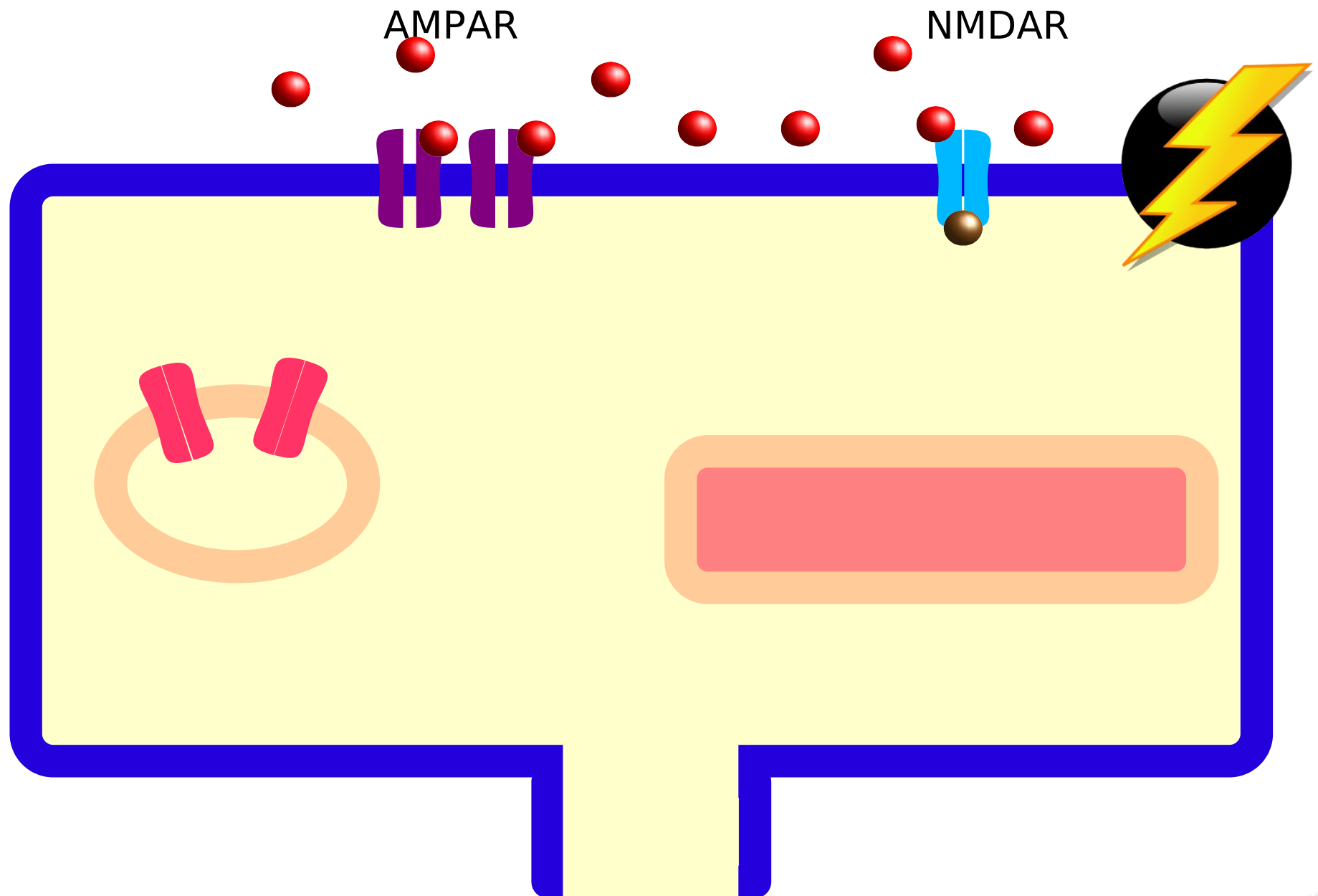


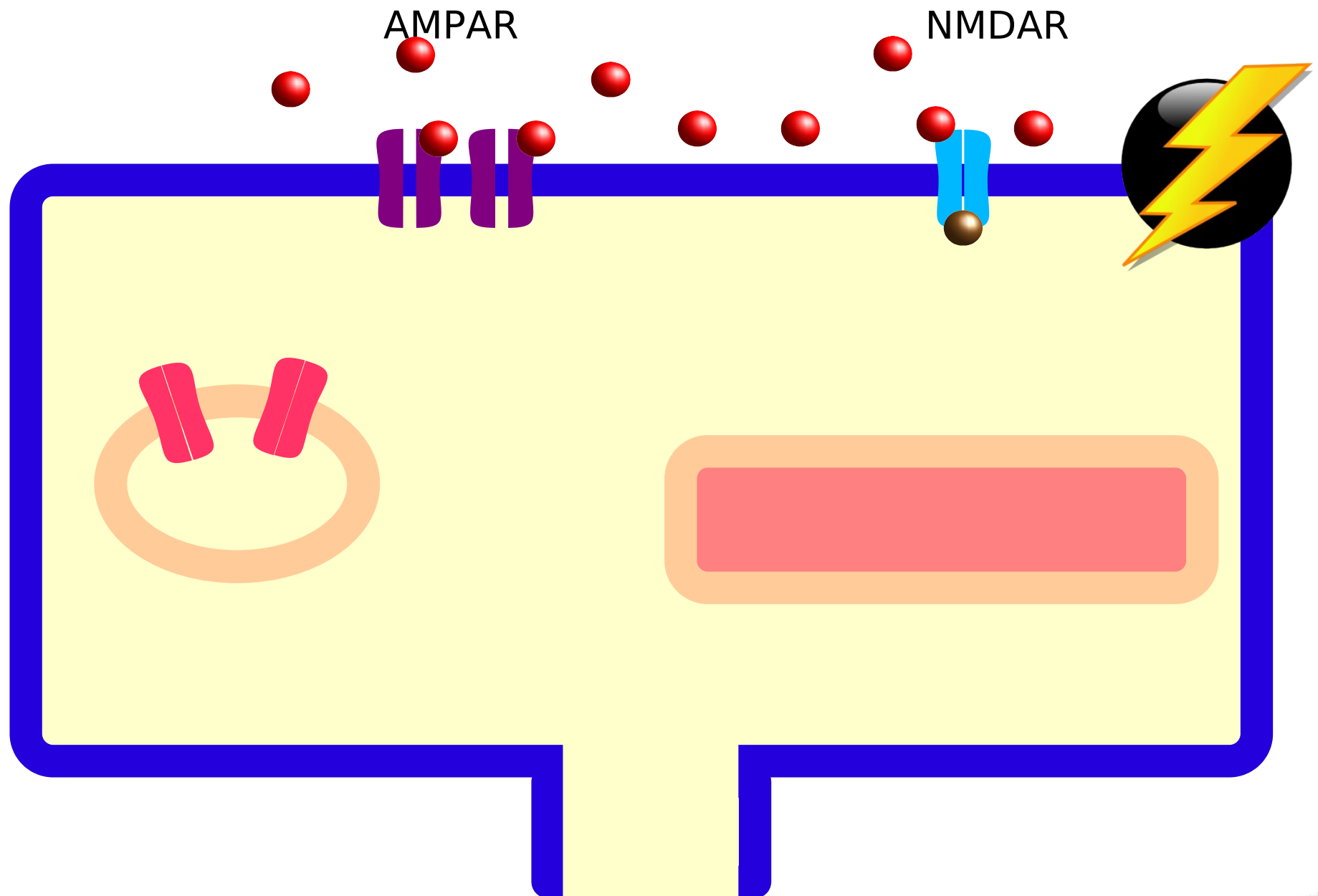


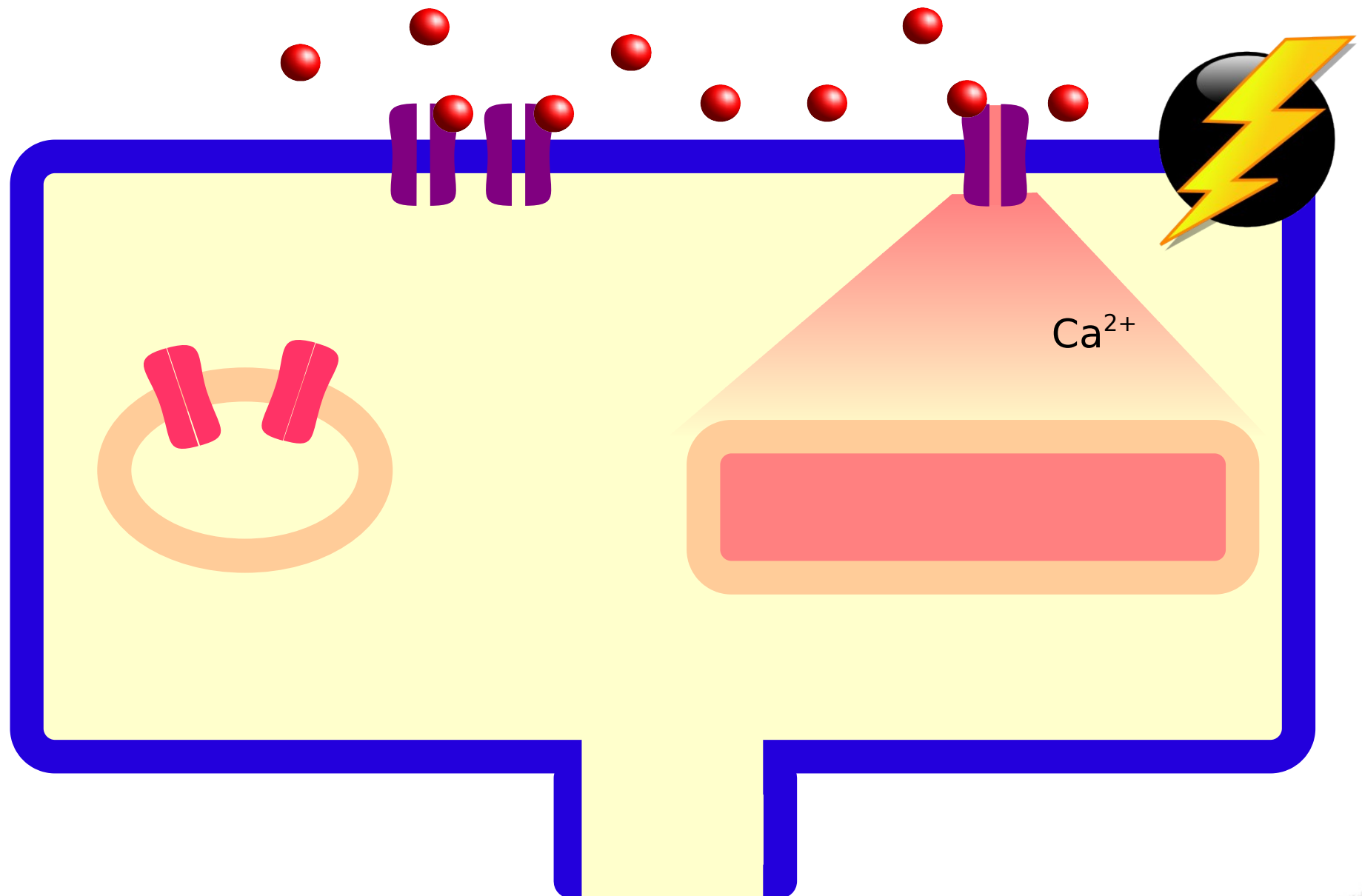
glutamate

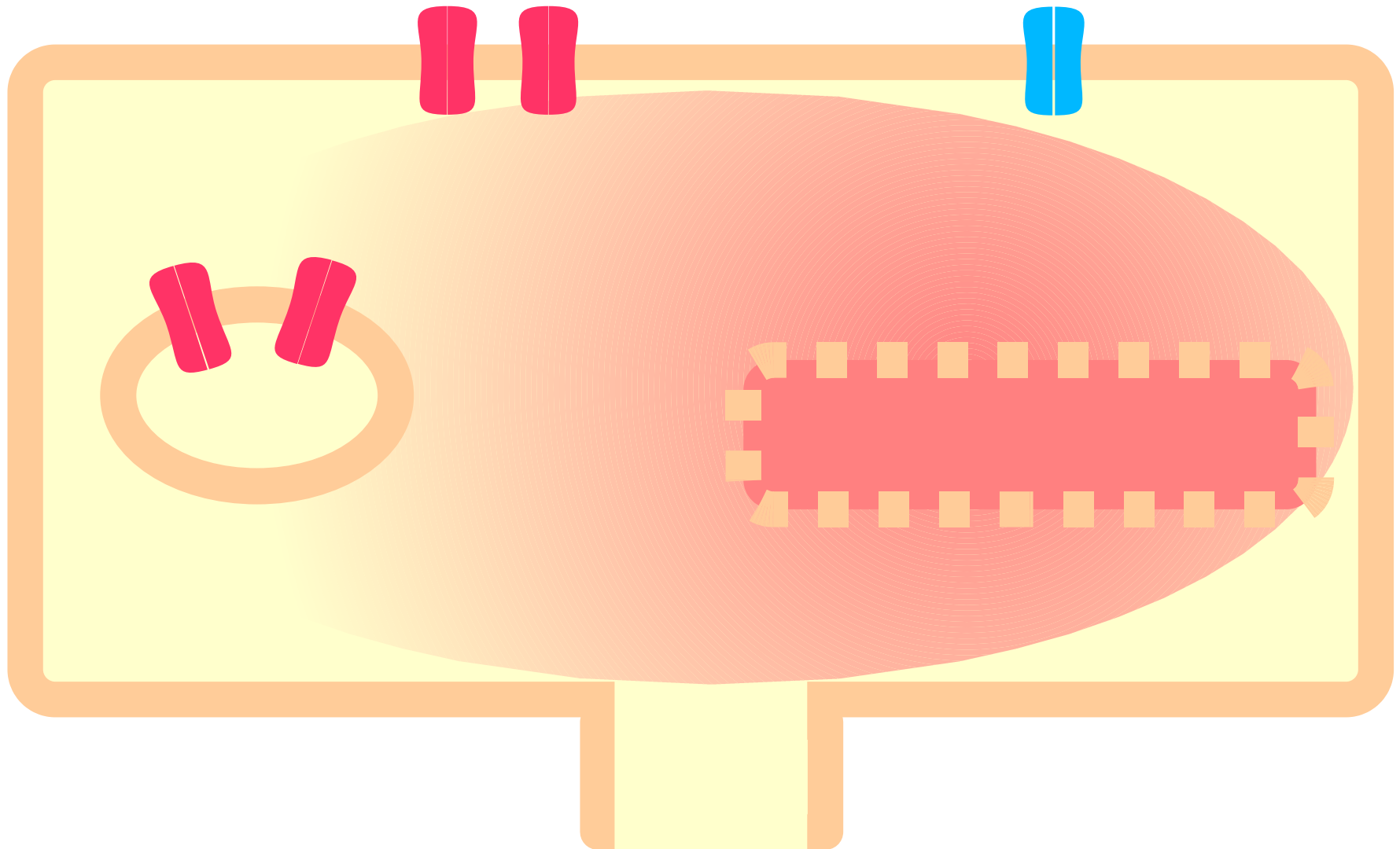


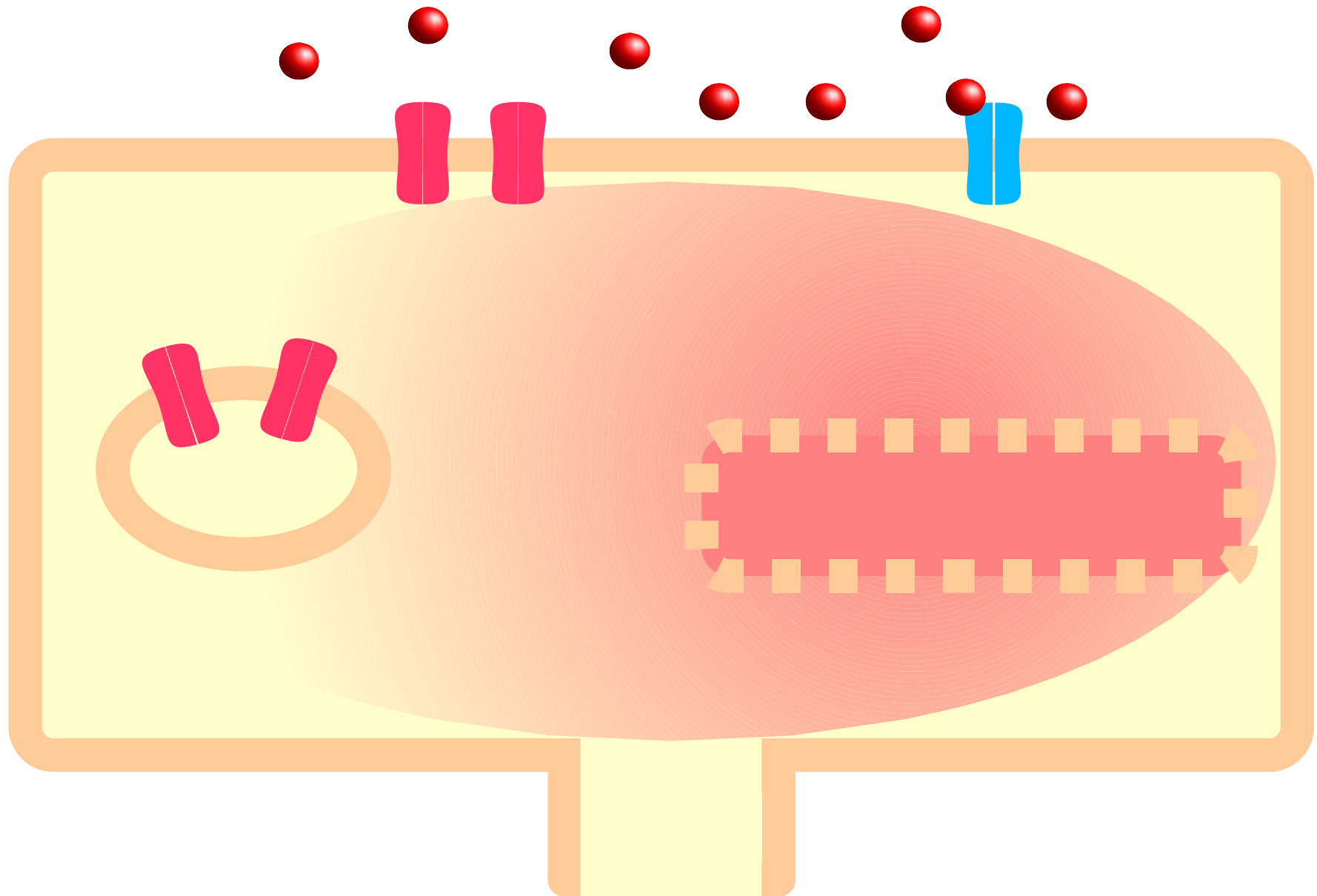


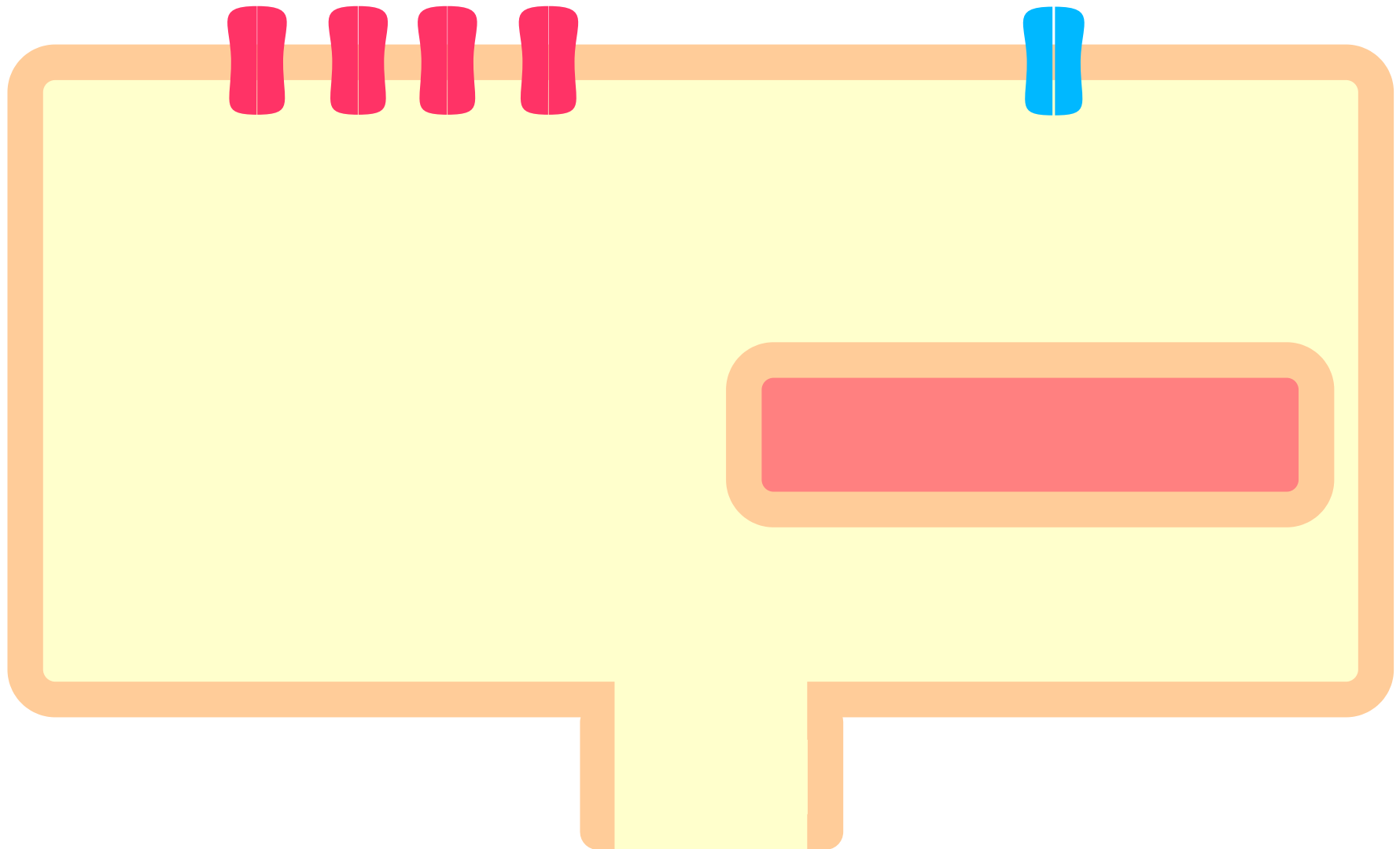


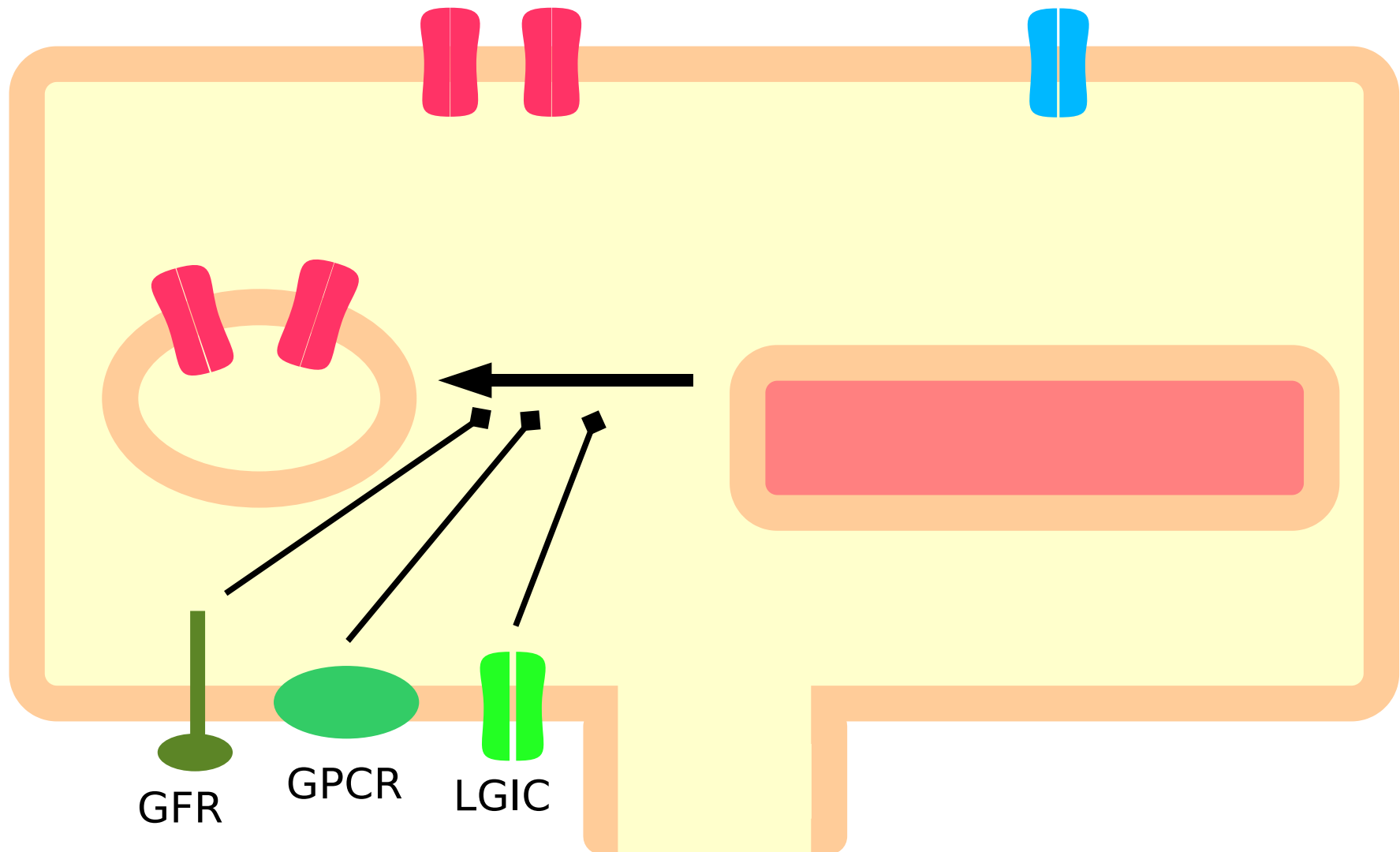


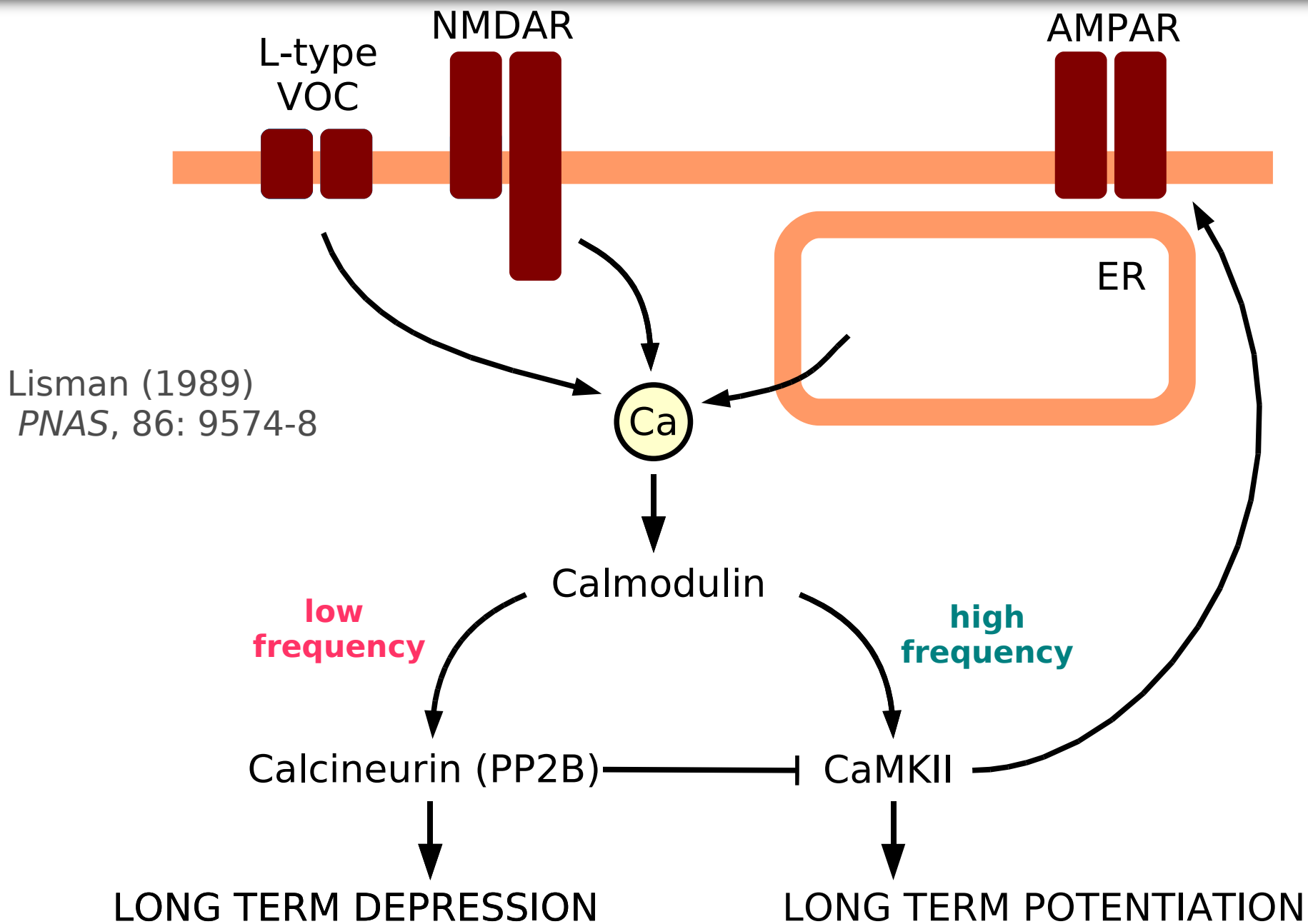










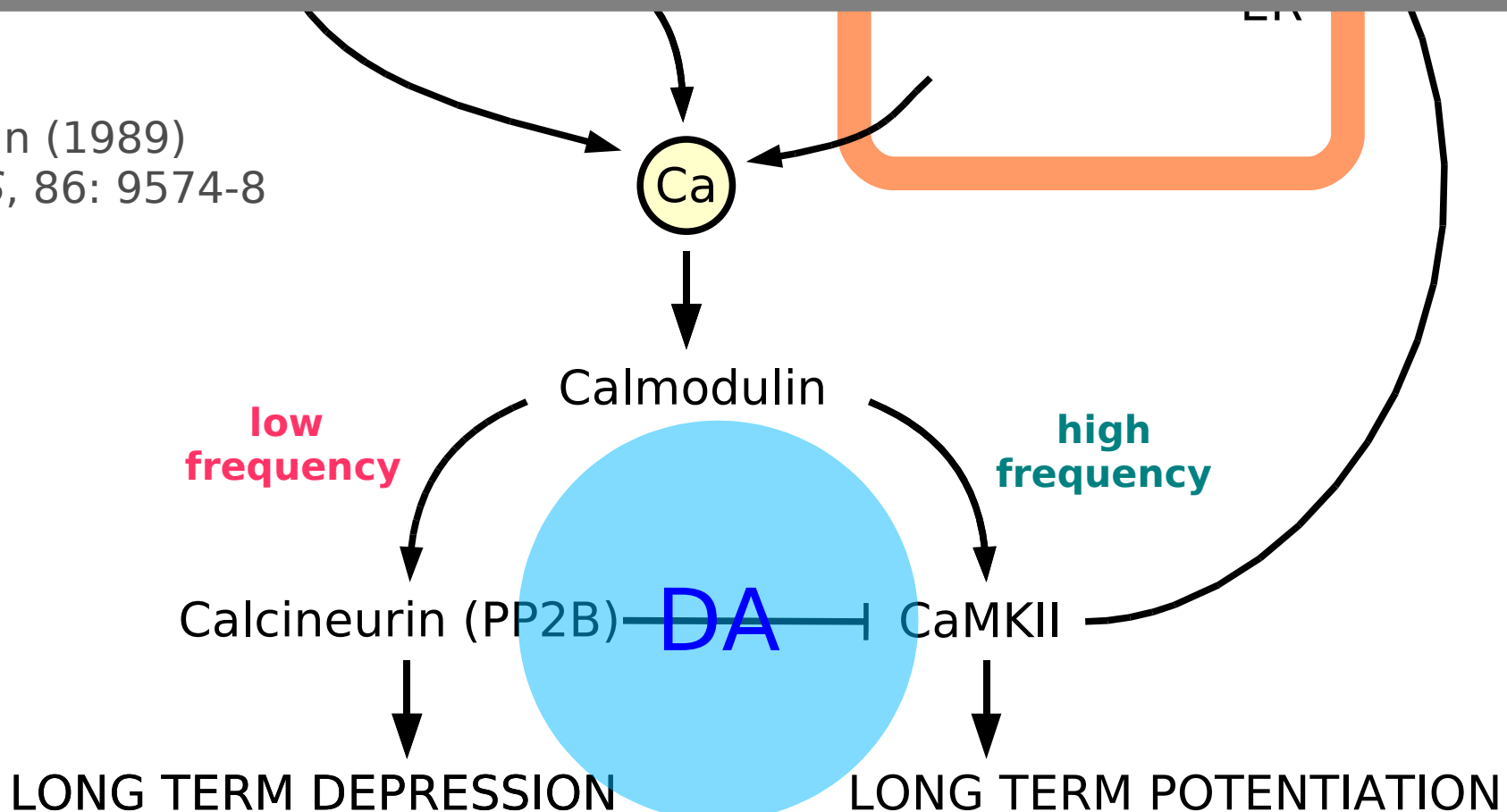




Fernandez E, Schiappa R, Girault JA, Le Novère N (2006) DARPP-32 is a robust integrator of dopamine and glutamate signals. *PLoS Computational Biology*, 2(12): e176.

Le Novère N, Li L, Girault JA (2008) DARPP-32: Molecular integration of phosphorylation potential. *Cellular and Molecular Life Sciences*, 65: 2125-2127

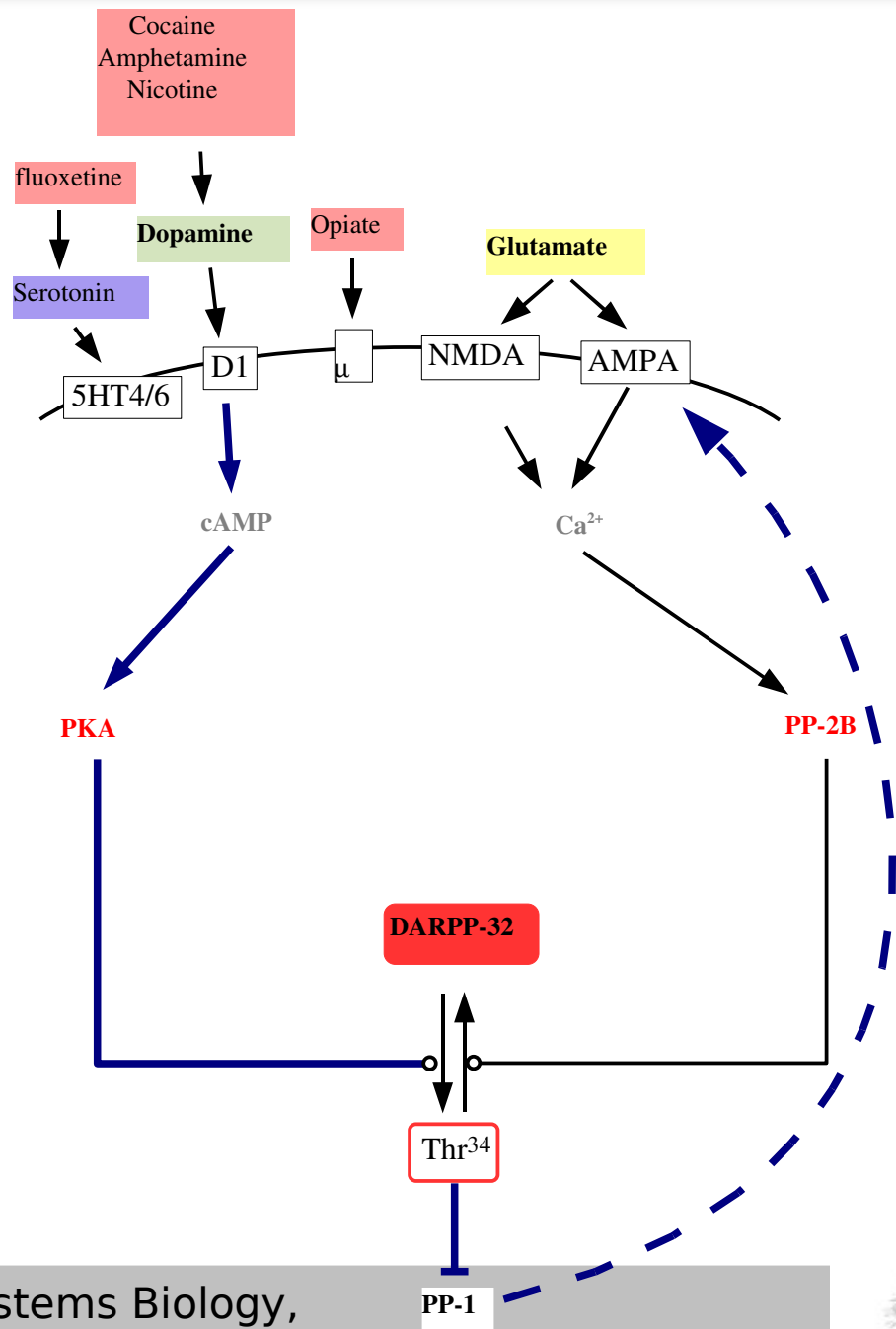
Lisman (1989)
PNAS, 86: 9574-8

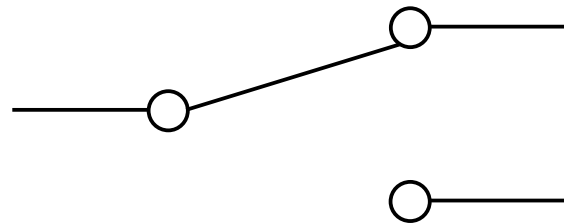


Fernandez E, Schiappa R, Girault JA,
Le Novère N (2006) DARPP-32 is a robust
integrator of dopamine and glutamate signals.
PLoS Computational Biology, 2(12): e176.

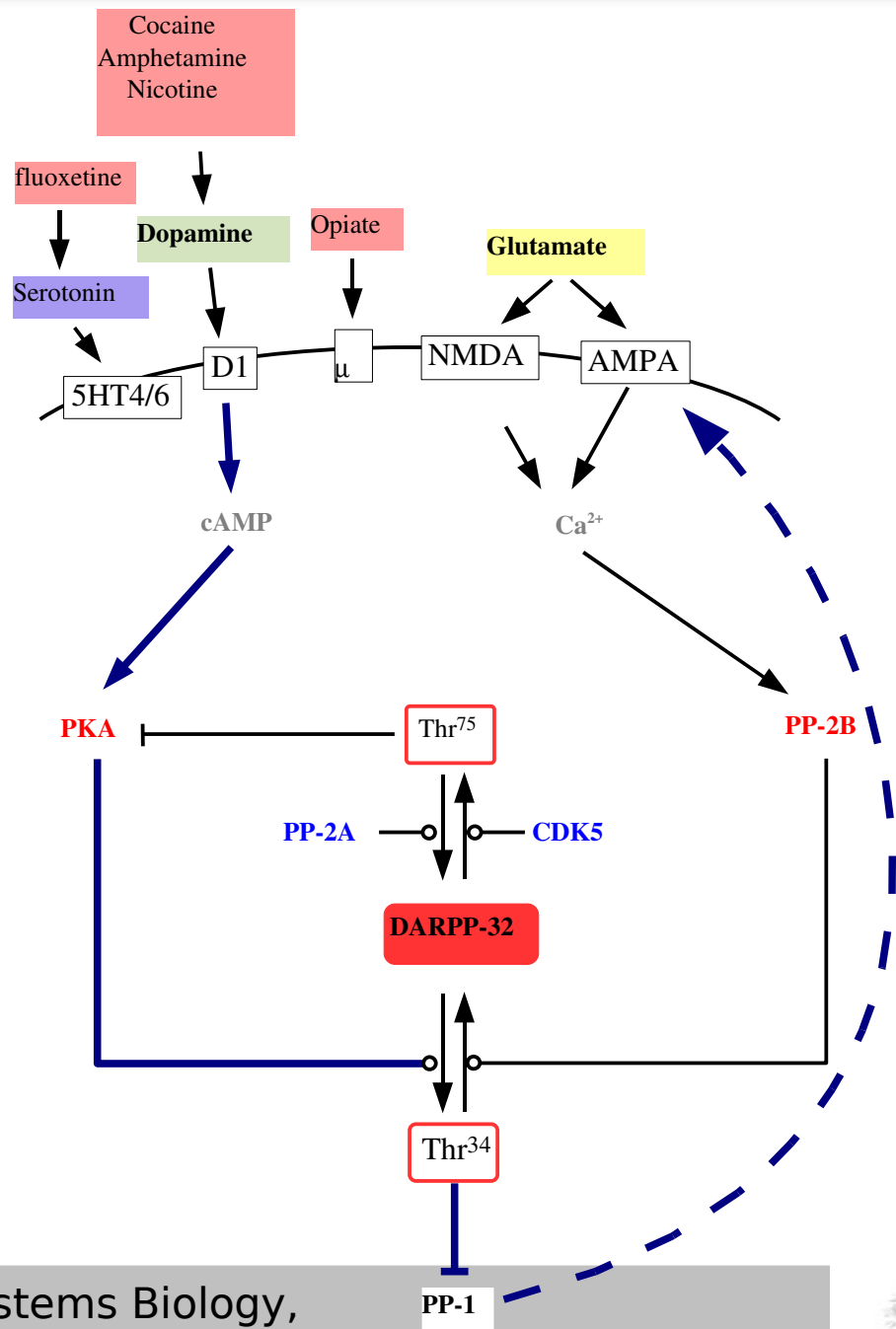


Textbook's view

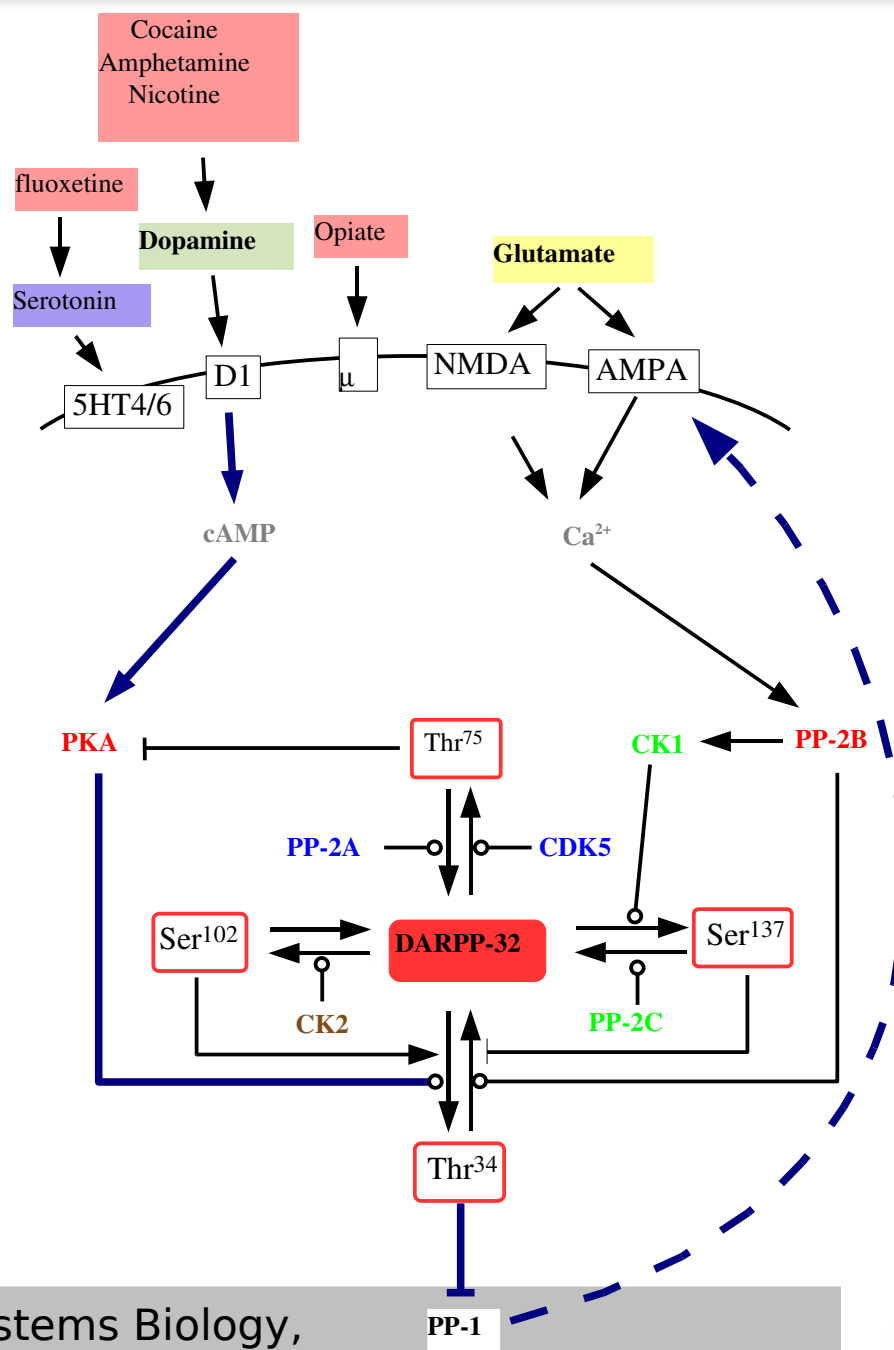




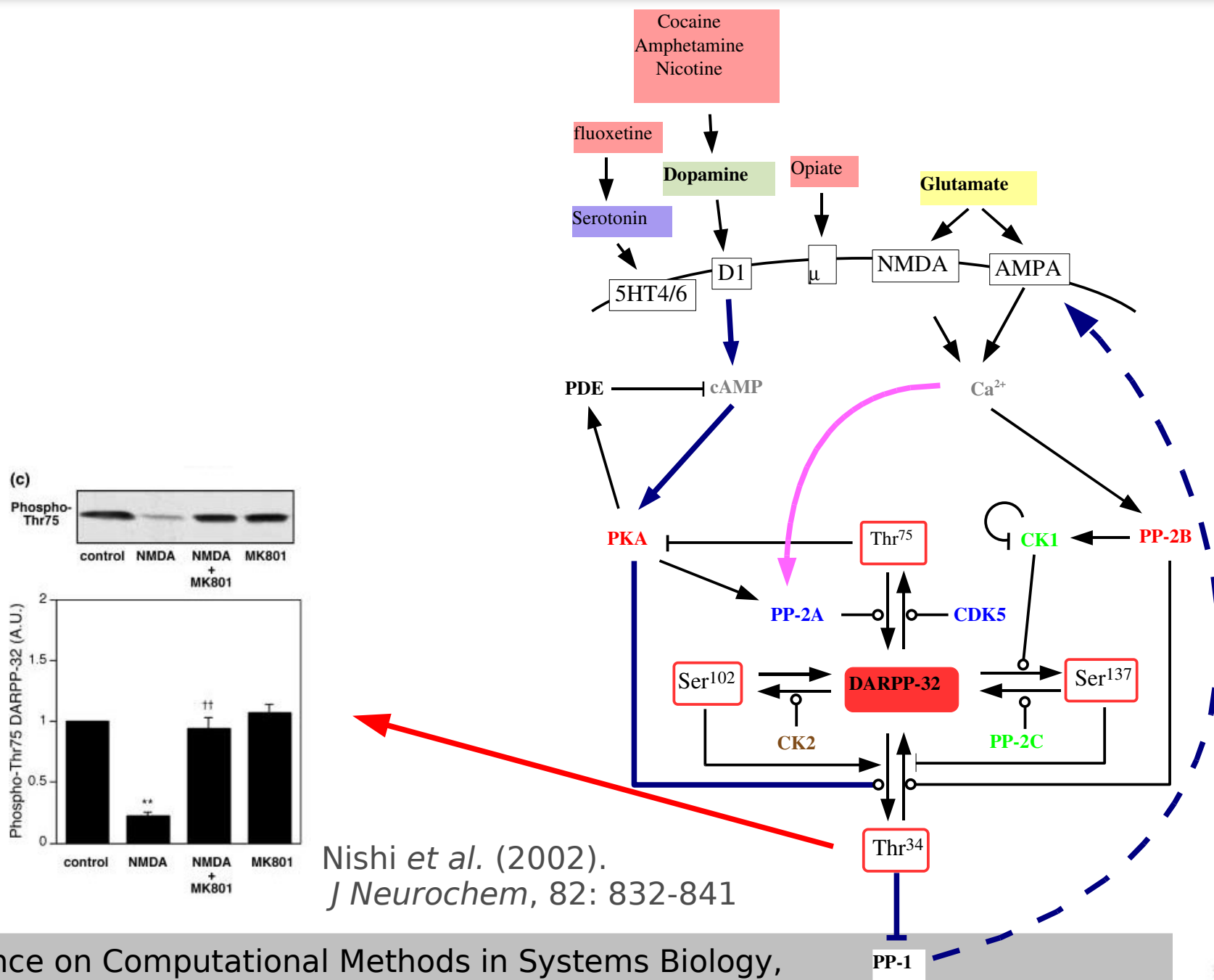
Review article's view

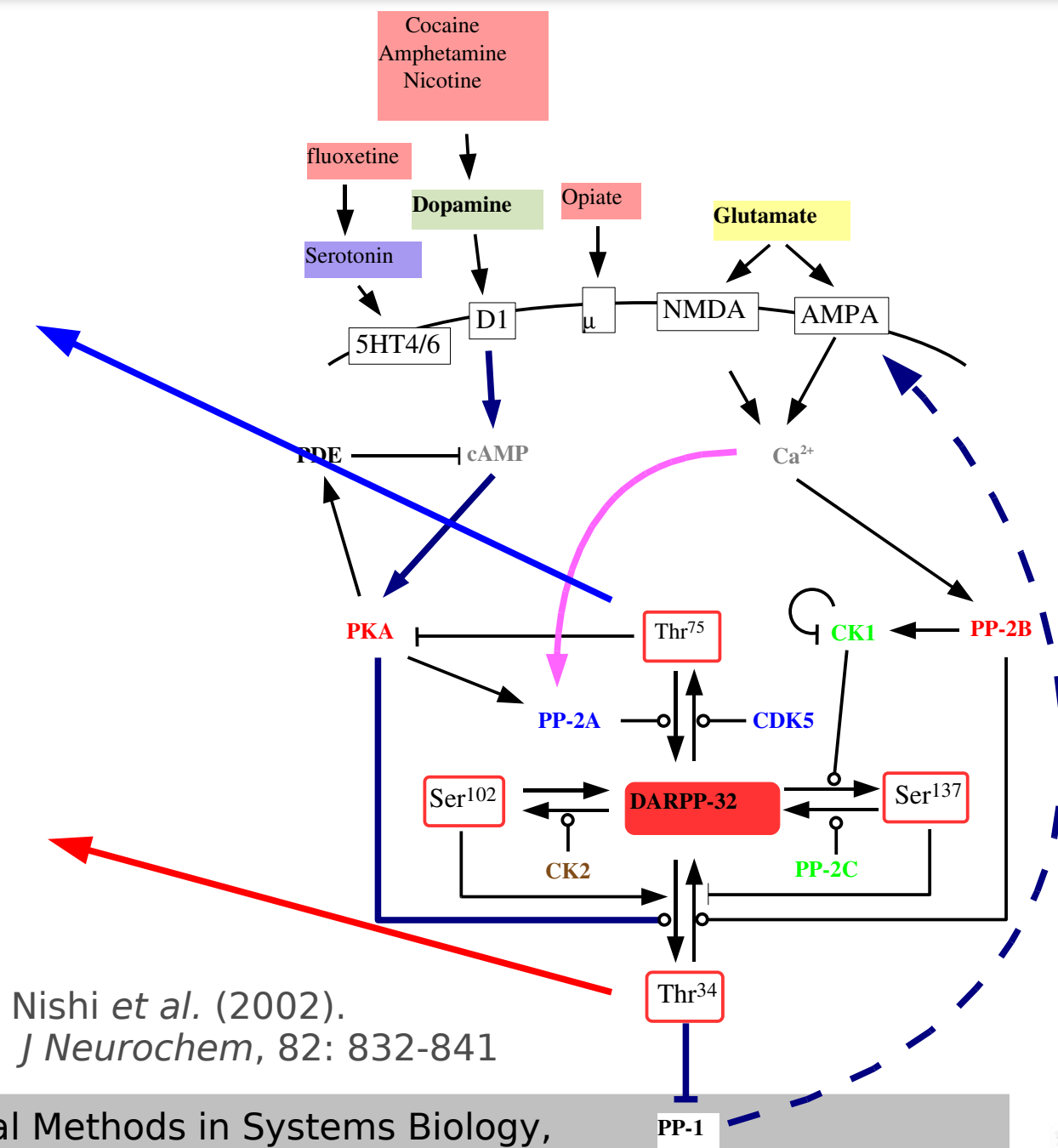
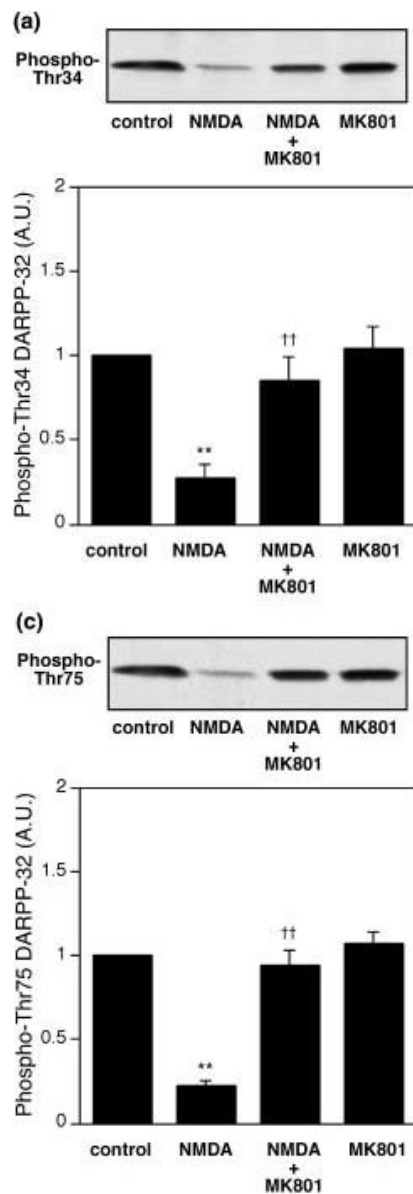


DARPP-32 is a “hub” for phosphatases and kinases in the post-synaptic compartment



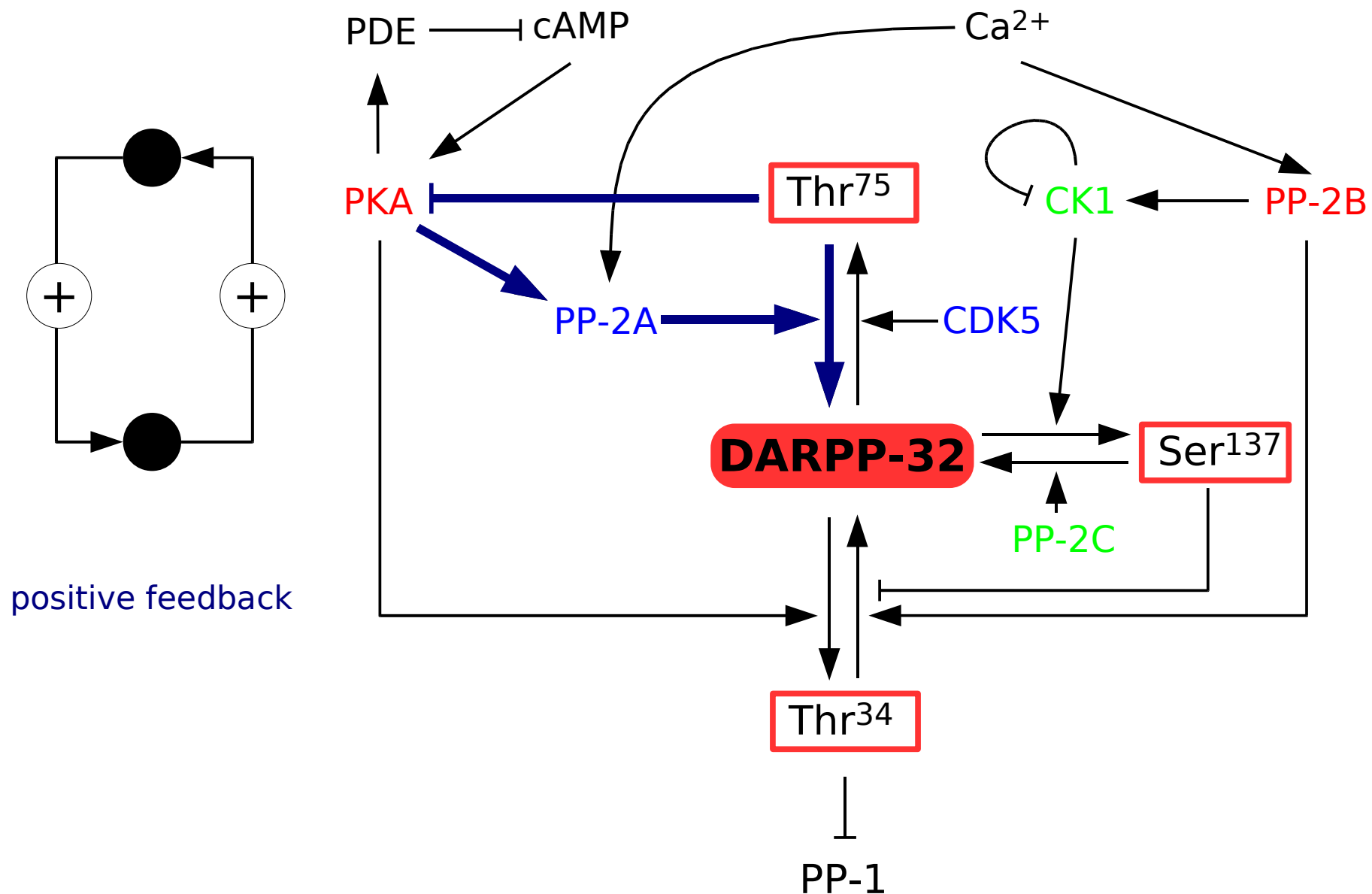


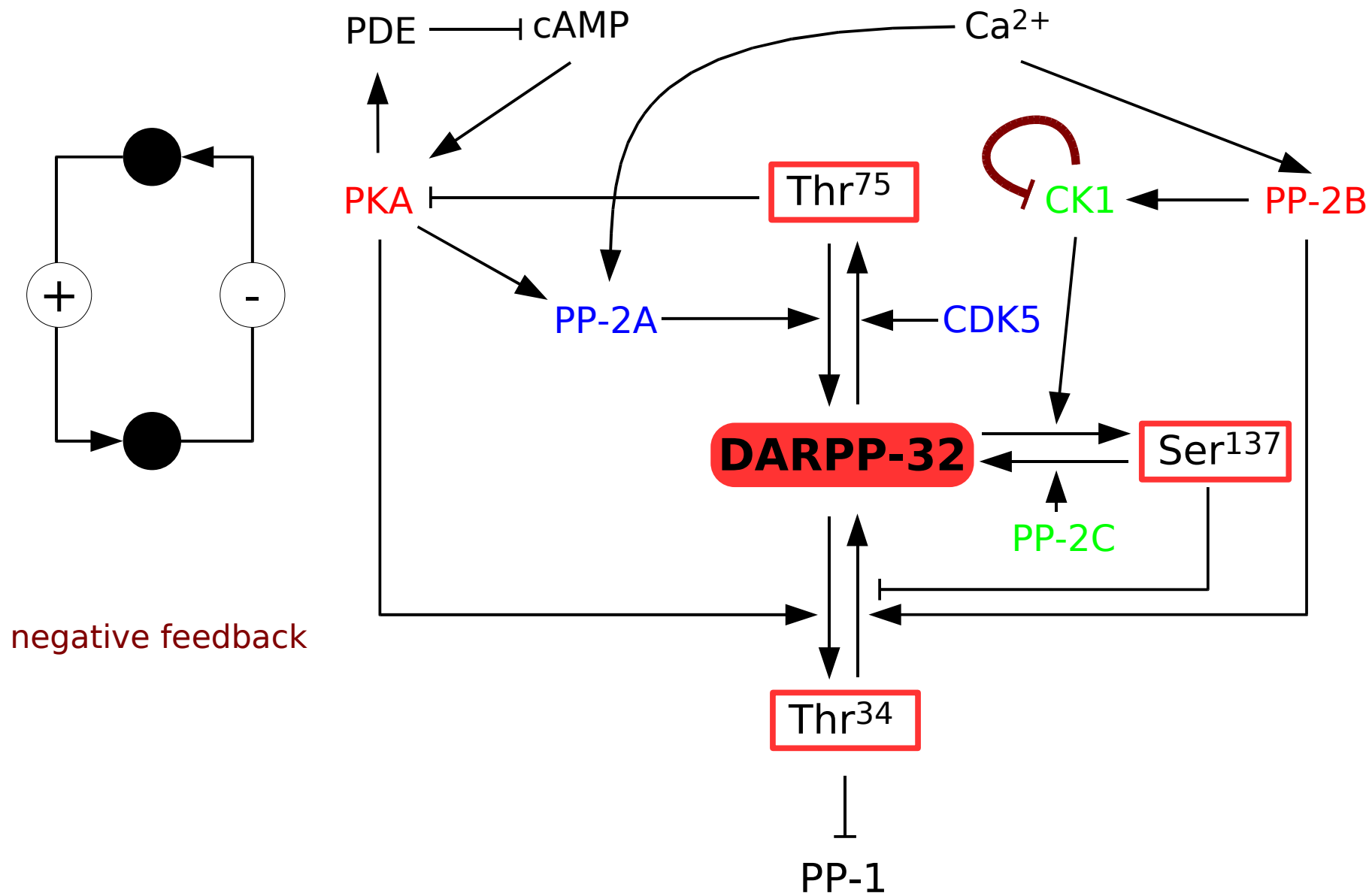


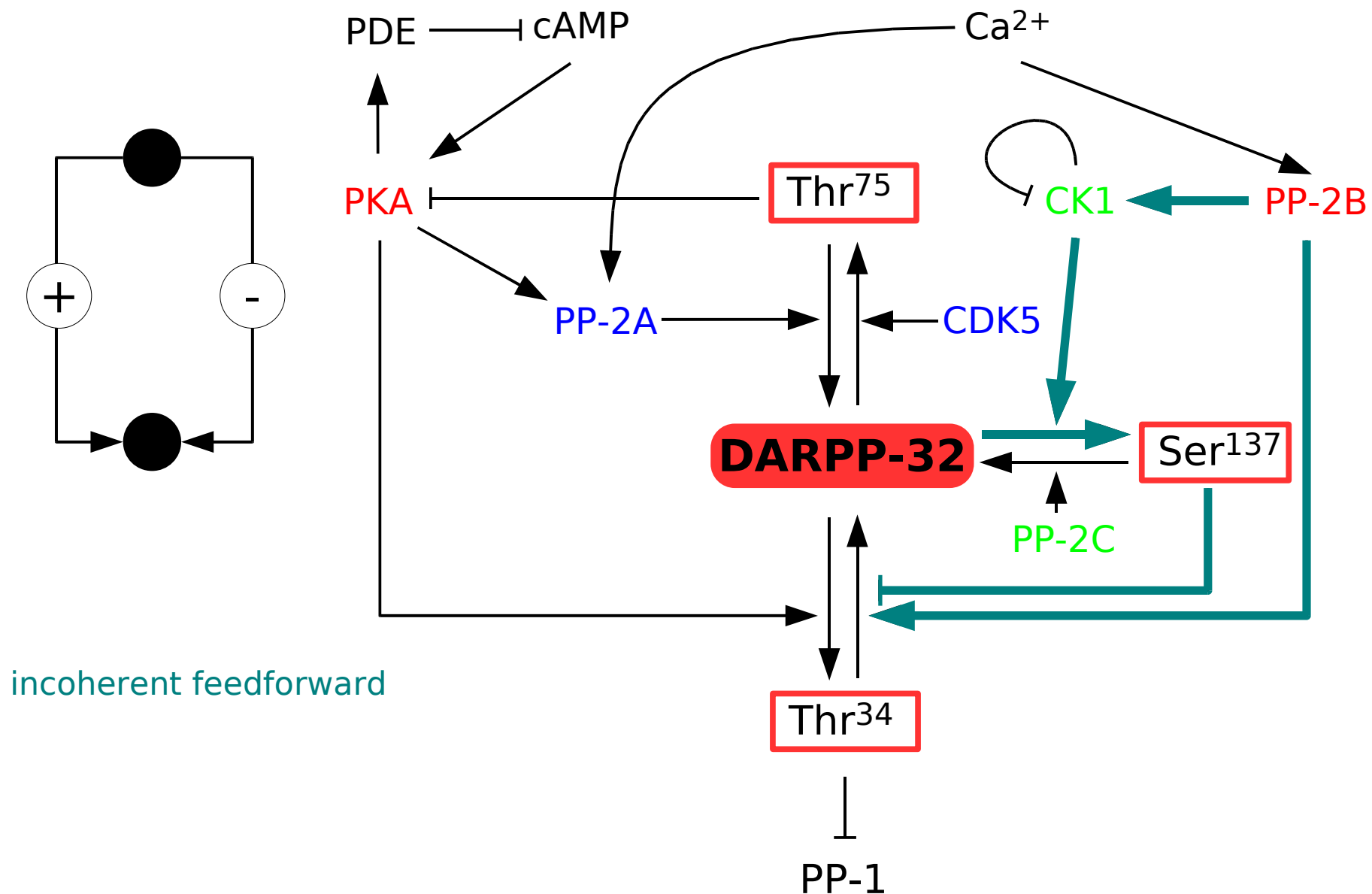


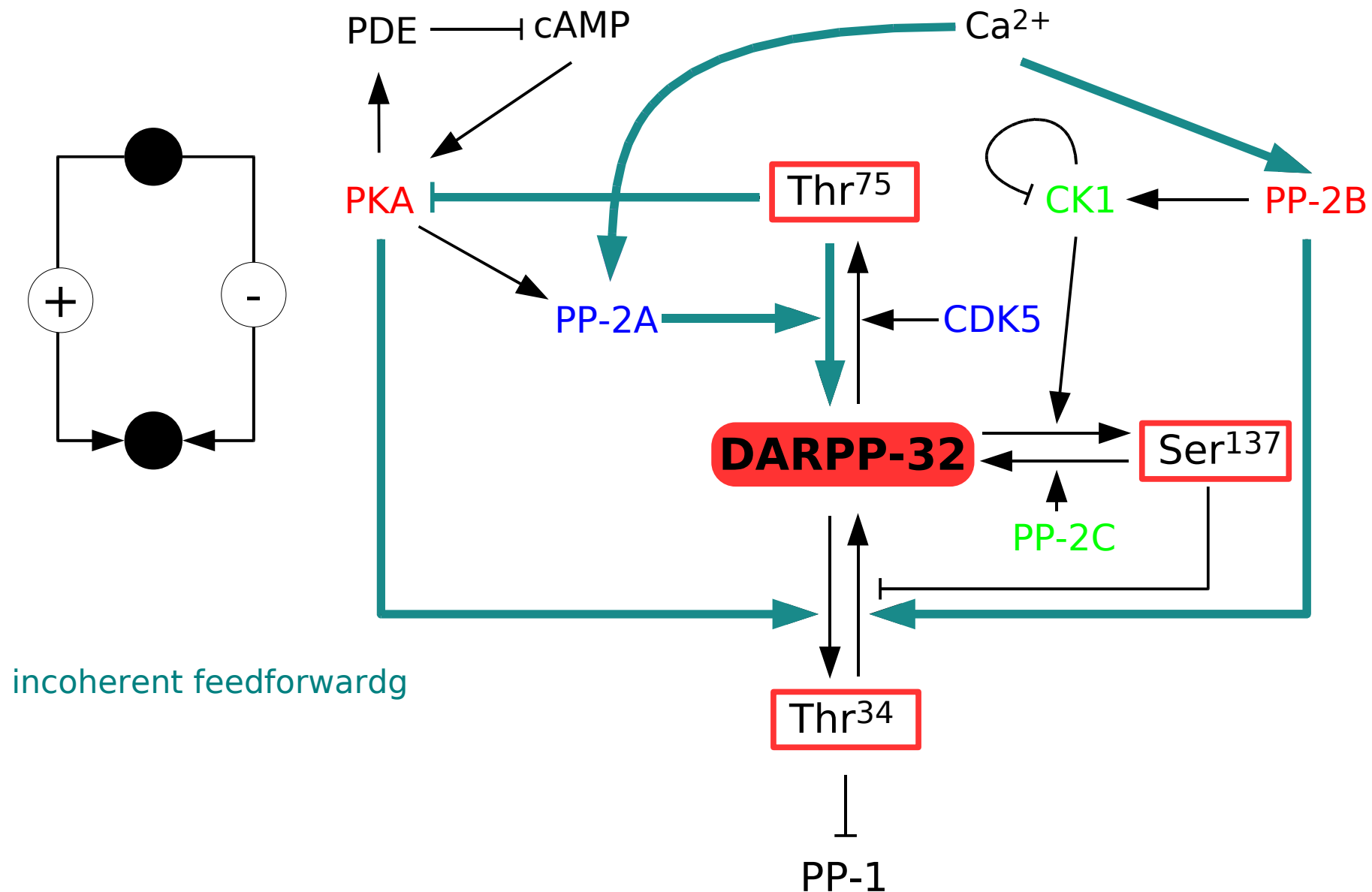
Nishi et al. (2002).
J Neurochem, 82: 832-841

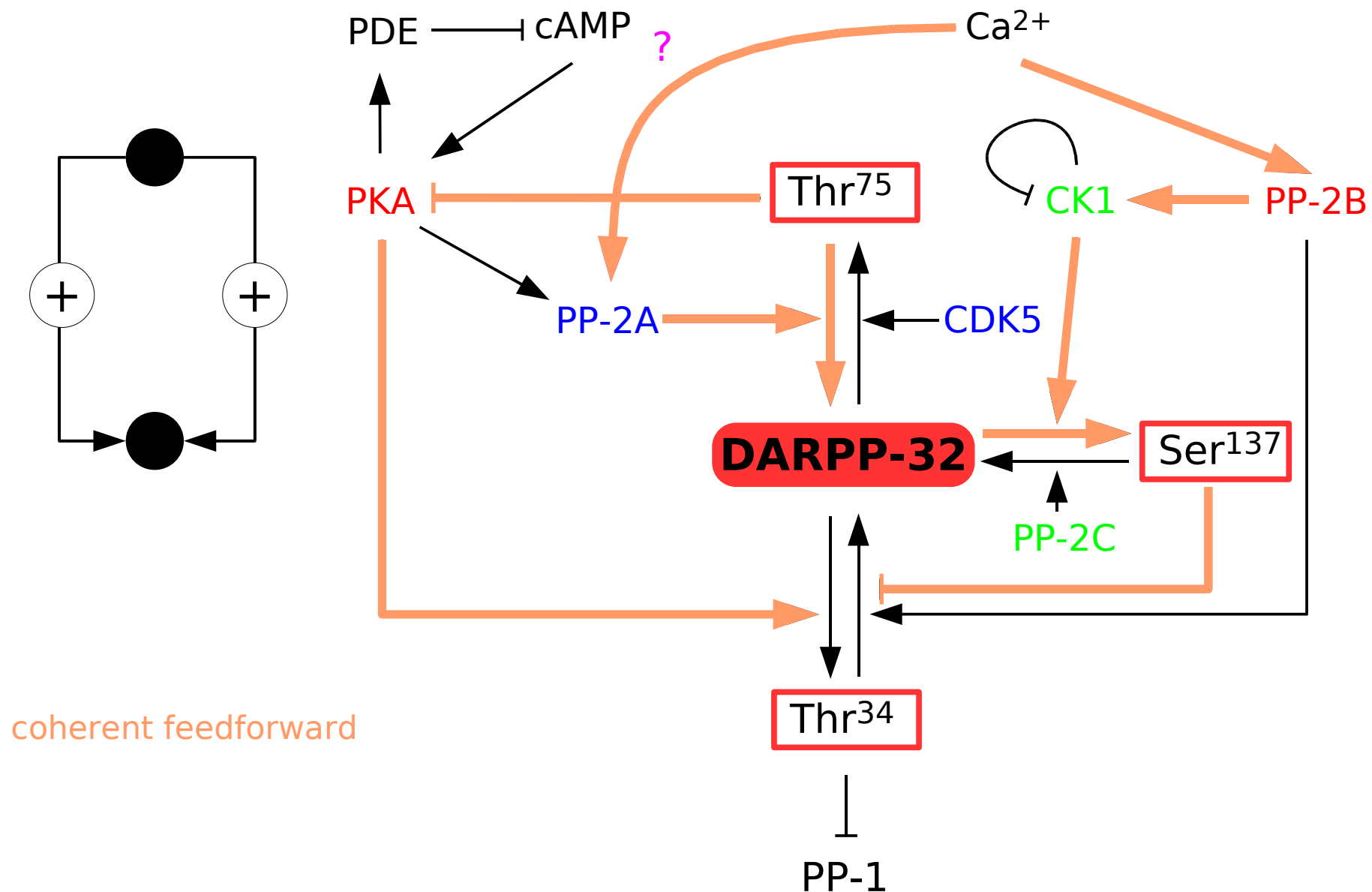


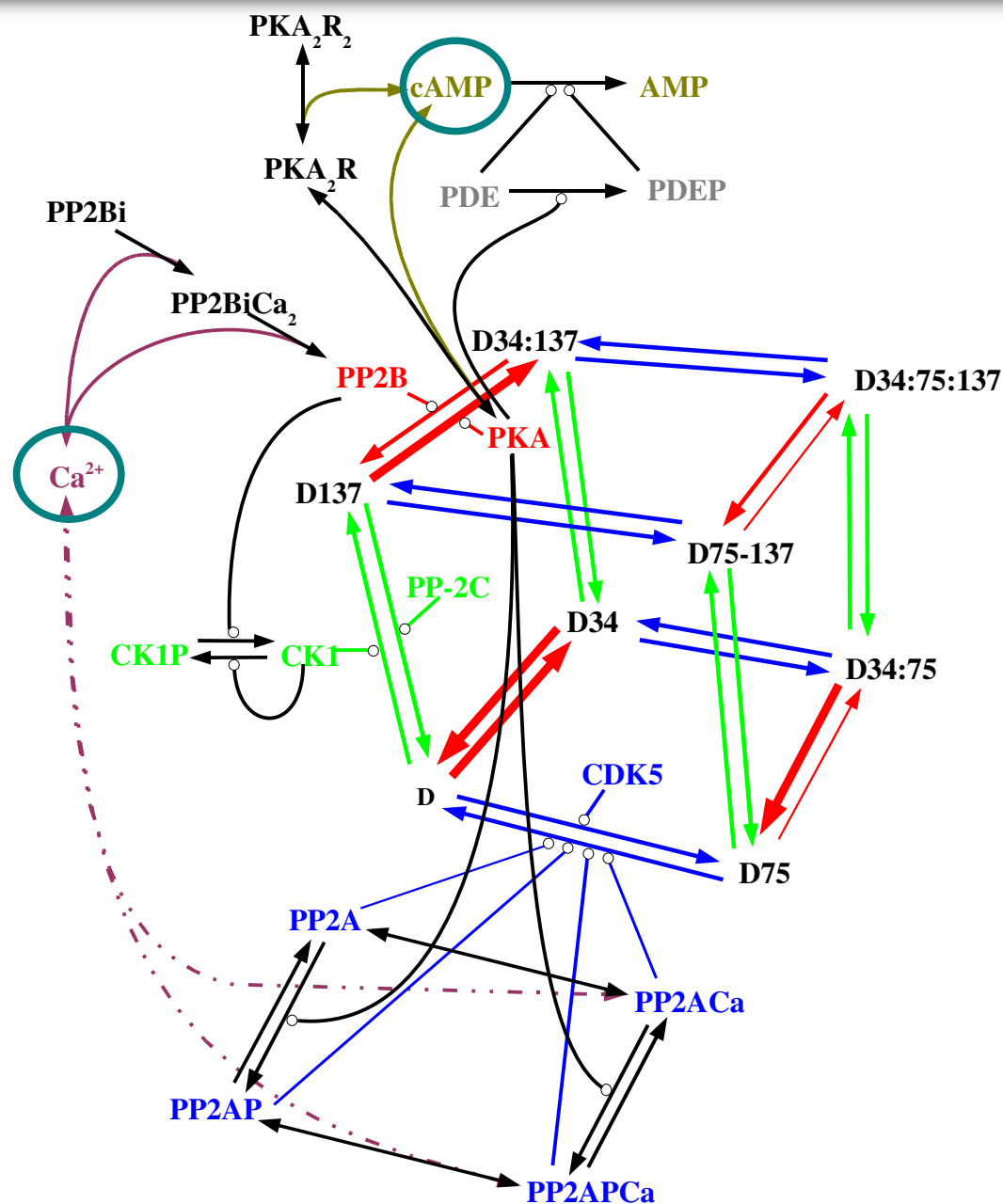












- $D + CDK5 \rightleftharpoons D_CDK5 \rightarrow D75 + CDK5$
- $D75 + PP2A \rightleftharpoons D75_PP2A \rightarrow D + PP2A$
- $D75 + PP2ACa \rightleftharpoons D75_PP2ACa \rightarrow D + PP2ACa$
- $D75 + PP2AP \rightleftharpoons D75_PP2AP \rightarrow D + PP2AP$
- $D75 + PP2APCa \rightleftharpoons D75_PP2APCa \rightarrow D + PP2APCa$
- $D137 + CDK5 \rightleftharpoons D137_CDK5 \rightarrow D75-137 + CDK5$
- $D75-137 + PP2A \rightleftharpoons D75-137_PP2A \rightarrow D137 + PP2A$
- $D75-137 + PP2ACa \rightleftharpoons D75-137_PP2ACa \rightarrow D137 + PP2ACa$
- $D75-137 + PP2AP \rightleftharpoons D75-137_PP2AP \rightarrow D137 + PP2AP$
- $D75-137 + PP2APCa \rightleftharpoons D75-137_PP2APCa \rightarrow D137 + PP2APCa$
- $D34 + CDK5 \rightleftharpoons D34_CDK5 \rightarrow D34-75 + CDK5$
- $D34-75 + PP2A \rightleftharpoons D34-75_PP2A \rightarrow D34 + PP2A$
- $D34-75 + PP2ACa \rightleftharpoons D34-75_PP2ACa \rightarrow D34 + PP2ACa$
- $D34-75 + PP2AP \rightleftharpoons D34-75_PP2AP \rightarrow D34 + PP2AP$
- $D34-75 + PP2APCa \rightleftharpoons D34-75_PP2APCa \rightarrow D34 + PP2APCa$
- $D34-137 + CDK5 \rightleftharpoons D34-137_CDK5 \rightarrow D34-75-137 + CDK5$
- $D34-75-137 + PP2A \rightleftharpoons D34-75-137_PP2A \rightarrow D34-137 + PP2A$
- $D34-75-137 + PP2ACa \rightleftharpoons D34-75-137_PP2ACa \rightarrow D34-137 + PP2ACa$
- $D34-75-137 + PP2AP \rightleftharpoons D34-75-137_PP2AP \rightarrow D34-137 + PP2AP$
- $D34-75-137 + PP2APCa \rightleftharpoons D34-75-137_PP2APCa \rightarrow D34-137 + PP2APCa$
- $D + CK1 \rightleftharpoons D_CK1 \rightarrow D137 + CK1$
- $D137 + PP2C \rightleftharpoons D137_PP2C \rightarrow D + PP2C$
- $D75 + CK1 \rightleftharpoons D75_CK1 \rightarrow D75-137 + CK1$
- $D75-137 + PP2C \rightleftharpoons D75-137_PP2C \rightarrow D75 + PP2C$
- $D34 + CK1 \rightleftharpoons D34_CK1 \rightarrow D34-137 + CK1$
- $D34-75 + PP2C \rightleftharpoons D34-75_PP2C \rightarrow D75 + PP2C$
- $D34-75 + CK1 \rightleftharpoons D34-75_CK1 \rightarrow D34-75-137 + CK1$
- $D34-75-137 + PP2C \rightleftharpoons D34-75-137_PP2C \rightarrow D34-75 + PP2C$
- $CK1 + CK1 \rightleftharpoons CK1_CK1 \rightarrow CK1P + CK1$
- $CK1P + PP2B \rightleftharpoons CK1P_PP2B \rightarrow CK1 + PP2B$

79 species, 151 reactions

- $D + PKA \rightleftharpoons D_PKA \rightarrow D34 + PKA$
- $D34 + PP2B \rightleftharpoons D34_PP2B \rightarrow D + PP2B$
- $D75 + PKA \rightleftharpoons D75_PKA \rightarrow D34-75 + PKA$
- $D34-75 + PP2B \rightleftharpoons D34-75_PP2B \rightarrow D75 + PP2B$
- $D137 + PKA \rightleftharpoons D137_PKA \rightarrow D34-137 + PKA$
- $D34-137 + PP2B \rightleftharpoons D34-137_PP2B \rightarrow D137 + PP2B$
- $D75-137 + PKA \rightleftharpoons D75-137_PKA \rightarrow D34-75-137 + PKA$
- $D34-75-137 + PP2B \rightleftharpoons D34-75-137_PP2B \rightarrow D75-137 + PP2B$
- $PP2A + PKA \rightleftharpoons PP2A_PKA \rightarrow PP2AP + PKA$
- $PP2ACa + PKA \rightleftharpoons PP2ACa_PKA \rightarrow PP2APCa + PKA$
- $\emptyset \rightarrow Ca^{2+}$
- $Ca^{2+} \rightarrow \emptyset$
- $PP2Bi + 2Ca \rightleftharpoons PP2Bi_Ca2$
- $PP2Bi_Ca + 2Ca \rightleftharpoons PP2B$
- $PP2A + Ca \rightleftharpoons PP2ACa$
- $PP2AP + Ca \rightleftharpoons PP2APCa$
- $R2_PKA2 + cAMP \rightleftharpoons cAMP_R2_PKA2$
- $cAMP_R2_PKA2 + cAMP \rightleftharpoons cAMP2_R2_PKA2$
- $cAMP2_R2_PKA2 + cAMP \rightleftharpoons cAMP3_R2_PKA2$
- $cAMP3_R2_PKA2 + cAMP \rightleftharpoons cAMP4_R2_PKA2$
- $cAMP4_R2_PKA2 \rightleftharpoons cAMP4_R2_PKA + PKA$
- $cAMP4_R2_PKA \rightleftharpoons cAMP4_R2 + PKA$
- $PKA + PDE \rightleftharpoons PKA_PDE \rightarrow PKA + PDEP$
- $cAMP + PDE \rightleftharpoons cAMP_PDE \rightarrow AMP + PDE$
- $cAMP + PDEP \rightleftharpoons cAMP_PDEP \rightarrow AMP + PDEP$

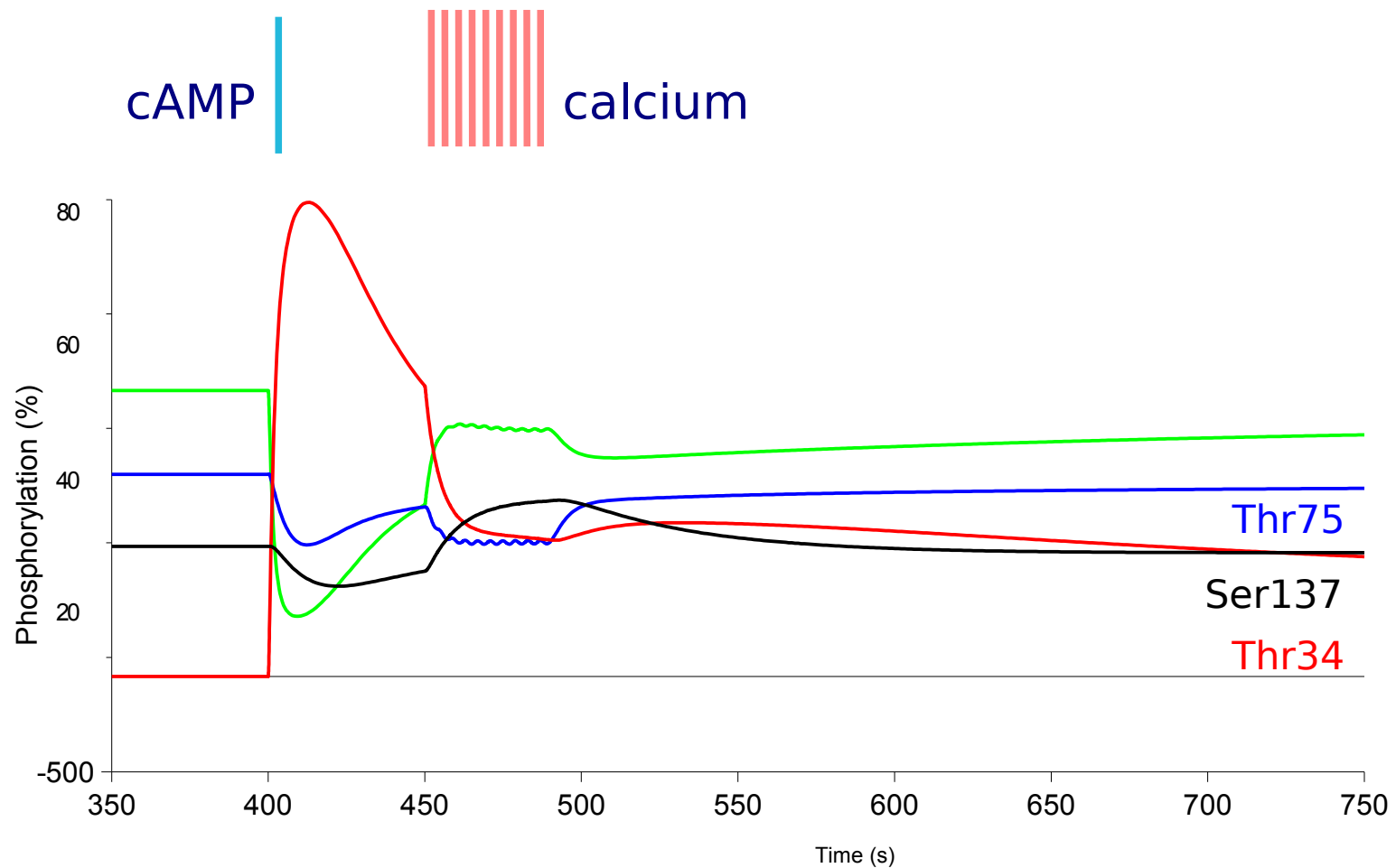


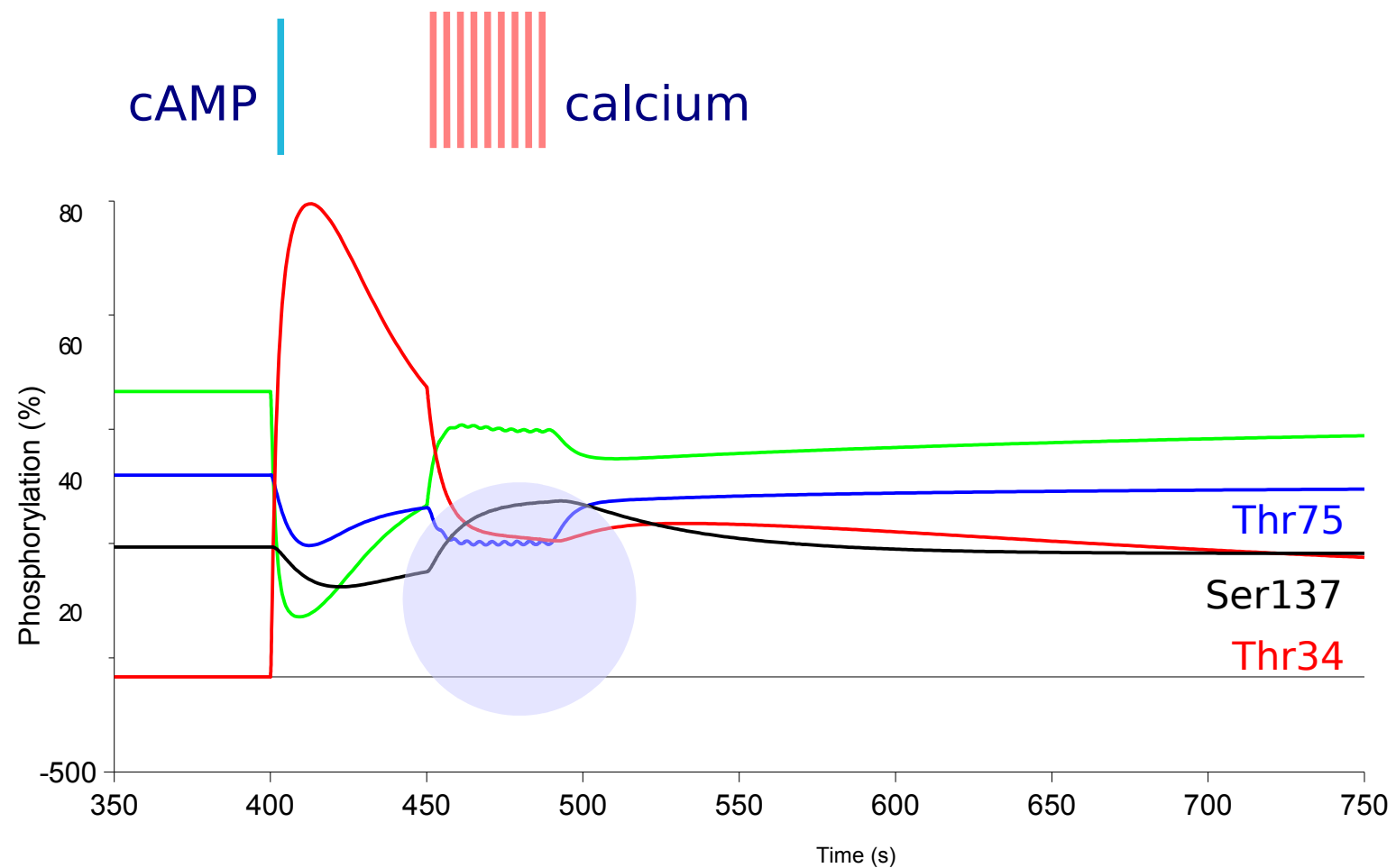


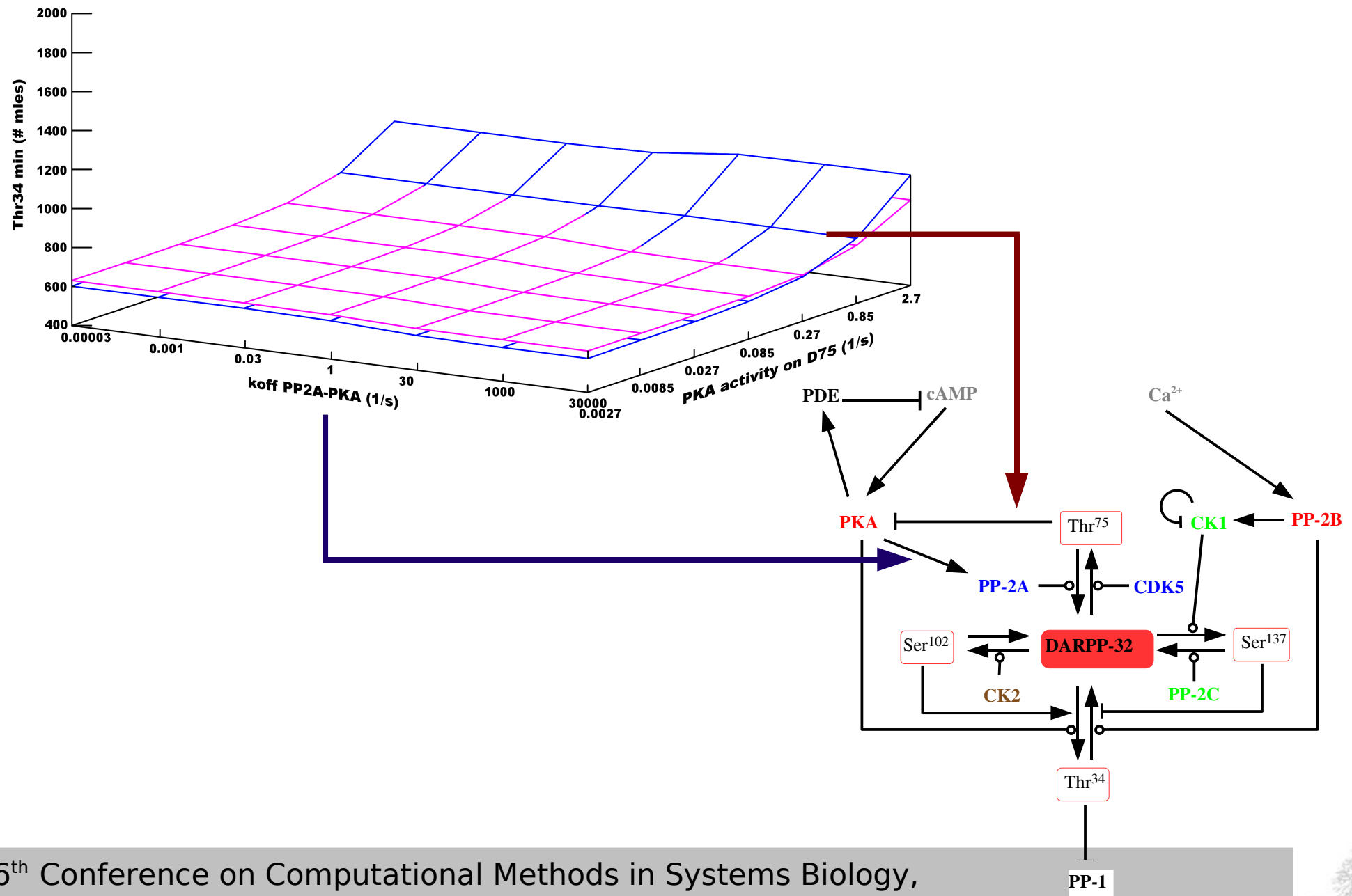
E-Cell

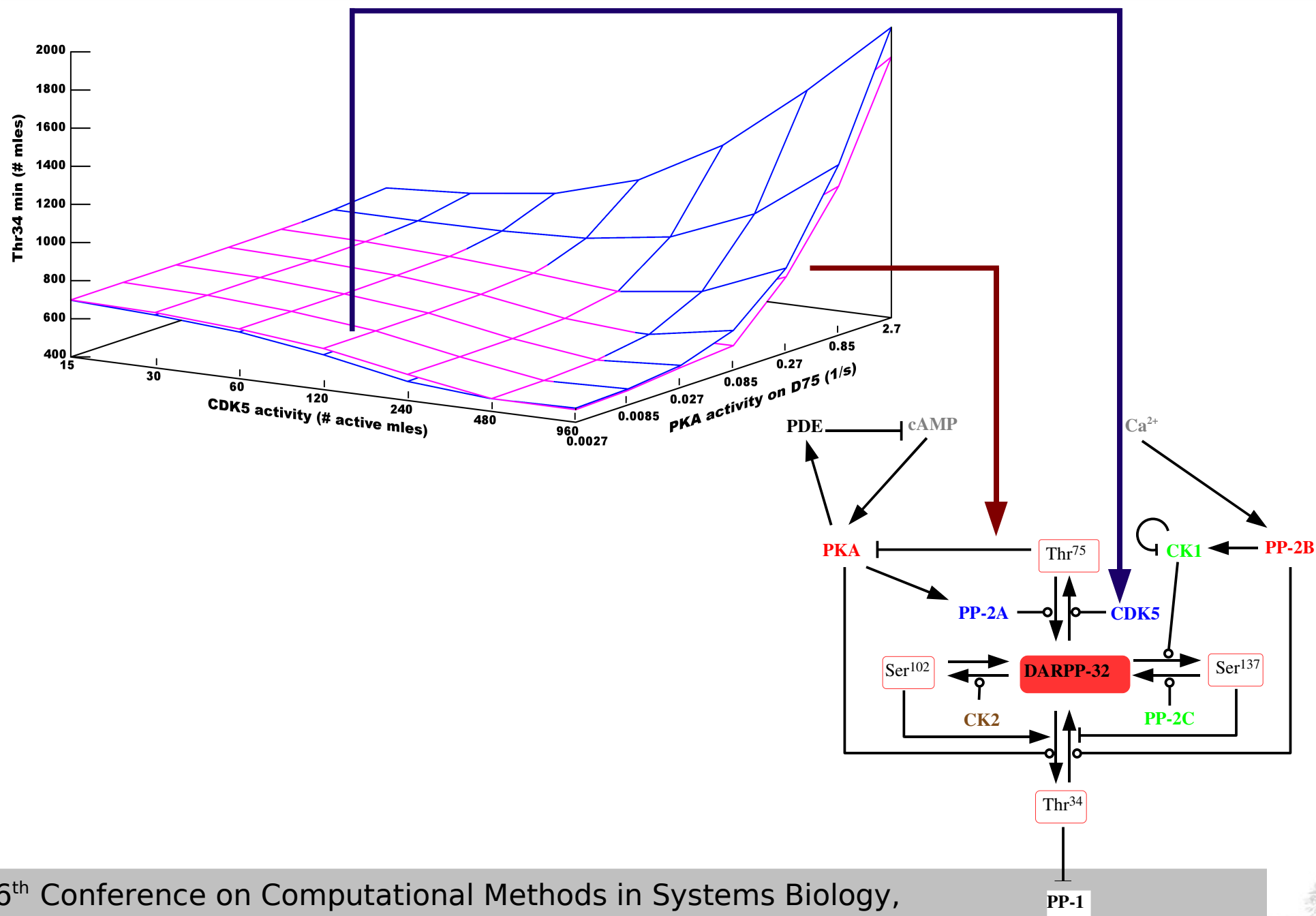
- Discrete simulator (# molecules)
- Hybrid simulator (stochastic and deterministic)
- Advanced numerical solver
- multi-compartments
- Python scripting interface ➡ arbitrary events
- Command-line + GUI

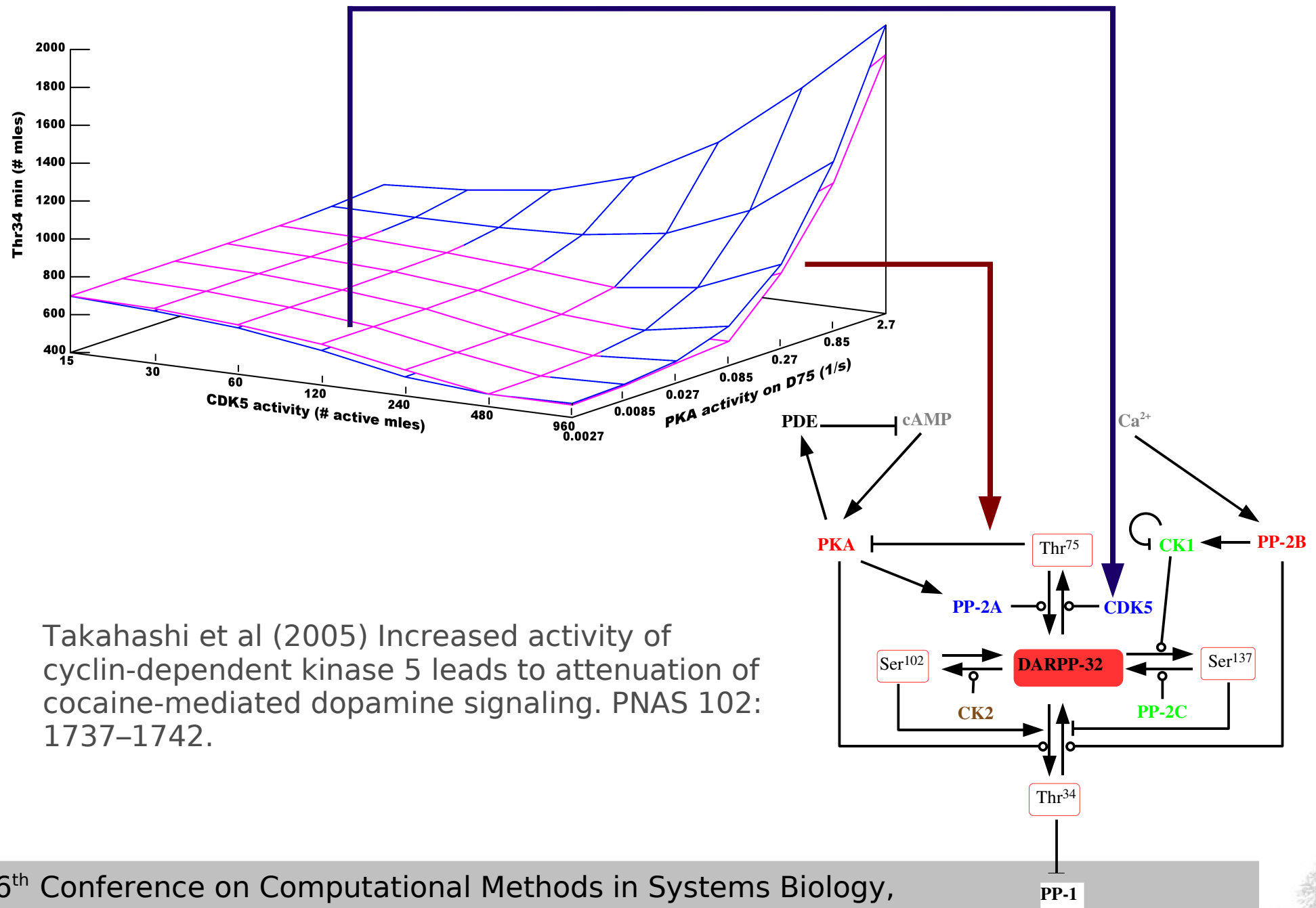






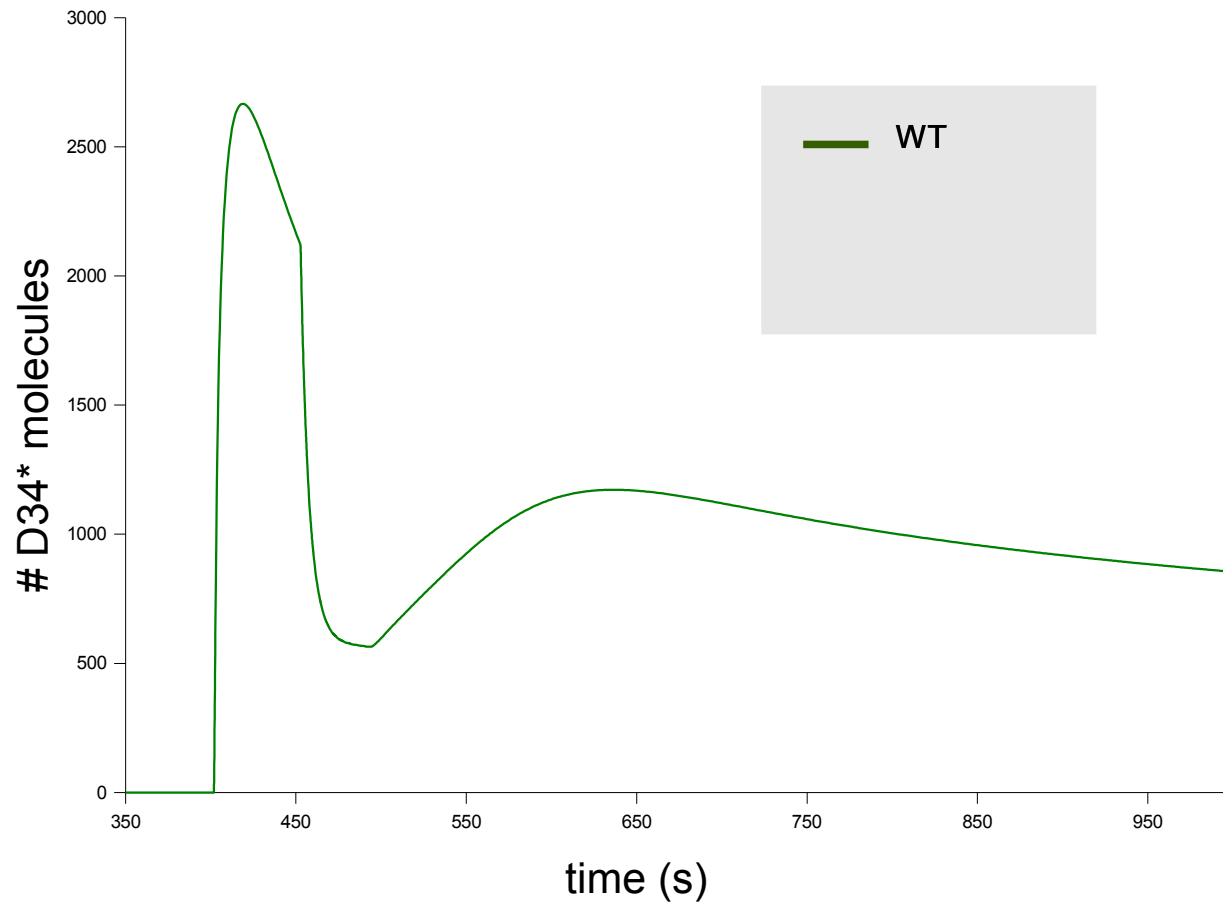


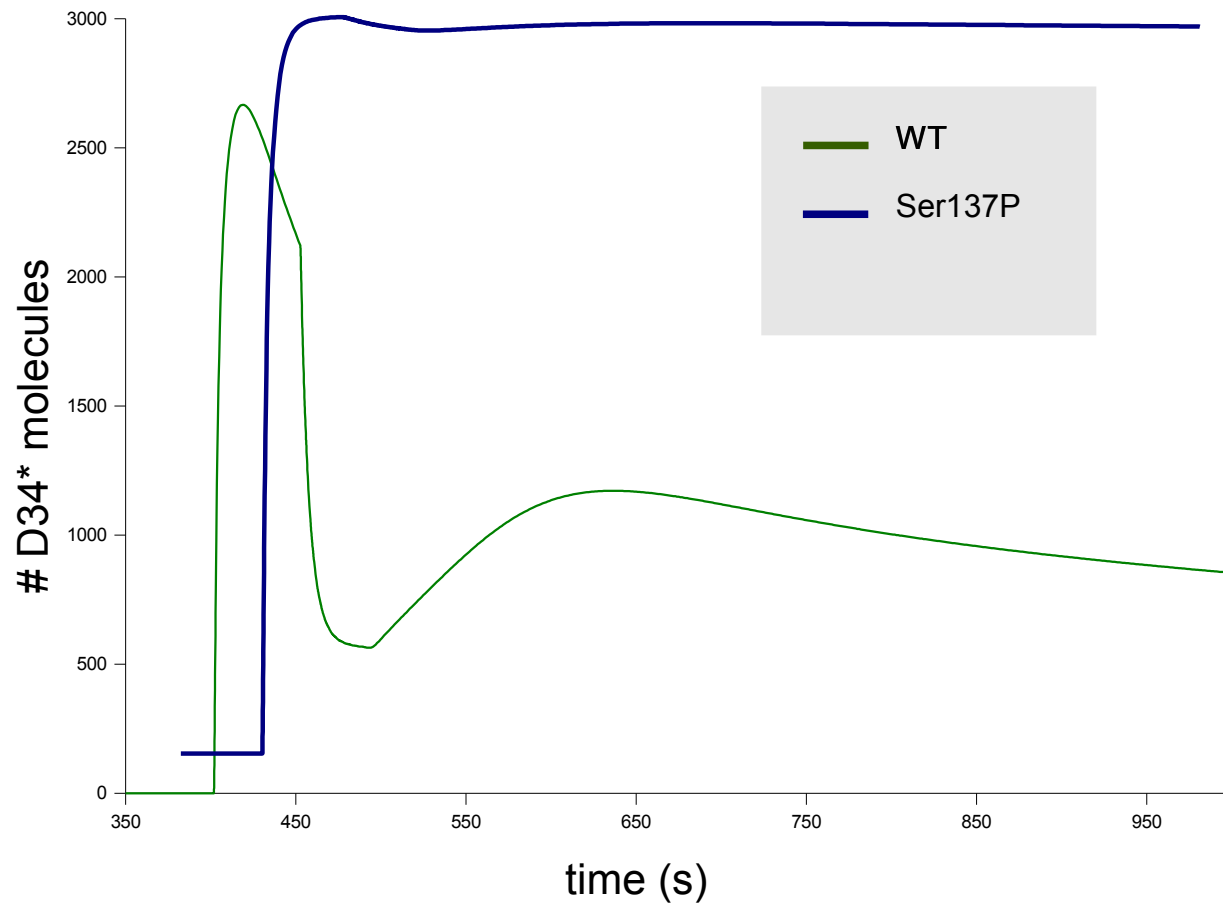


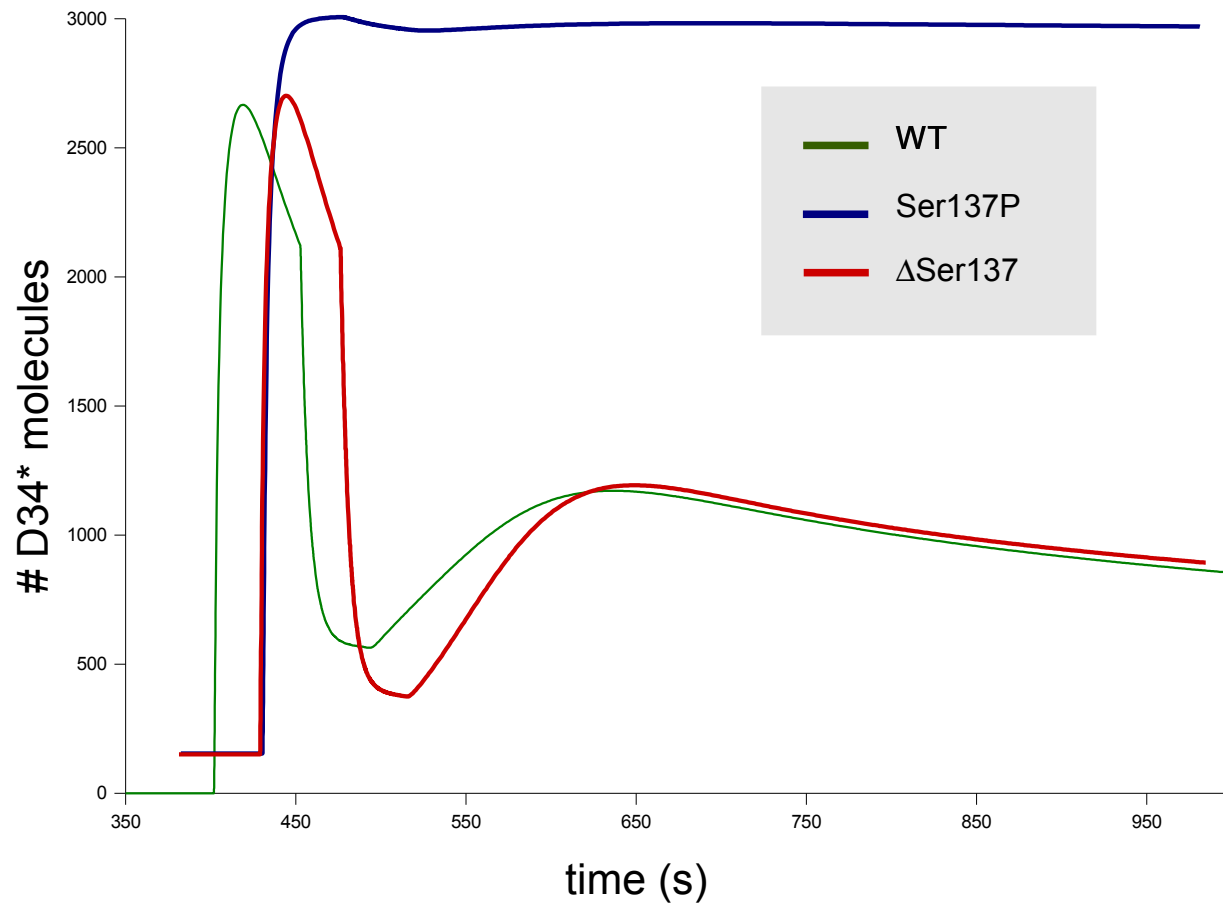


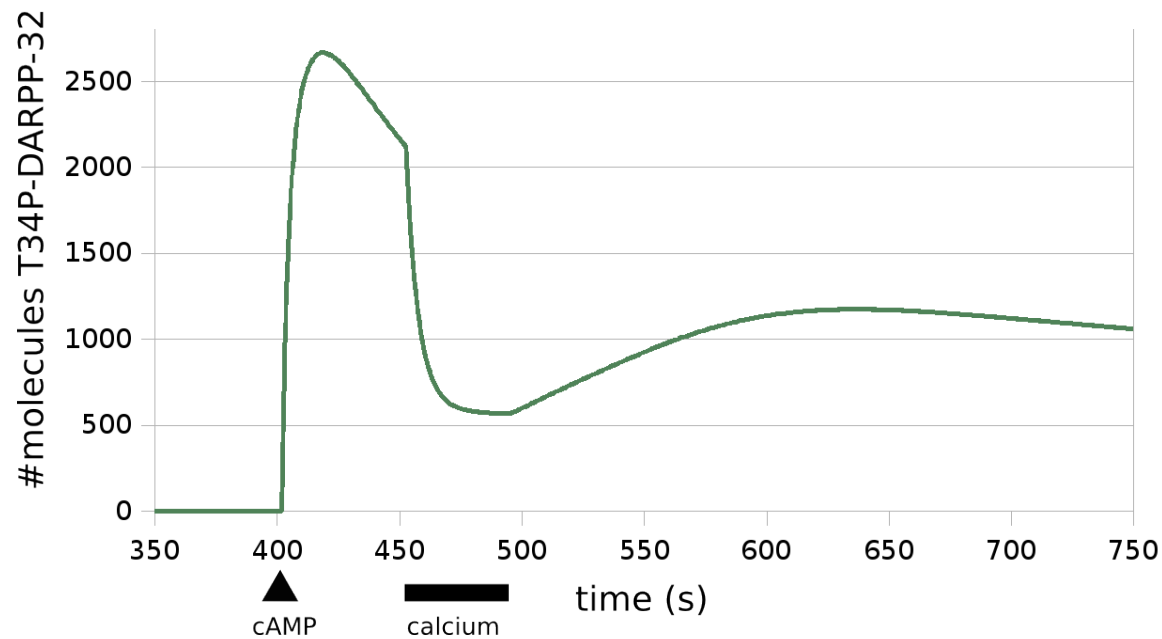
Takahashi et al (2005) Increased activity of cyclin-dependent kinase 5 leads to attenuation of cocaine-mediated dopamine signaling. PNAS 102: 1737-1742.

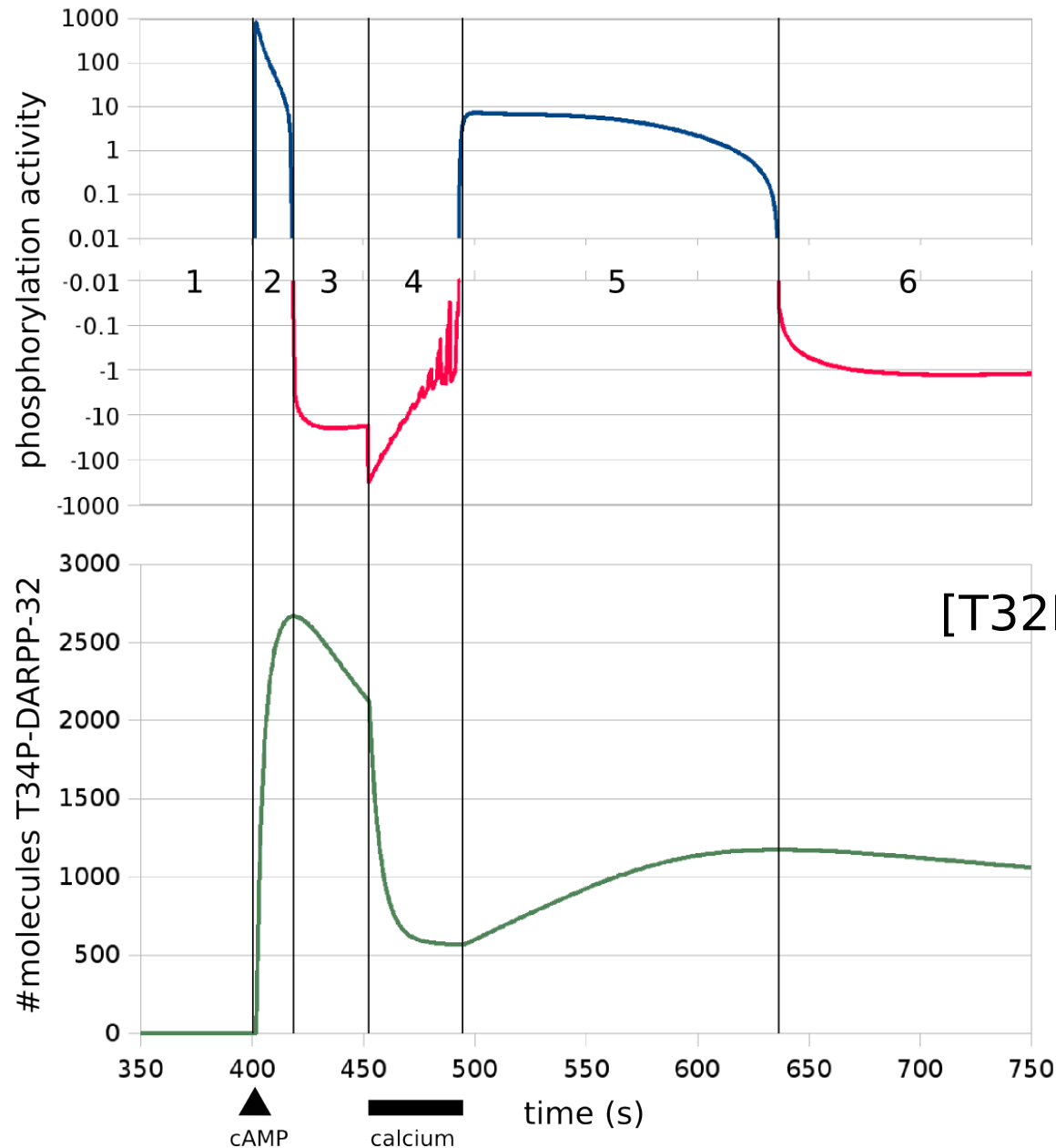






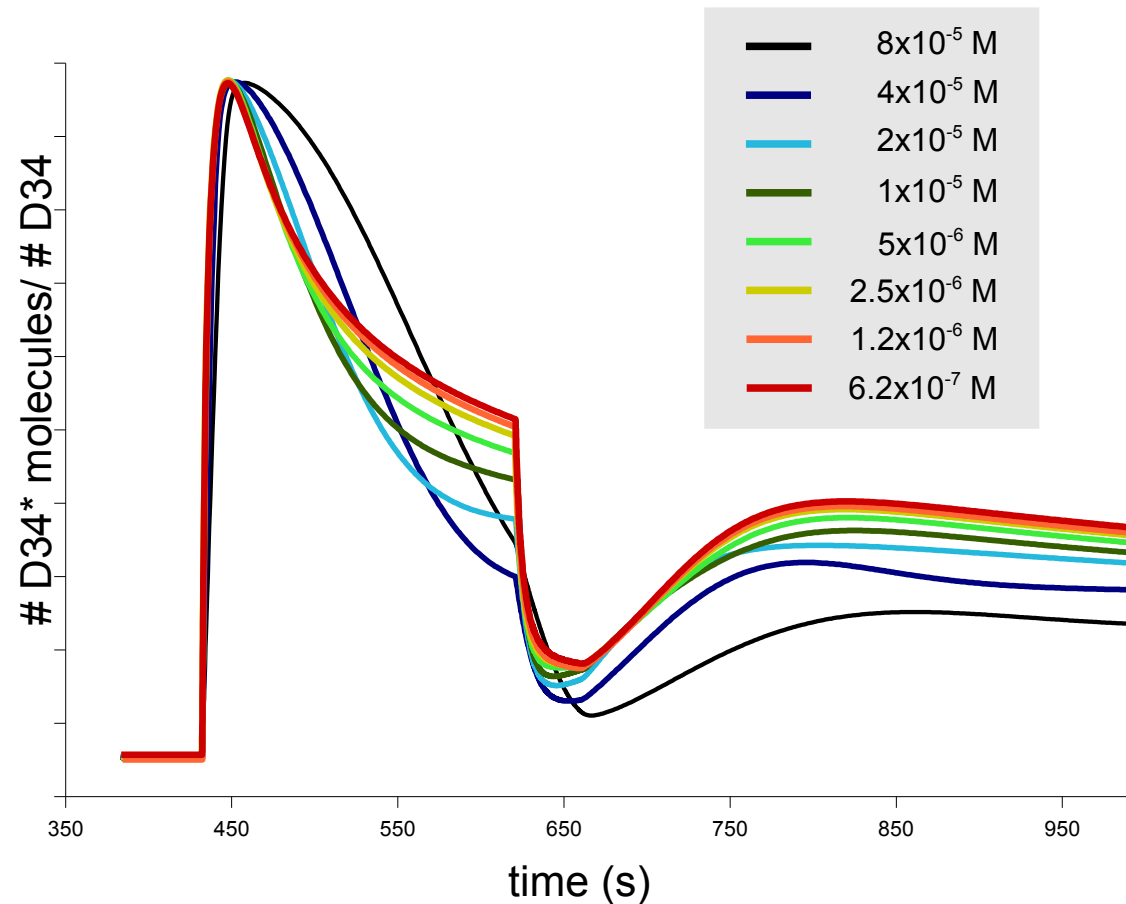


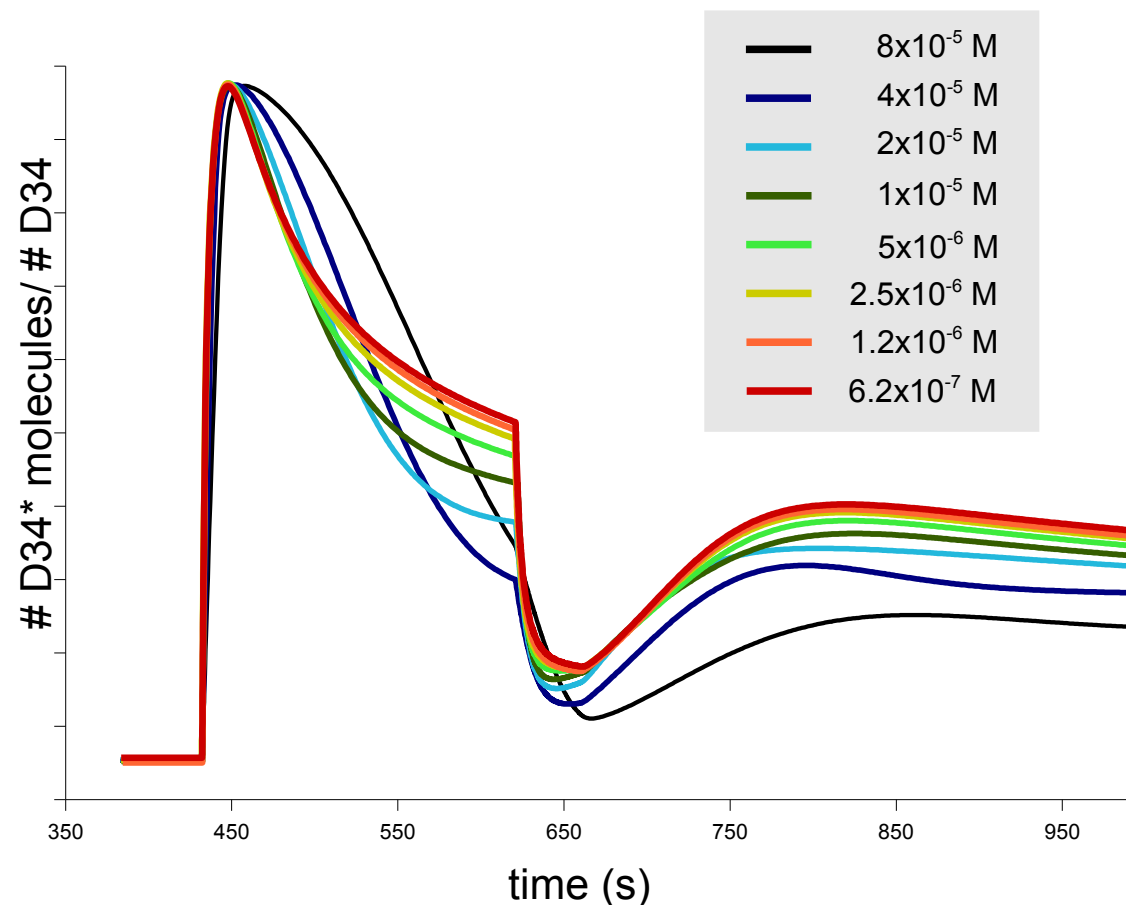




$$[T32P] = \int (\sum[kin] - \sum[ppase]) dt$$

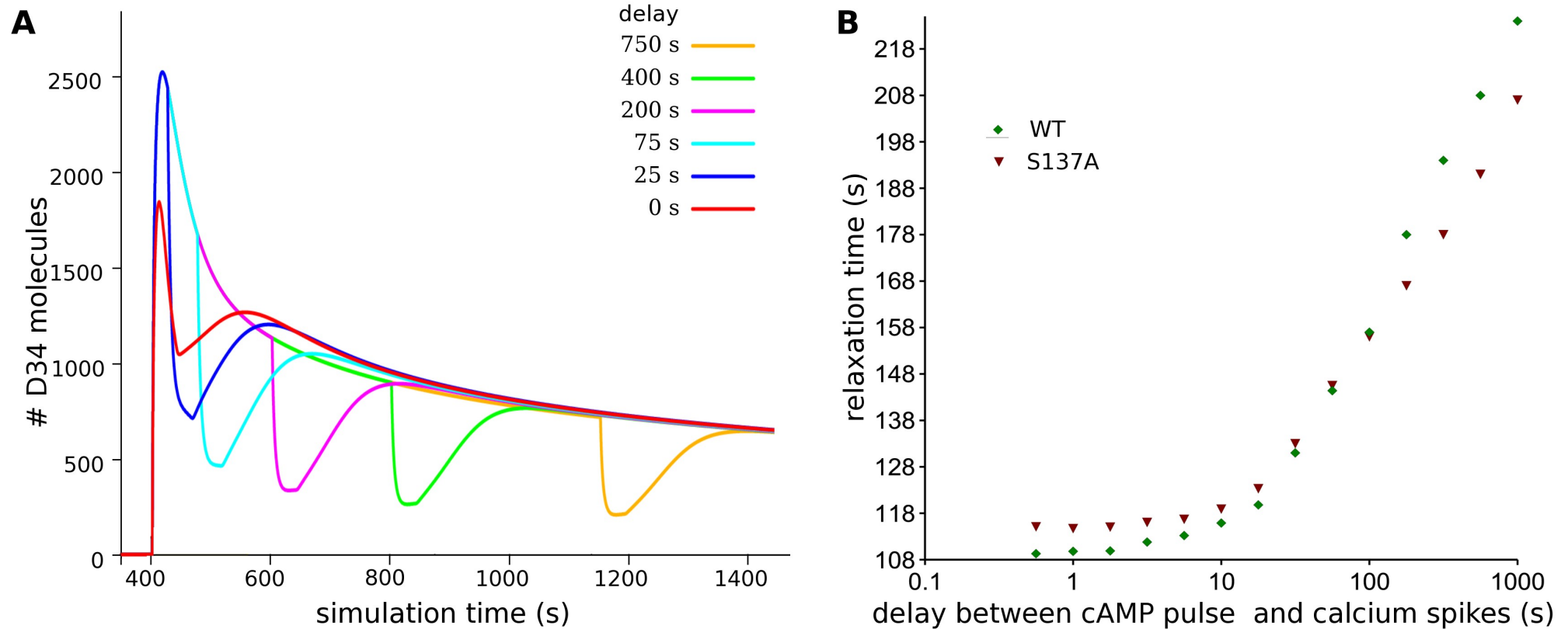






Heterozygous DARPP-32 +/- do not display phenotypes





- E-cell developers
(Institute of Advanced Biosciences, Tsuruoka)

Kouichi Takahashi

Kazunari Kaizu

Gabor Bereczki

- Collaborators
(INSERM U536, Paris)

Jean-Antoine Girault

Denis Herve



Eric Fernandez



Renaud Schiappa



Lu Li



■ Pros

- Scale well-adapted to most available quantitative biochemical information
- Can easily be extended and/or modularised
- Extensive theoretical and computational toolkit

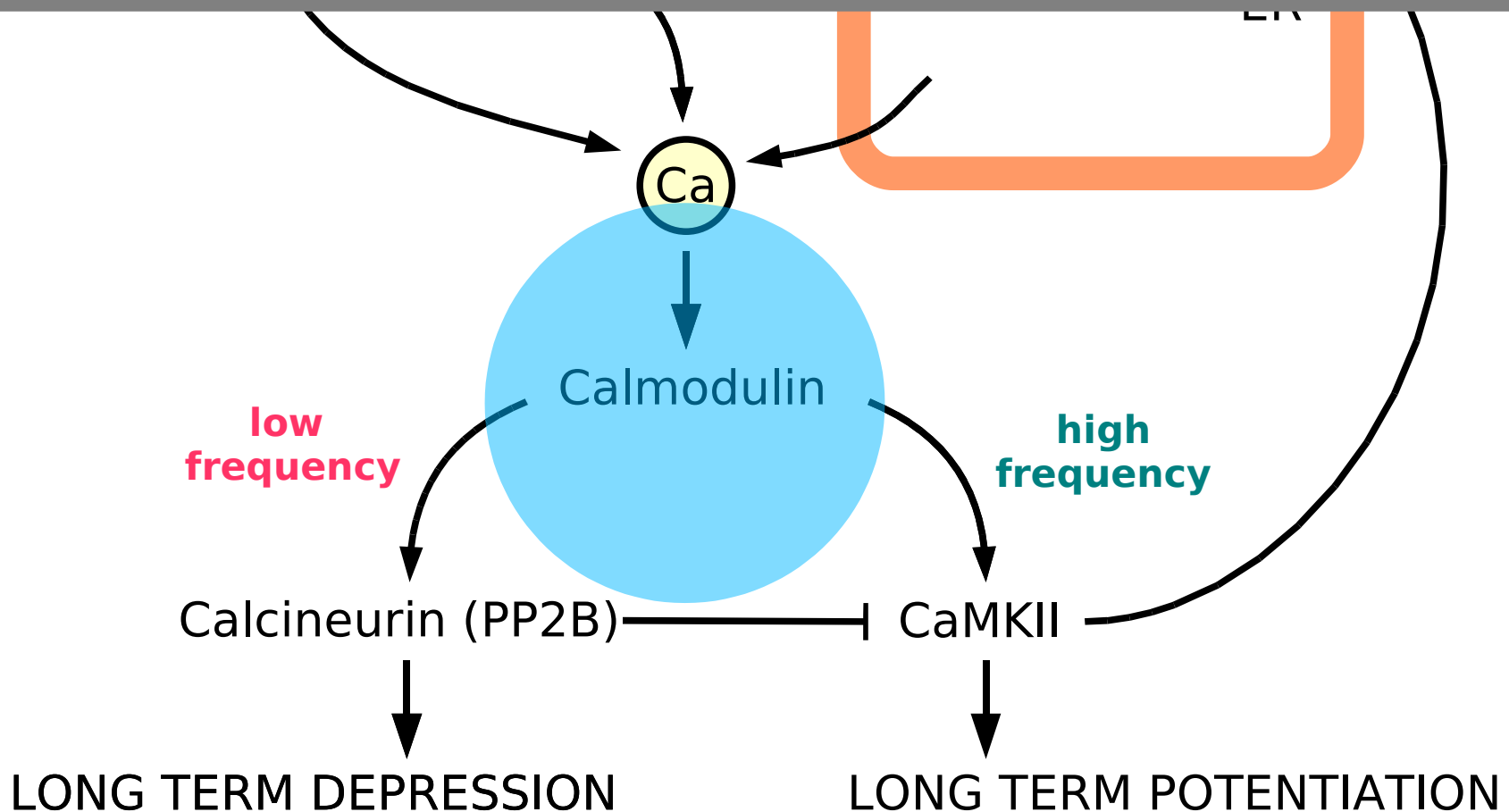
■ Cons

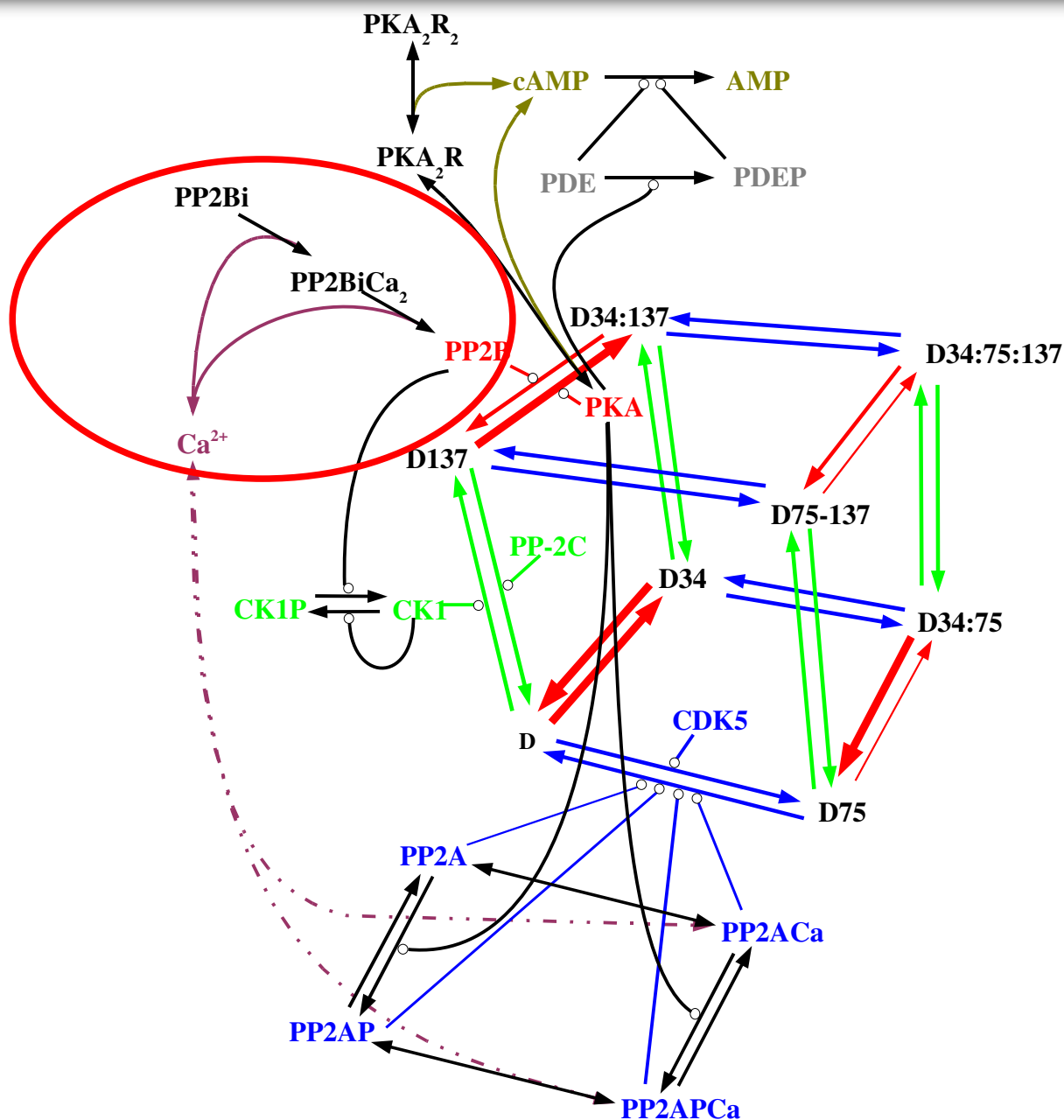
- Description of reactions is far from being realistic, and do not describe accurately the underlying mechanisms
- Sensitive to combinatorial explosion
- Dendritic spine is strongly anisotropic: Importance of distributions, gradients, crowding, scaffolding etc.

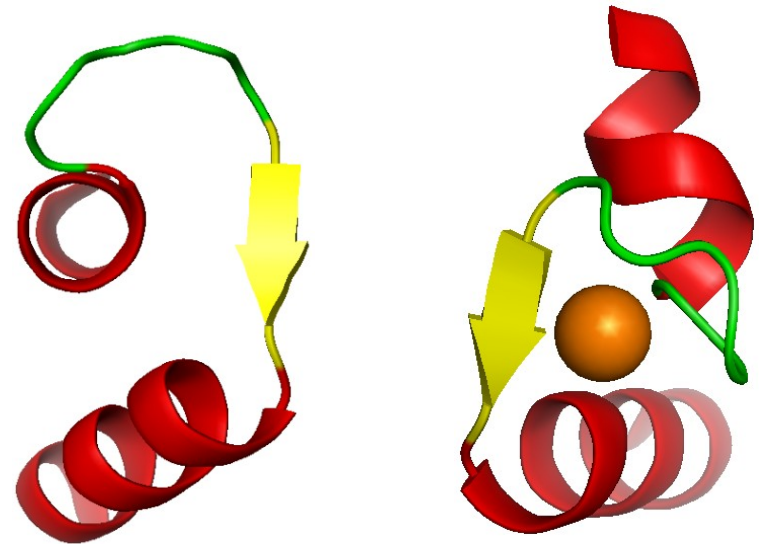


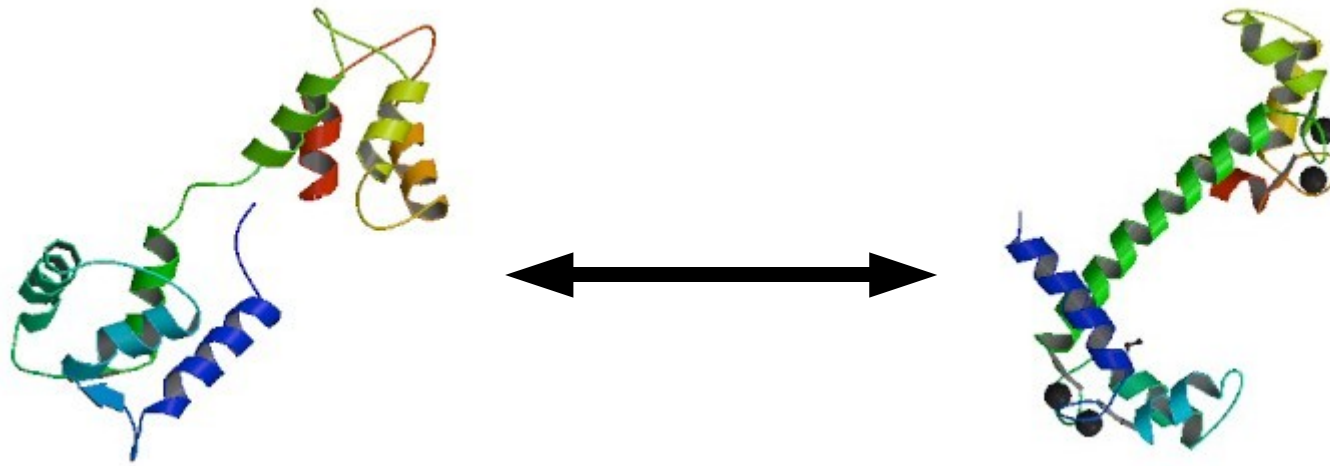


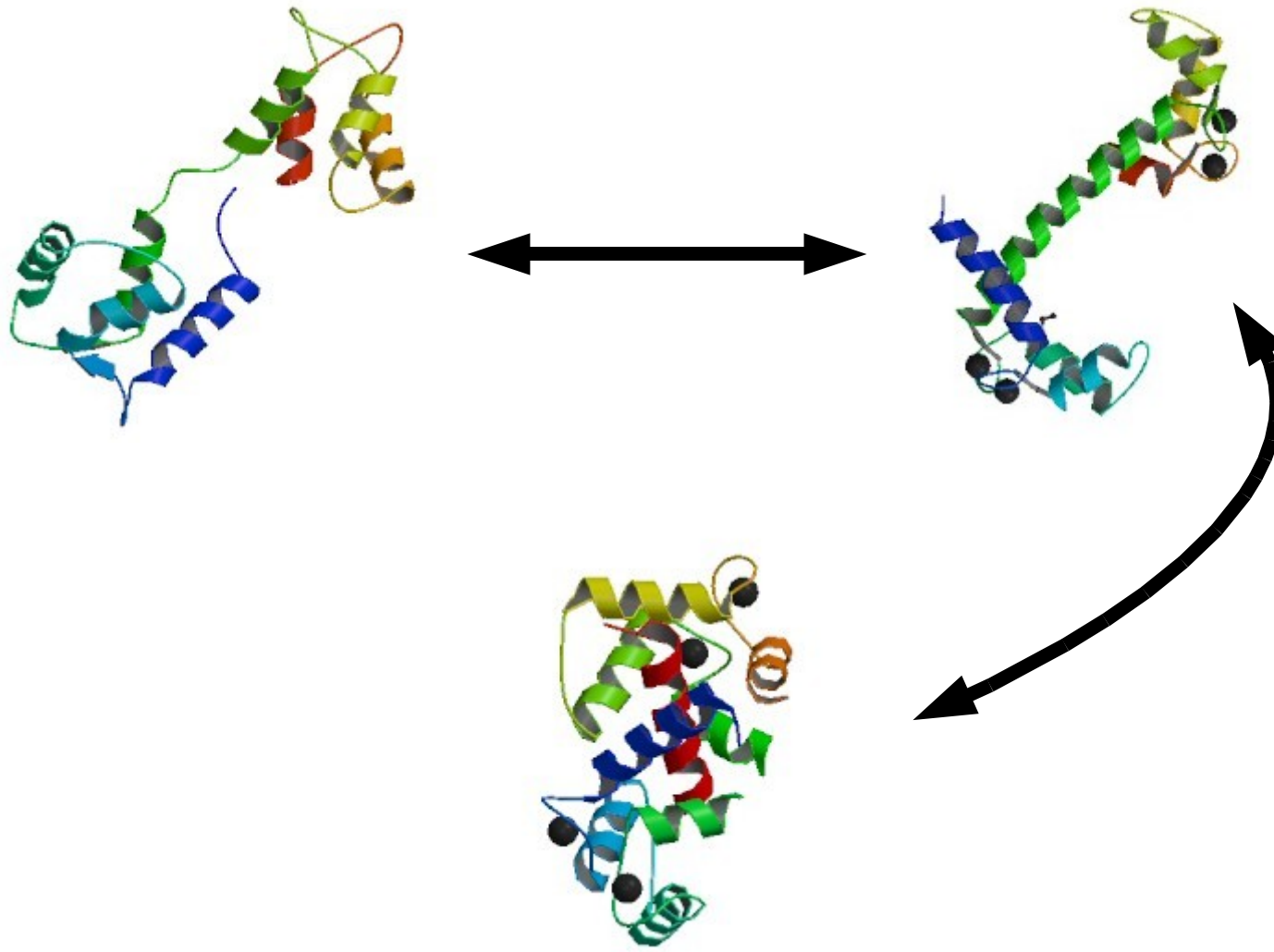
Stefan MI, Edelstein SJ, Le Novère N (2008) An allosteric model of calmodulin explains differential activation of PP2B and CaMKII. *Proceedings of the National Academy of Sciences USA*, 105:10768-10773
Stefan MI, Edelstein SJ, Le Novère N. Computing phenomenologic Adair-Klotz constants from microscopic MWC parameters. *submitted*





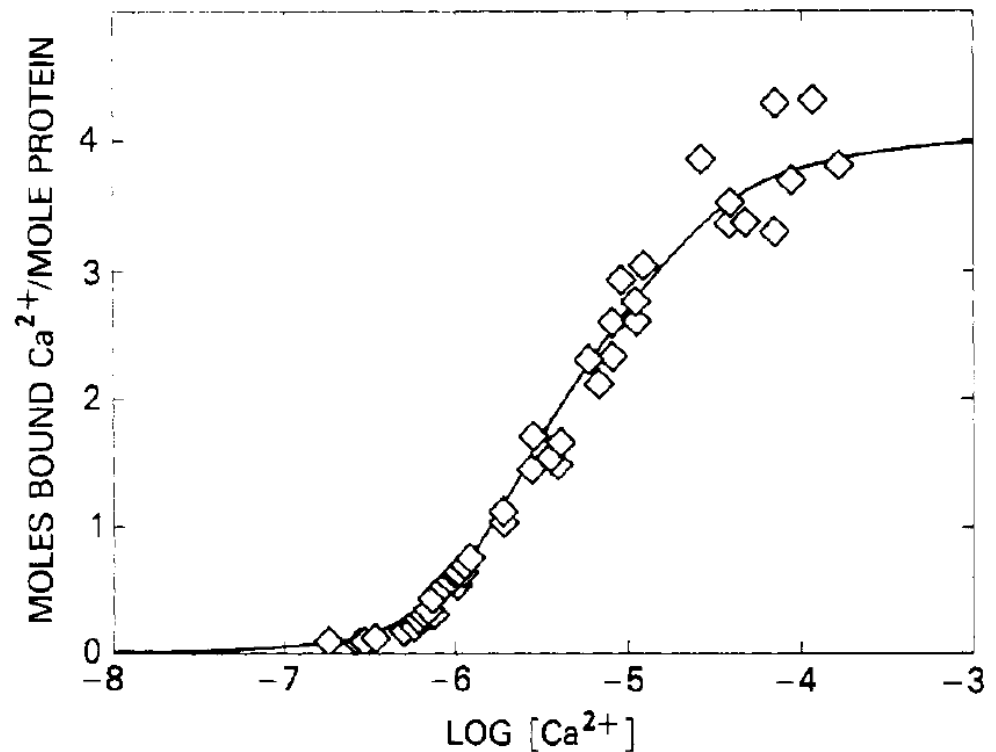






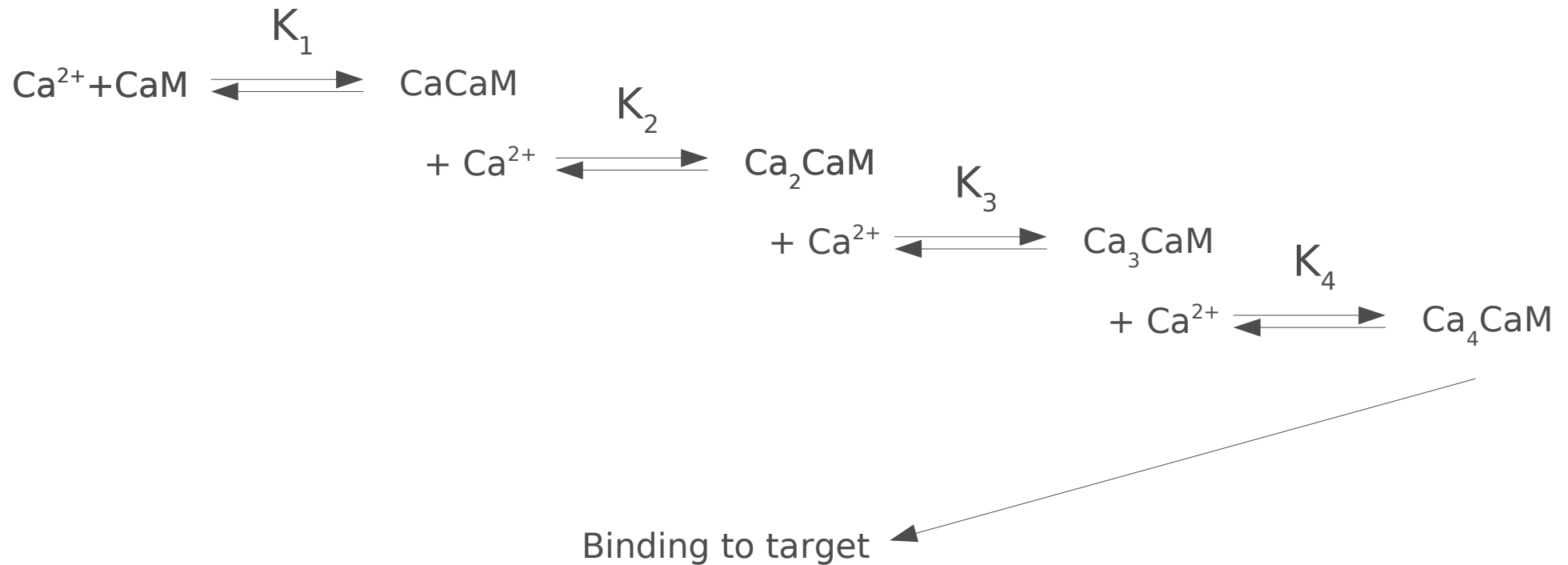
$$\bar{Y} = \frac{K_1[\text{Ca}] + 2K_1K_2[\text{Ca}]^2 + 3K_1K_2K_3[\text{Ca}]^3 + 4K_1K_2K_3K_4[\text{Ca}]^4}{1 + K_1[\text{Ca}] + K_1K_2[\text{Ca}]^2 + K_1K_2K_3[\text{Ca}]^3 + K_1K_2K_3K_4[\text{Ca}]^4}$$

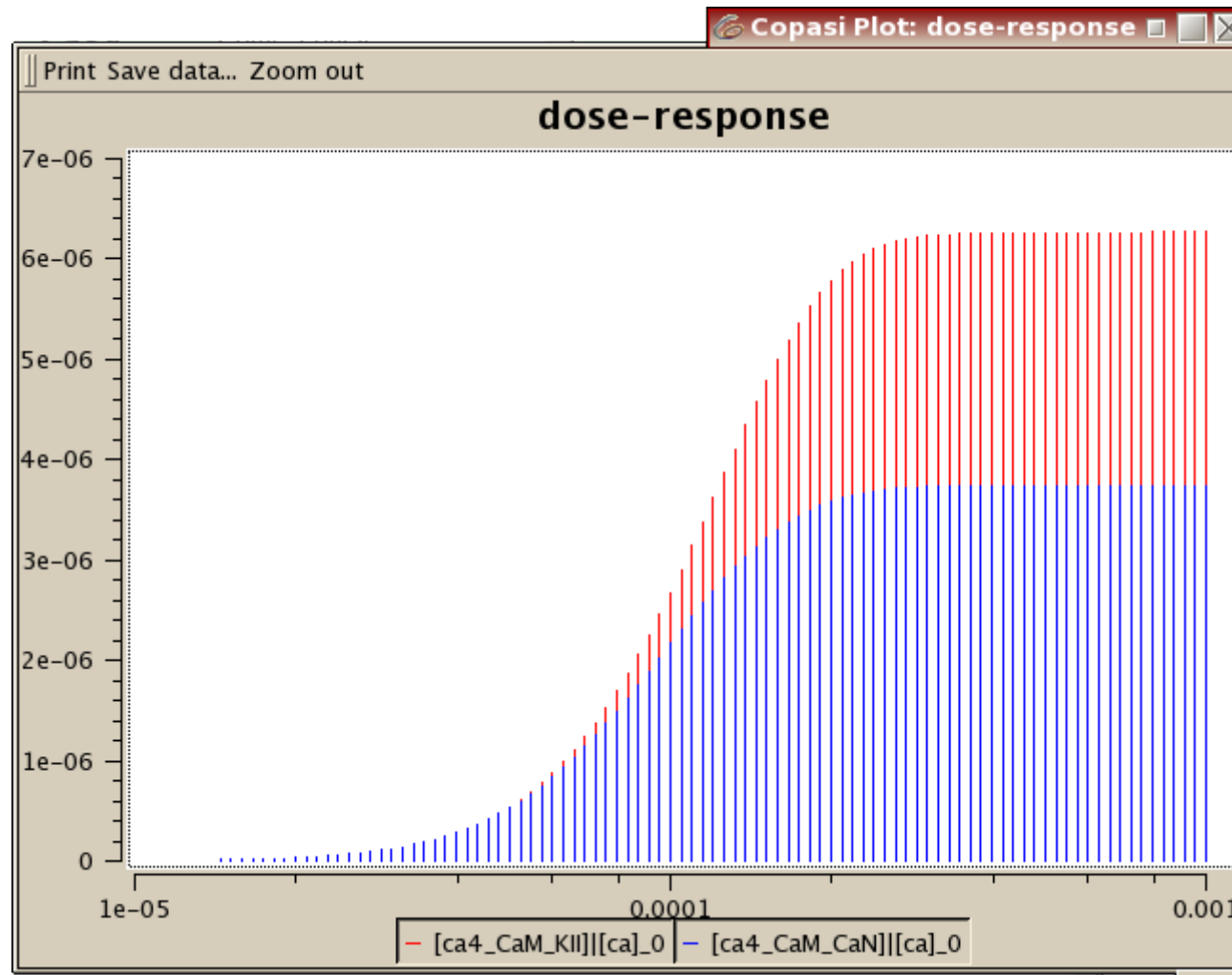
Adair (1925)
J Biol Chem, 63: 529-545



Crouch and Klee (1980)
Biochemistry, 19: 3692-3698







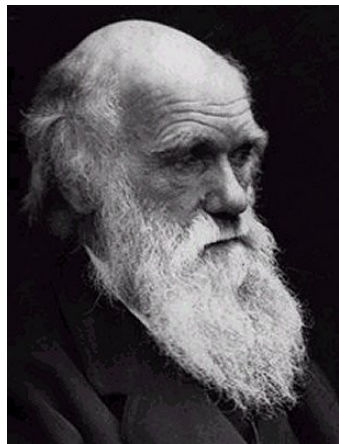
- Calmodulin bound to three calcium activates calcineurin
 - Kincaid and Vaughan (1986). *PNAS*, 83: 1193-1197
- Calmodulin bound to two calcium can bind CaMKII
 - Shifman et al (2006). *PNAS*, 103: 13968-13973
- Calmodulin affinity for calcium increases once bound to CaMKII
 - Shifman et al (2006) [but many previous reports on other targets: e.g. Burger et al (1983). *JBC*, 258: 14733-14739 ; Olwin et (1984). *JBC* 259: 10949-10955]
- Calcium activates both LTP and LTD through calmodulin
 - Lisman (1989) *PNAS*, 86: 9574-9578
 - High $[Ca^{2+}]$ (high freq) $\cong$ CaMKII ; Low $[Ca^{2+}]$ (low freq) $\cong$ Calcineurin



- Induction = BAD
(intelligent design, Lamarck's first law, antibody moulding, directed axonal growth, induced-fit ...)
- Physicaly non-sensical. Calcium has no inertia. Calcium cannot “trigger” a conformational change

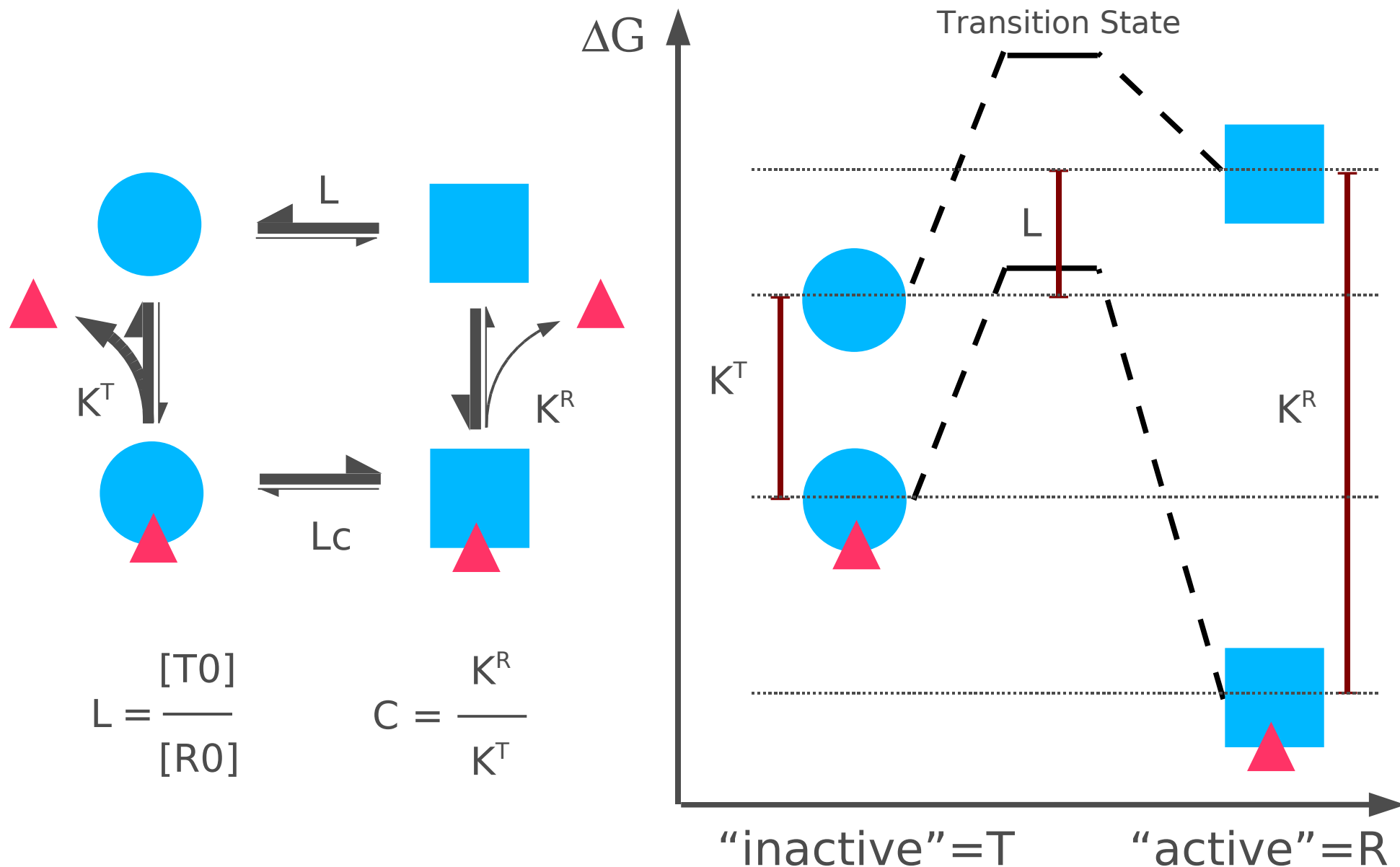


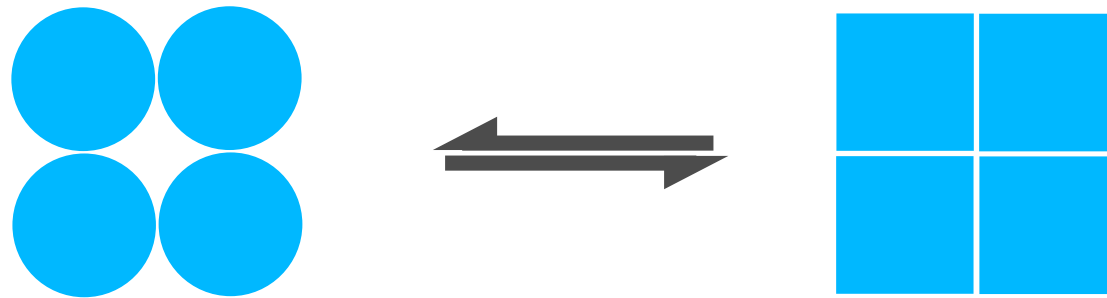
- **Induction = BAD**
(intelligent design, Lamarck's first law, antibody moulding, directed axonal growth, induced-fit ...)
- Physically non-sensical. Calcium has no inertia. Calcium cannot “trigger” a conformational change
- **Selection = GOOD**
(natural selection, clonal selection, synapse stabilisation, conformation stabilisation ...)

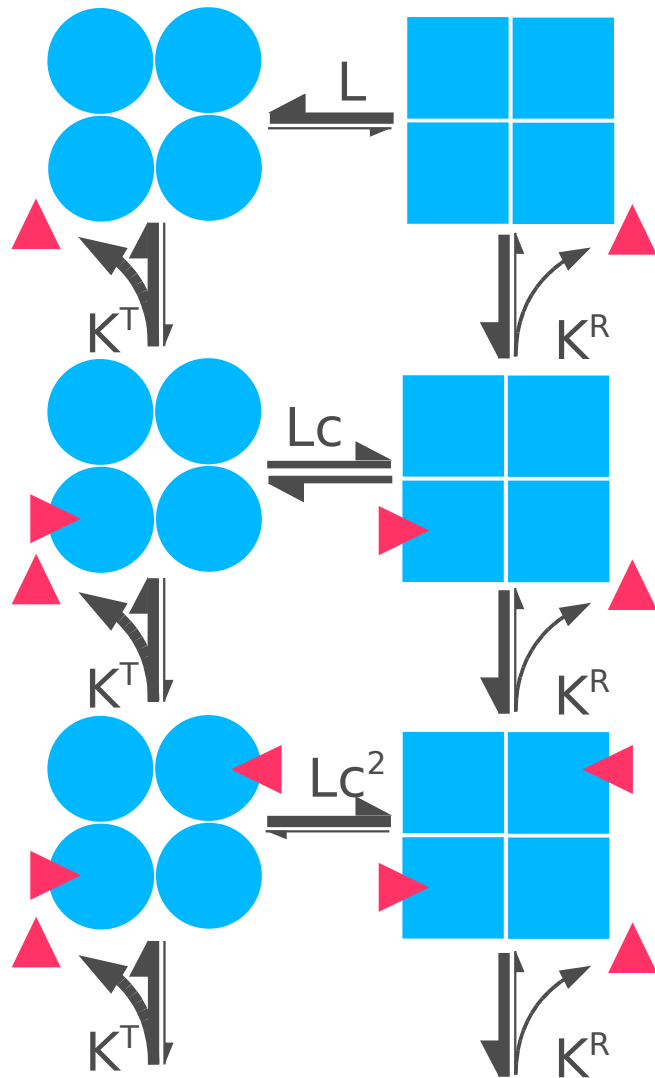


- ➔ Monod, Wyman, Changeux (1965). On the nature of allosteric transitions: a plausible model.
J Mol Biol, 12: 88-118



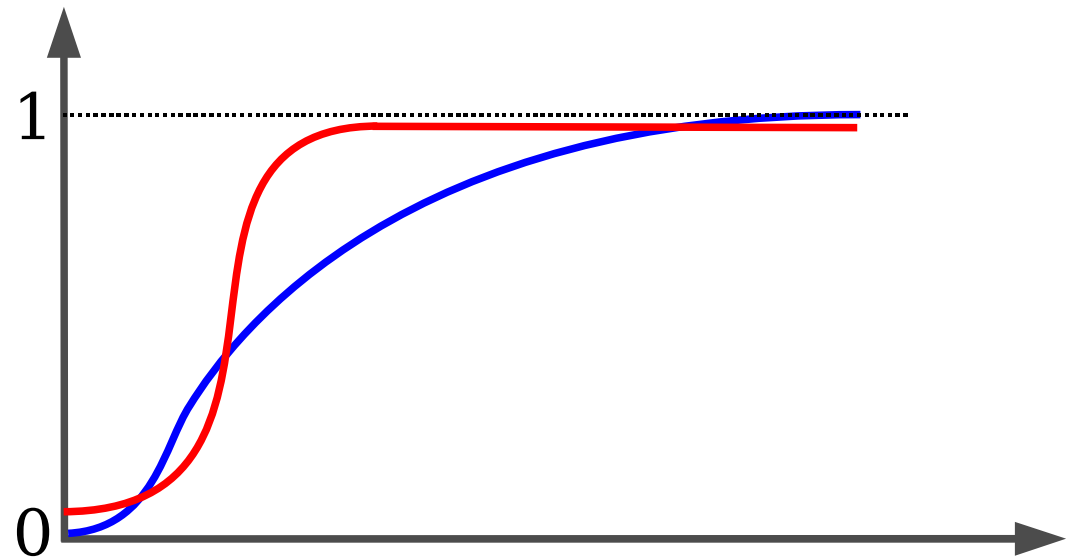






$$\bar{Y} = \frac{Lc\alpha(1 + c\alpha)^{n-1} + \alpha(1 + \alpha)^{n-1}}{L(1 + c\alpha)^n + (1 + \alpha)^n}$$

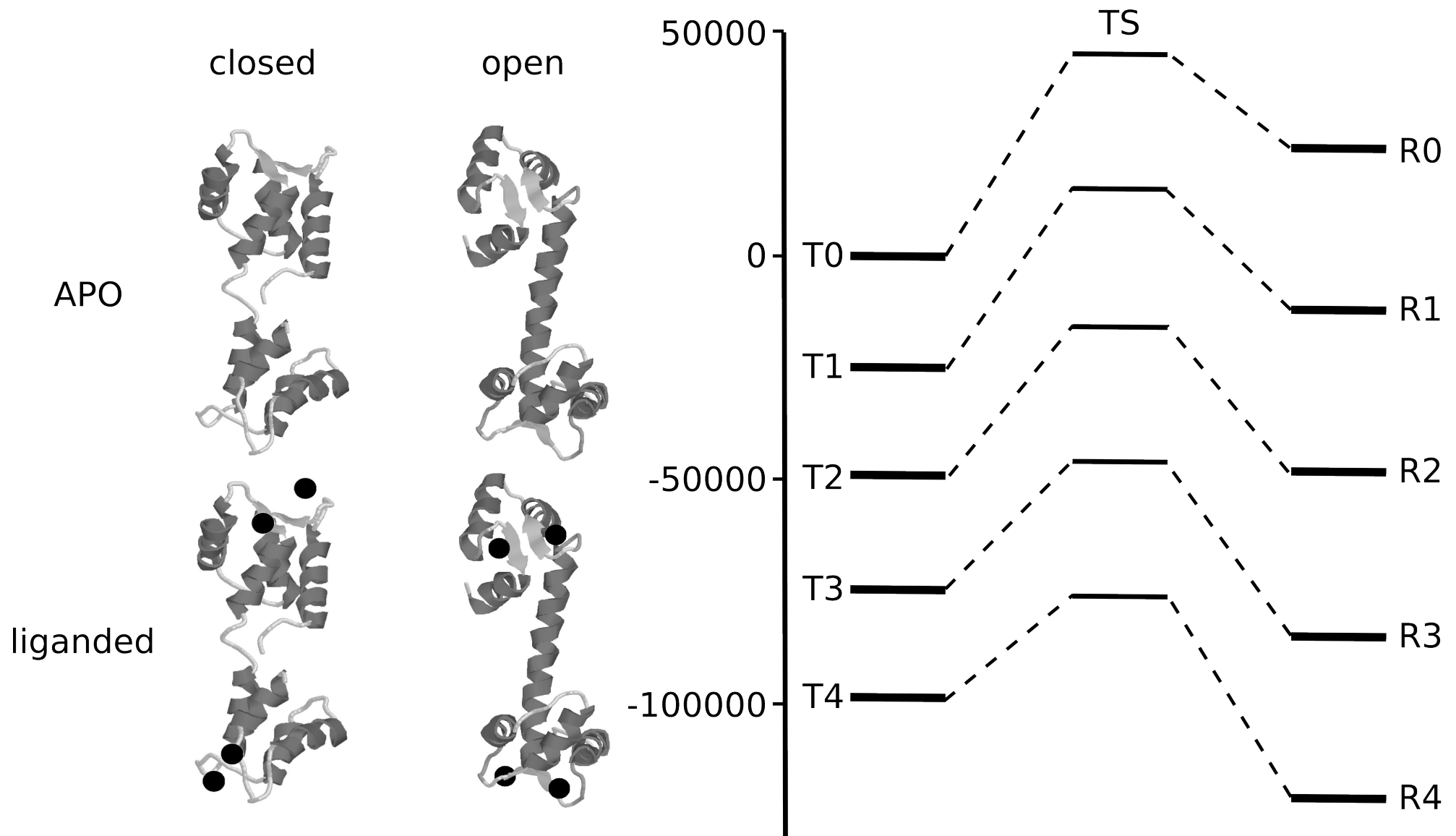
$$\bar{R} = \frac{(1 + \alpha)^n}{L(1 + c\alpha)^n + (1 + \alpha)^n}$$

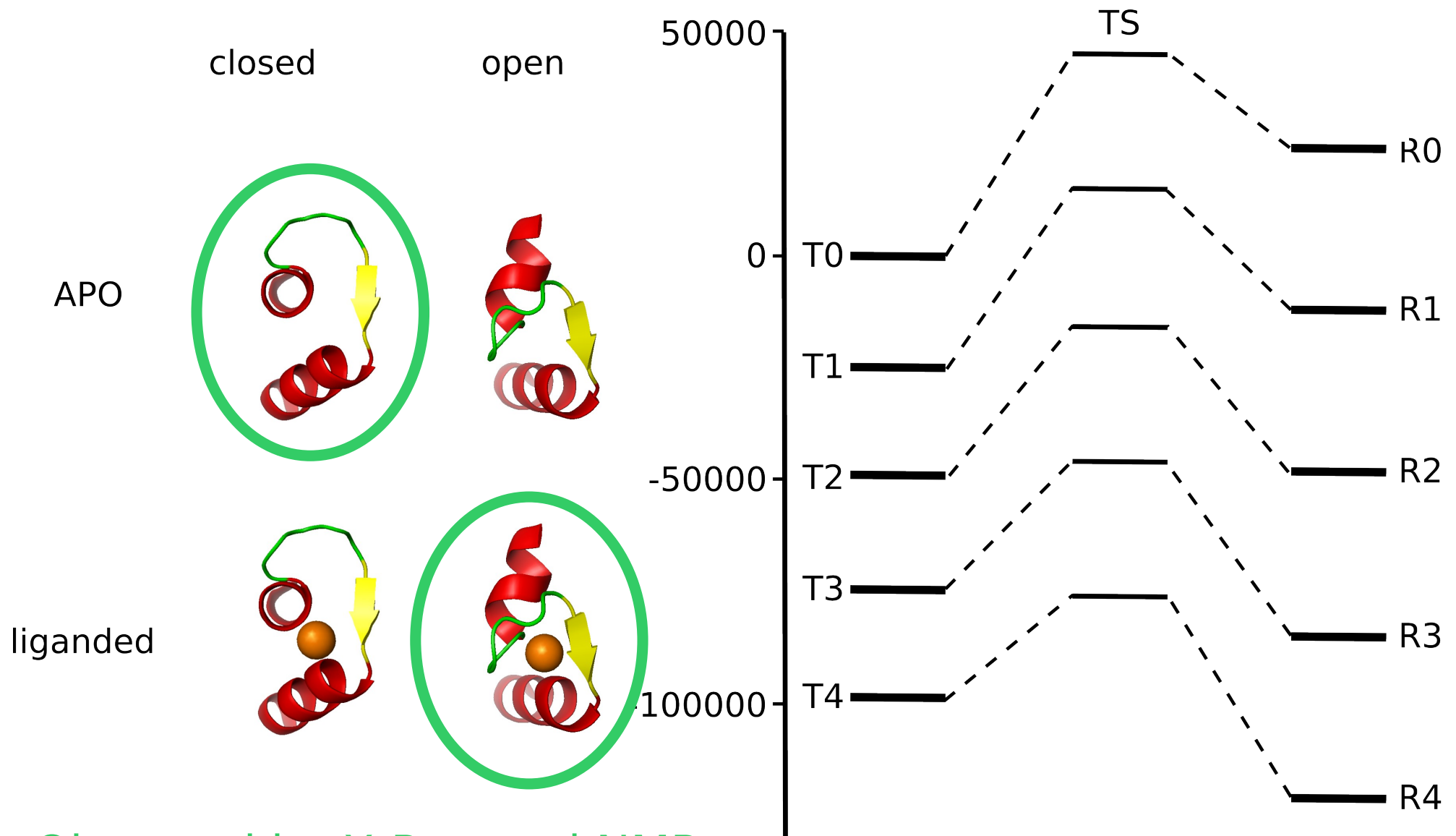


$$\bar{Y} = \frac{1}{n} \frac{\sum_i \left(\alpha_i \prod_{j \neq i} (1 + \alpha_j) \right) + L \prod_k \left(\frac{1 + e_k \gamma_k}{1 + \gamma_k} \right) \sum_i \left(c_i \alpha_i \prod_{j \neq i} (1 + c_j \alpha_j) \right)}{\prod_i (1 + \alpha_i) + L \prod_k \left(\frac{1 + e_k \gamma_k}{1 + \gamma_k} \right) \prod_i (1 + c_i \alpha_i)}$$

- $\alpha_i = [\text{ligand}] / K_{i,\text{lig}}^R$
- $\gamma_k = [\text{modulator}] / K_{k,\text{mod}}^R$
- $c_i = K_{i,\text{lig}}^R / K_{i,\text{lig}}^T$
- $e_k = K_{k,\text{mod}}^R / K_{k,\text{mod}}^T$

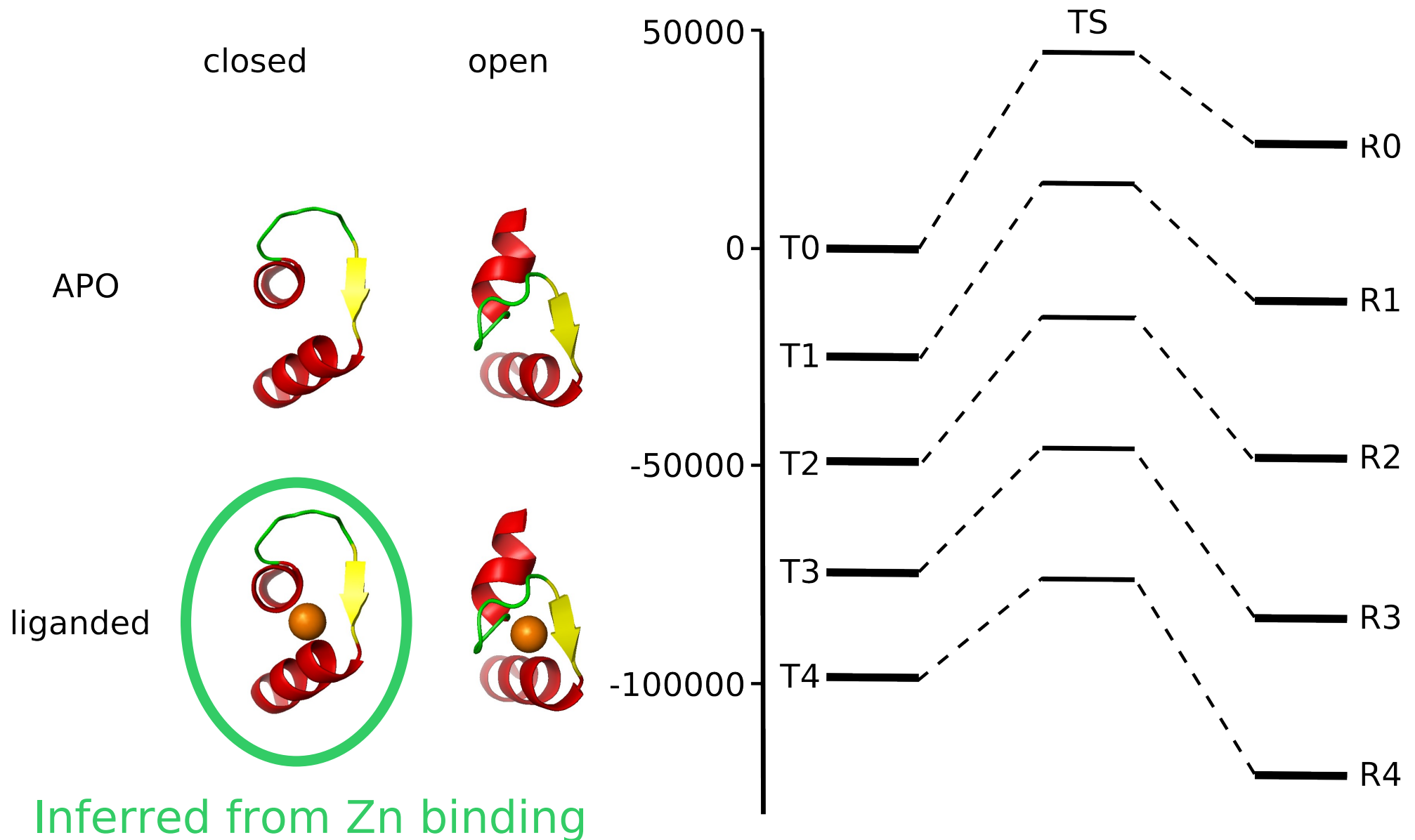


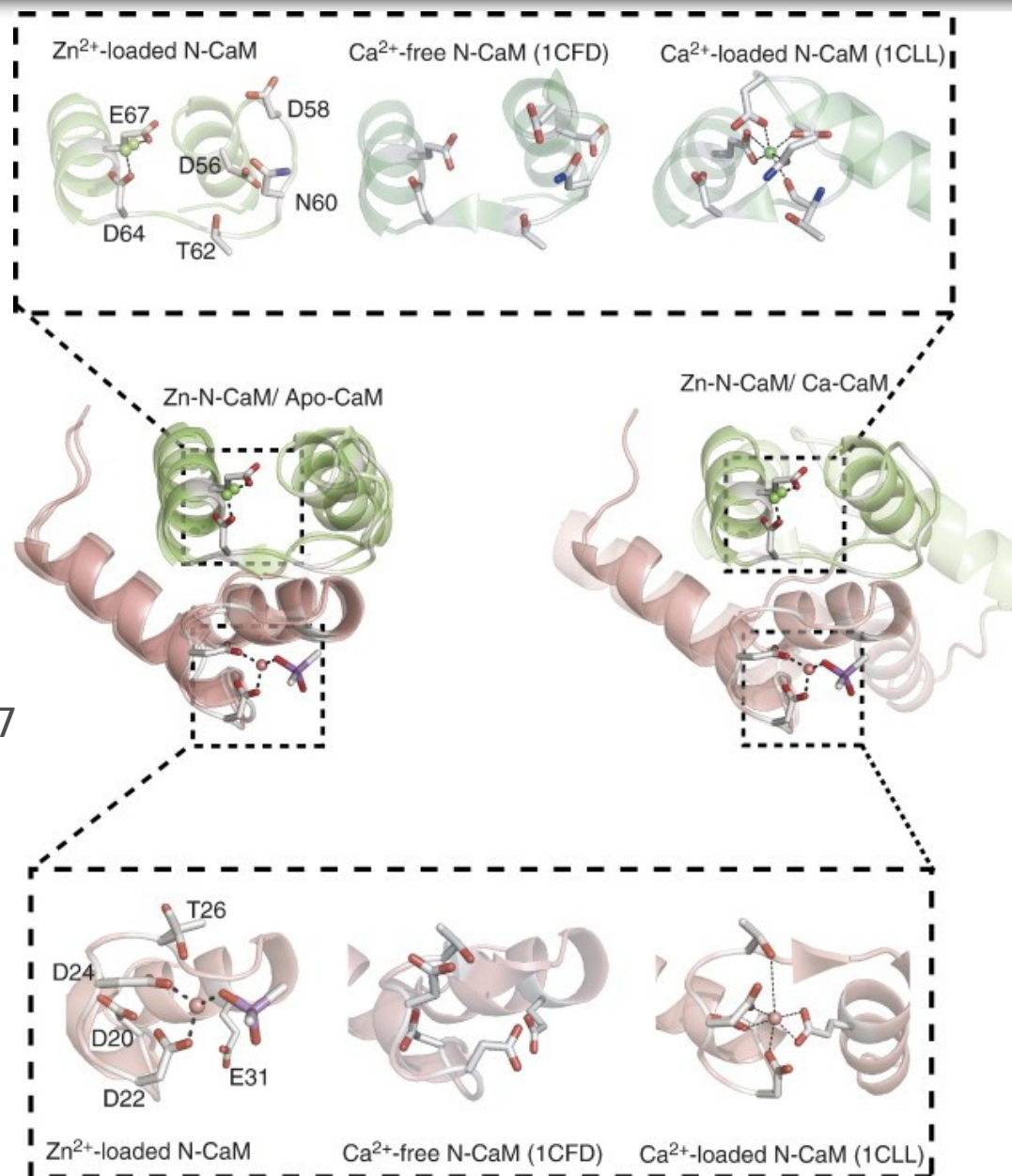




Observed by X-Ray and NMR

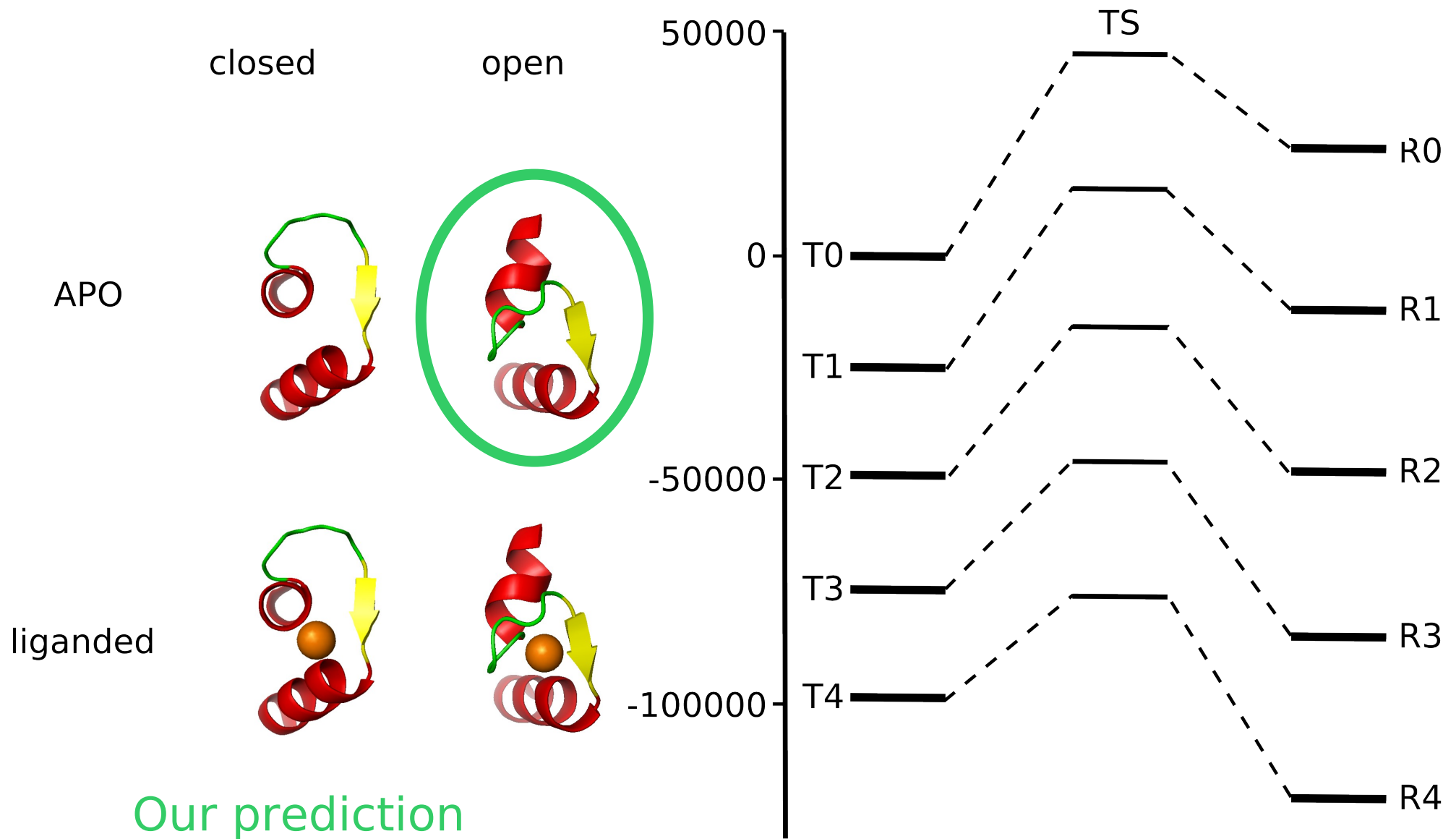


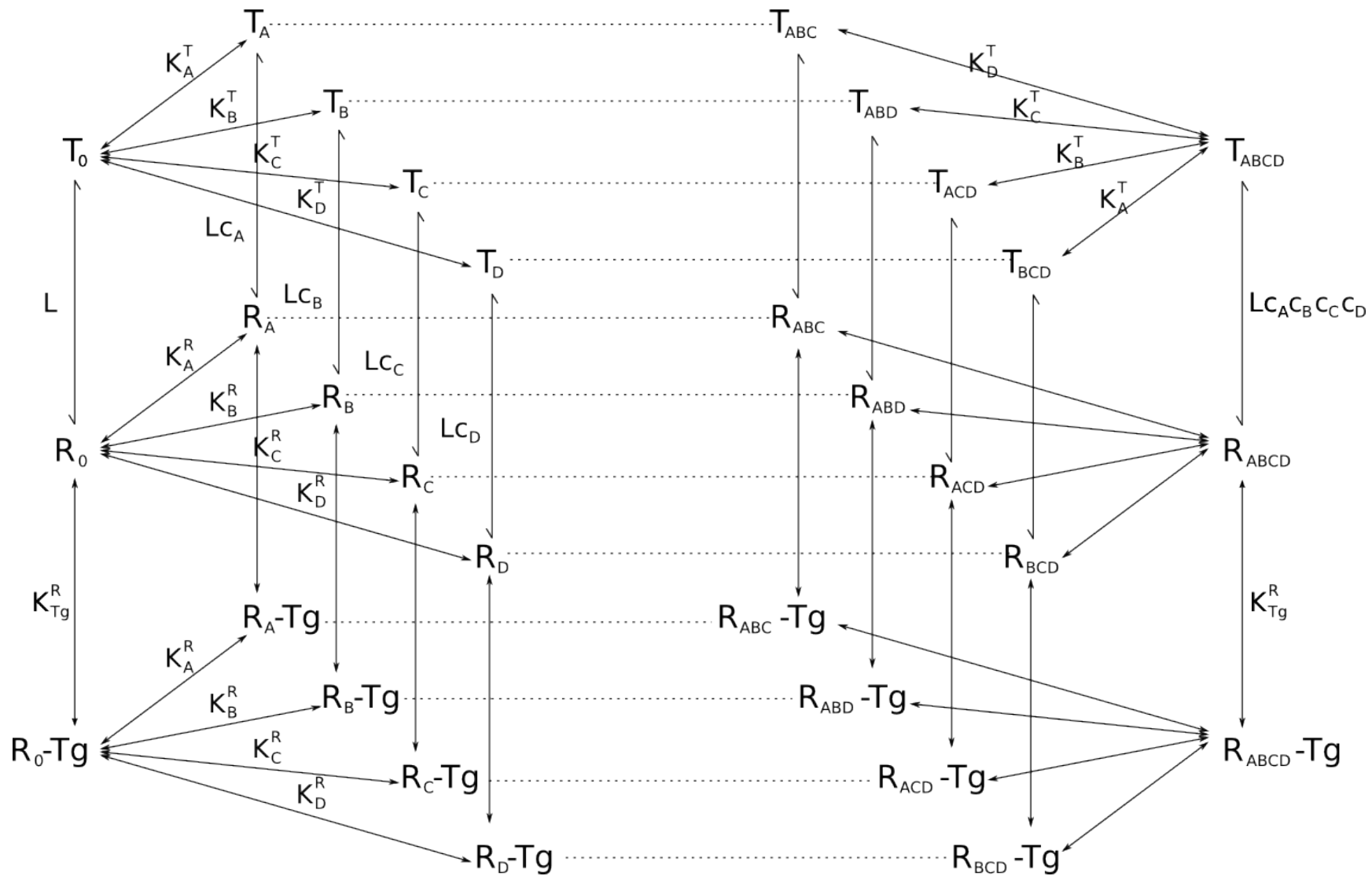




Warren et al. (2007).
J Mol Biol 374: 517-527







320 reactions



- Hypothesis for the whole model: free energy of conformational transition is evenly distributed: c is unique
- Additional simplification to determine L : affinities are identical

$$\bar{Y} = \frac{\alpha(1 + \alpha)^3 + L \left(\frac{1 + \gamma e}{1 + \gamma} \right) c \alpha (1 + c \alpha)^3}{(1 + \alpha)^4 + L \left(\frac{1 + \gamma e}{1 + \gamma} \right) (1 + c \alpha)^4}$$

- Determination of individual affinities:

$$\bar{Y} = 0.25 \frac{\sum_i \left(\alpha_i \prod_j (1 + \alpha_j) \right) + L \sum_i \left(c \alpha_i \prod_j (1 + c \alpha_j) \right)}{\prod_i (1 + \alpha_i) + L \prod_i (1 + c \alpha_i)}$$

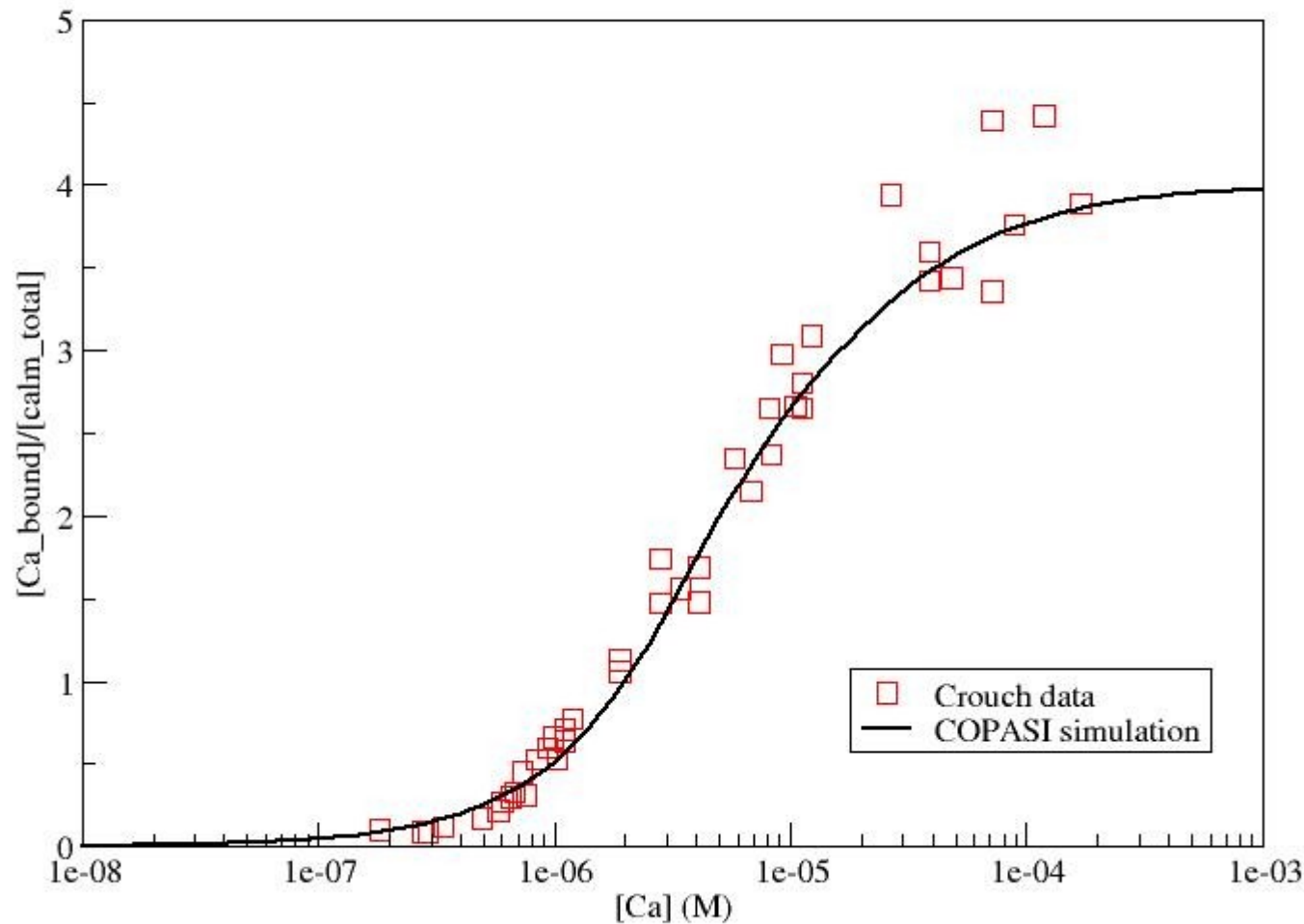


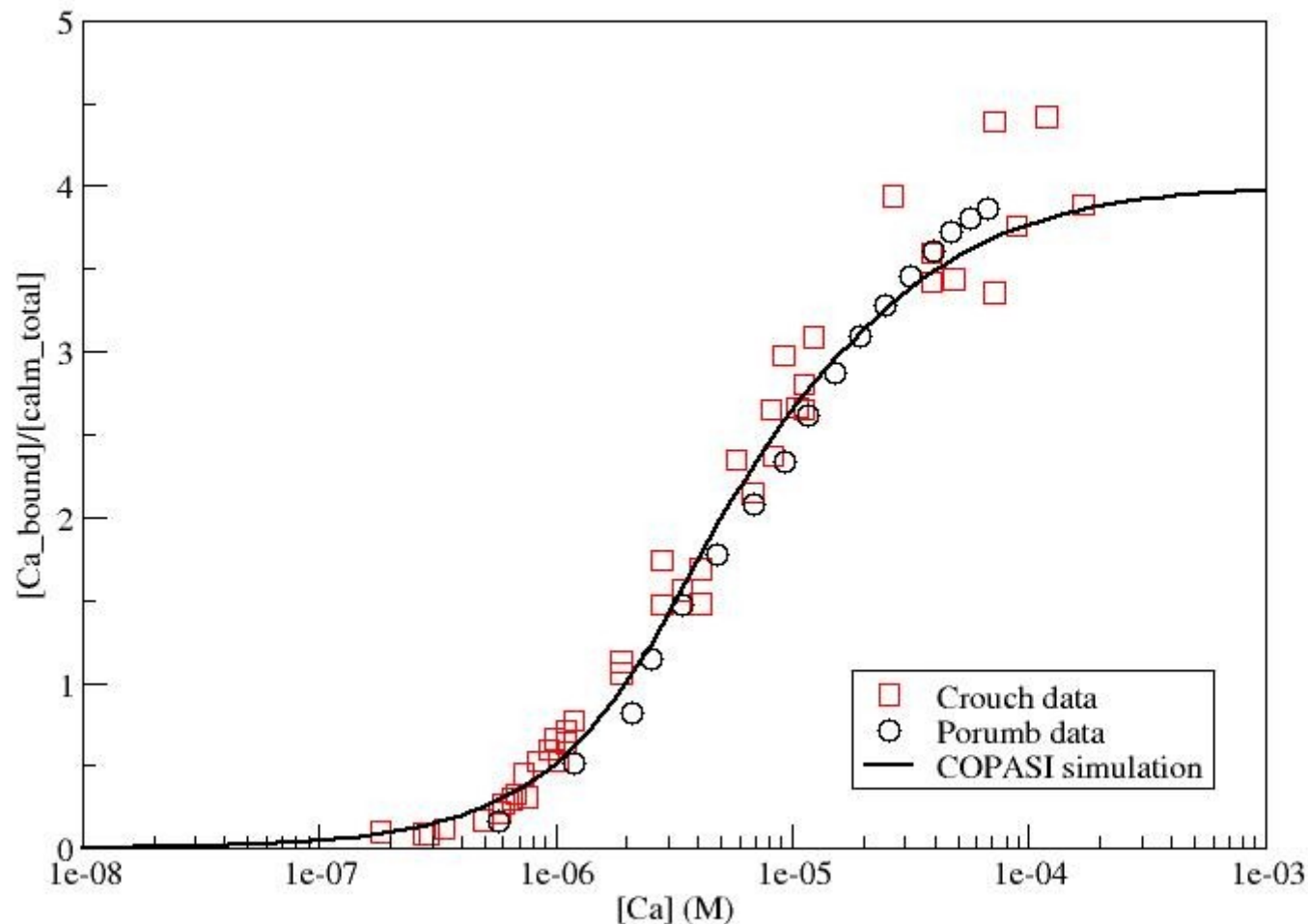
- Determination of c and L
 - Ca binding in presence of target: none, skMLCK, PhK5, CaATPase (Peersen et al (1997) *Prot Sci* 6: 794-807)
 - 100 000 parameter sets plus least-square
- Calcium dissociation constants
 - Complete CaM (Bayley et al (1996) *Prot Sci* 5: 1215-1228)
 - N and C term Mutants (Shifman et al (2006) *PNAS*, 103: 13968-13973)
 - R-only - skMLCK (Peersen et al (1997) *Prot Sci* 6: 794-807)
 - Systematic logarithmic sampling of the affinity space (coarse-grained, 50 values per affinity, then refined 66 values per affinity)
25 millions parameter sets

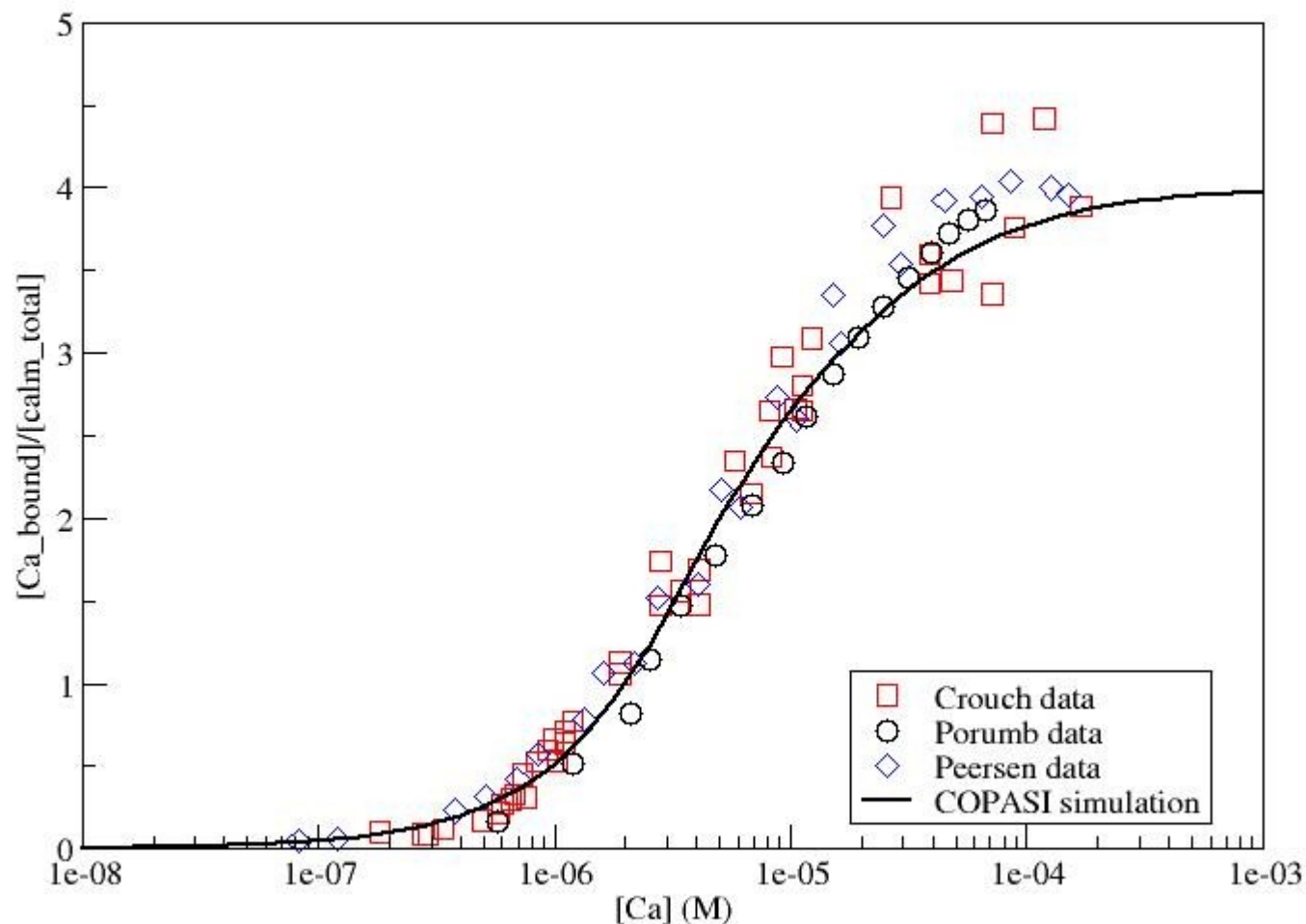
Simulations ran by COPASI (<http://www.copasi.org>)

fit done with SciLab (<http://www.scilab.org/>)







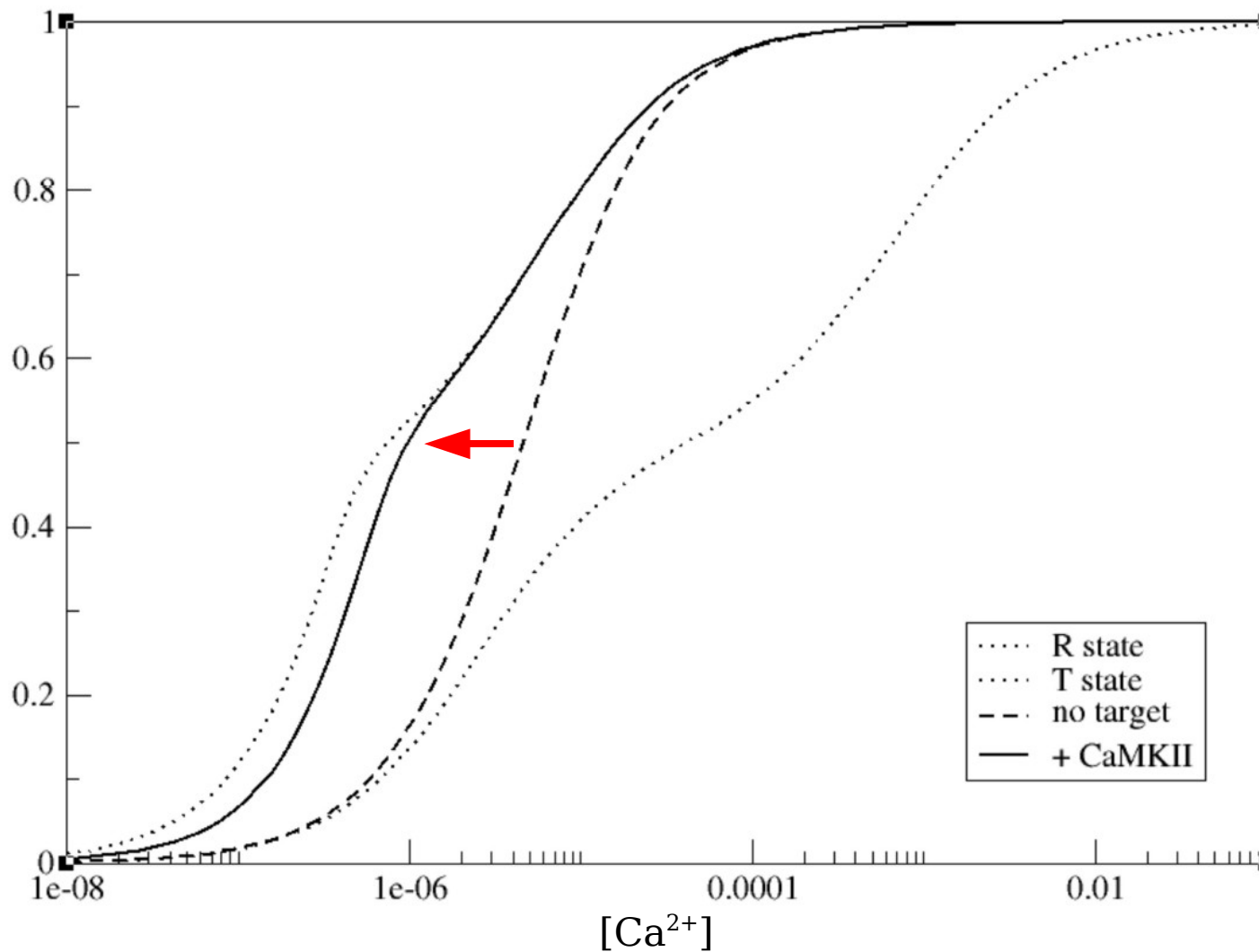


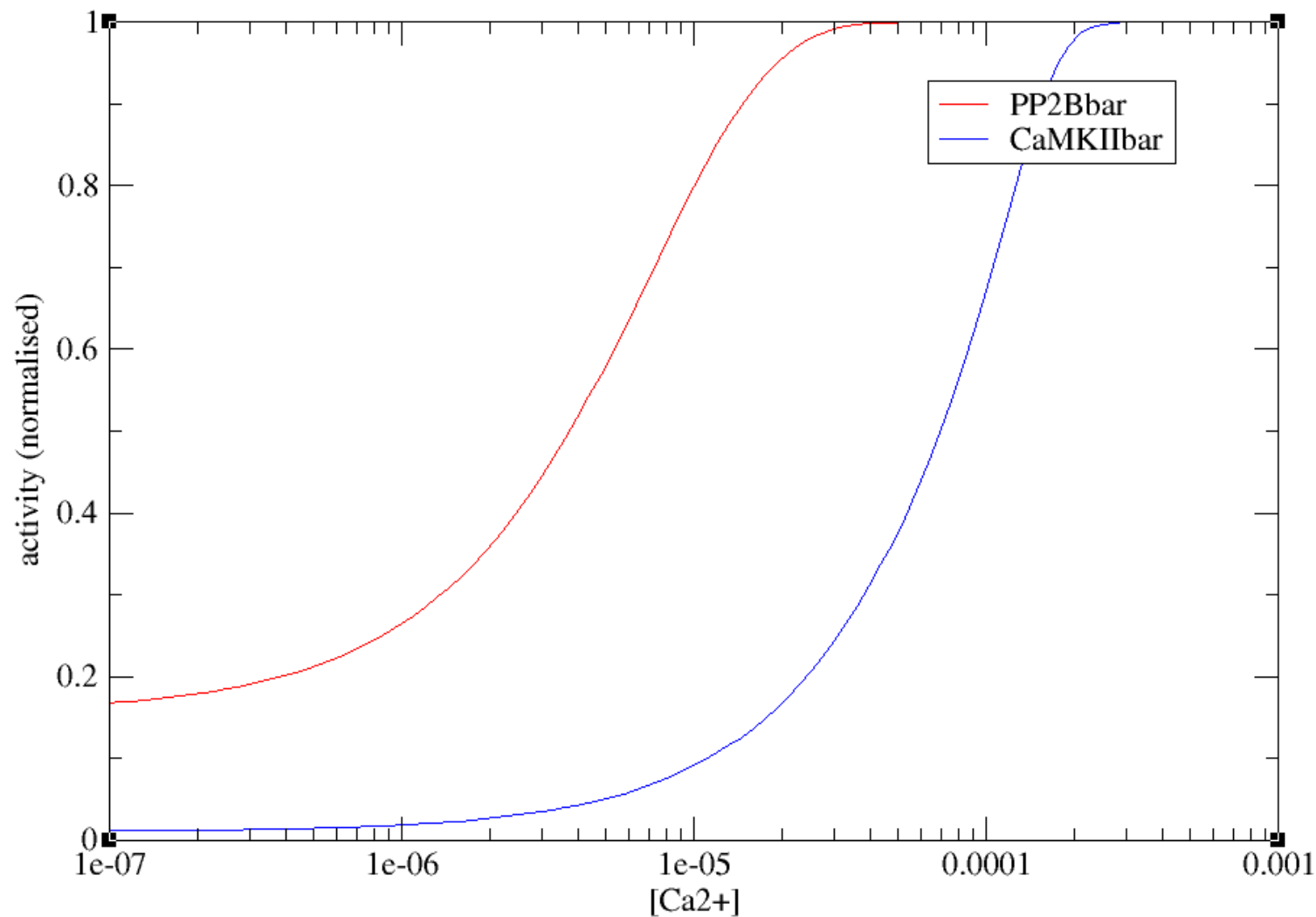
- Fractional activity depends on the number of calcium ions bound. E.g.:

$$\frac{R_2}{T_2} = \frac{6}{L(c_Ac_B + c_Ac_C + c_Ac_D + c_Bc_C + c_Bc_D + c_Cc_D)}$$

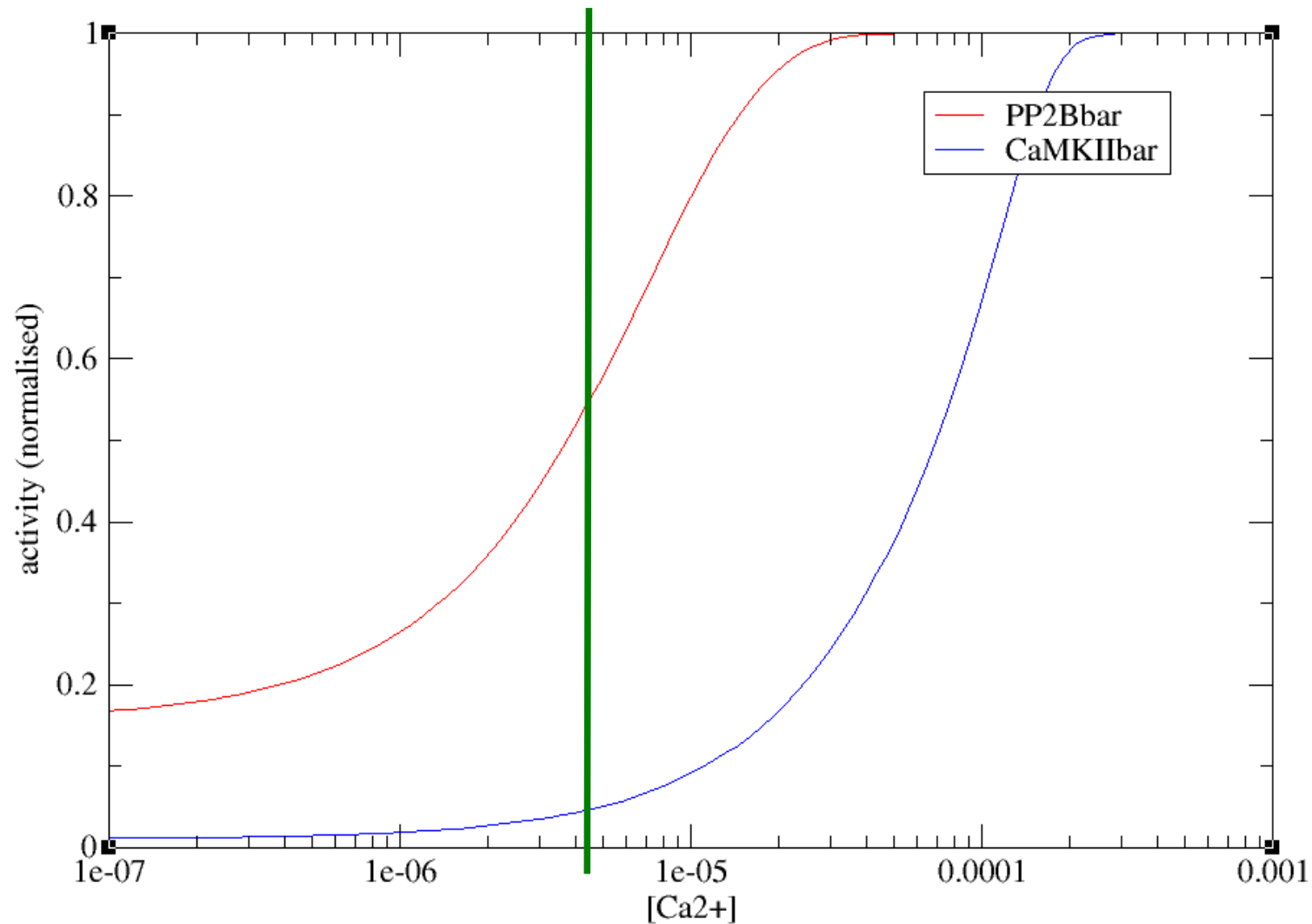
- $R_0/T_0 = 1/20000$ (1/L)
- $R_1/T_1 = 1/170$
- $R_2/T_2 = 0.69$  half-saturation = equi-probability
- $R_3/T_3 = 80$
- $R_4/T_4 = 10000$

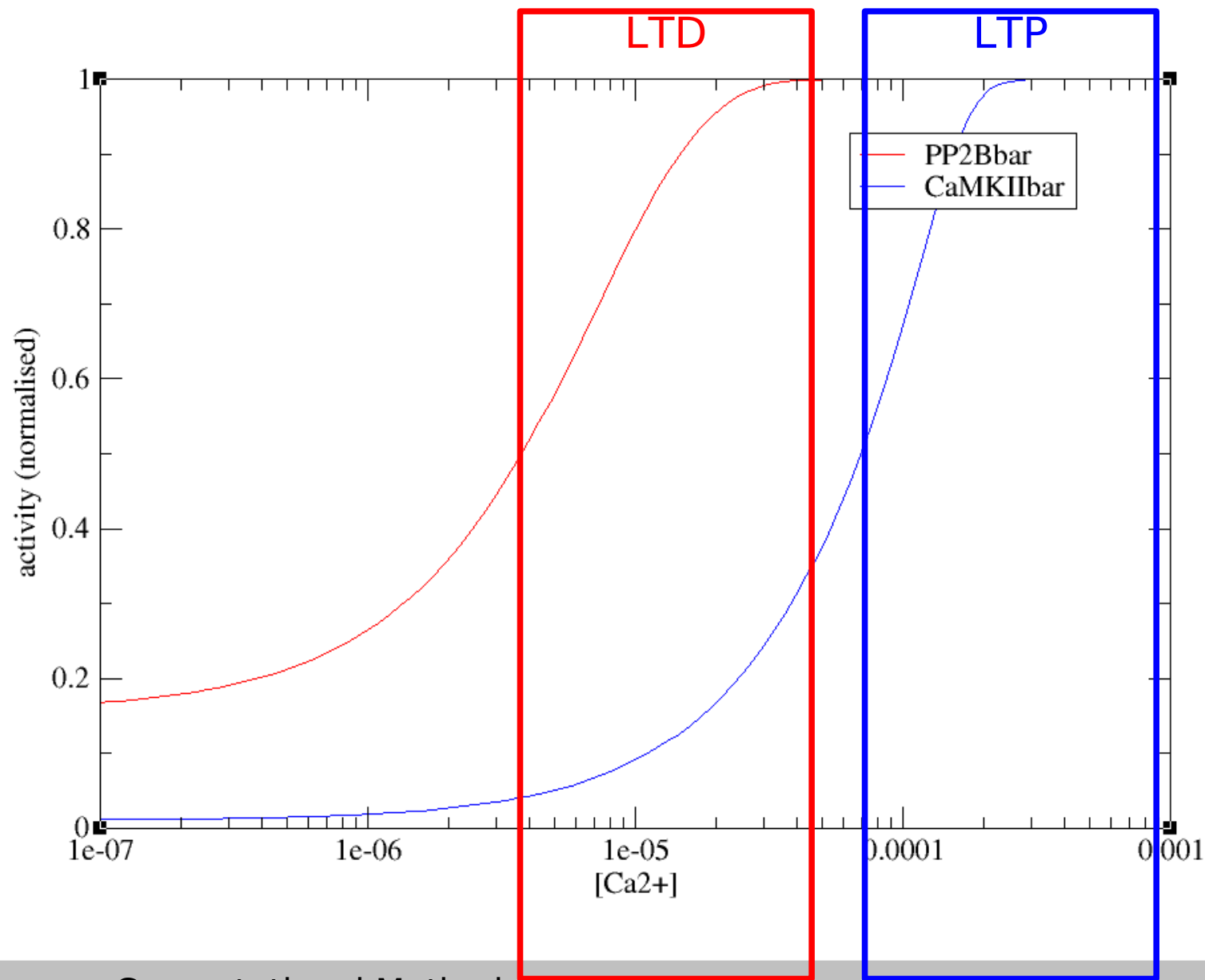






half saturation of calmodulin





- Developers of COPASI
 - Sven Sahle
 - Stefan Hoops
 - Ursula Kummer
 - Pedro Mendes
- Developers of Scilab
- Annalisa Pastore
- Stephen Martins



Melanie Stefan



Stuart Edelstsein

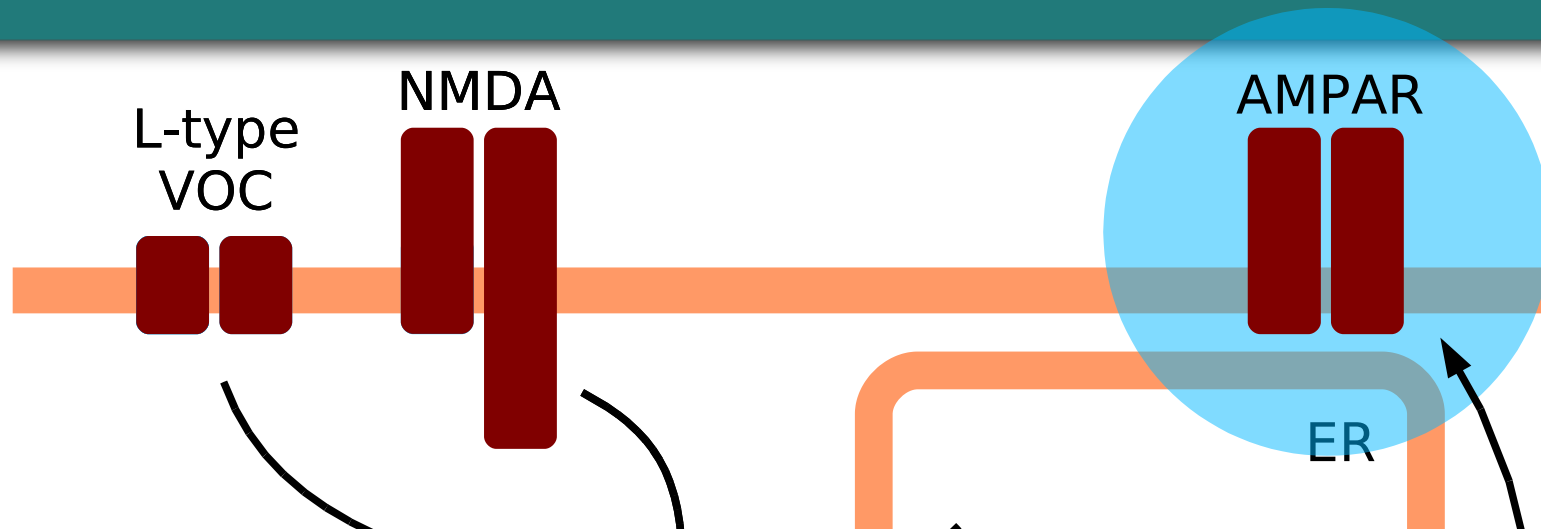
- Pros

- Trying to understand the “true” reaction mechanism
- Able to generate non-trivial predictions

- Cons

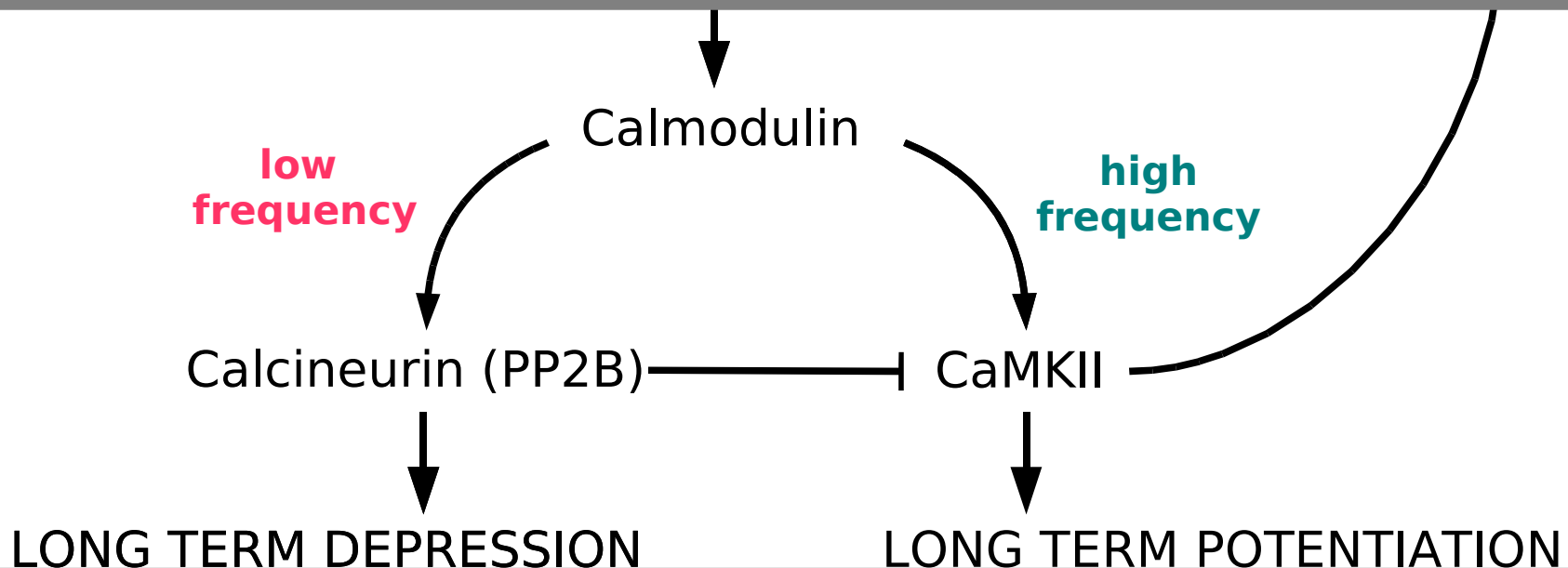
- Very sensitive to combinatorial explosion
- Almost complete lack of adequate quantitative data
- Cannot be easily embedded as such in larger models (but can be phenomenology abstracted)

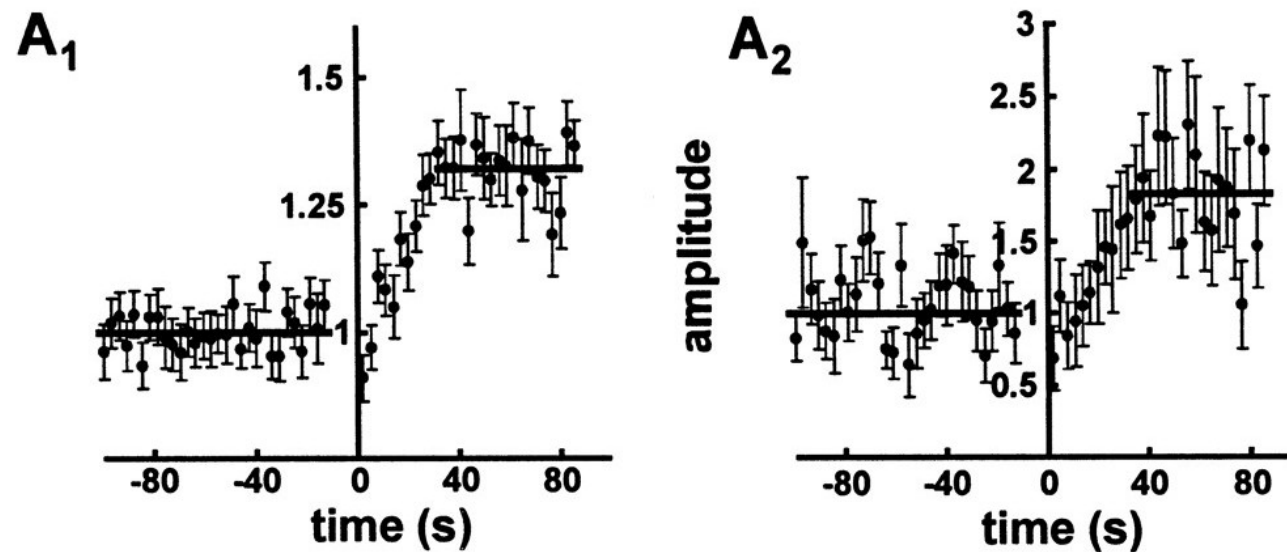


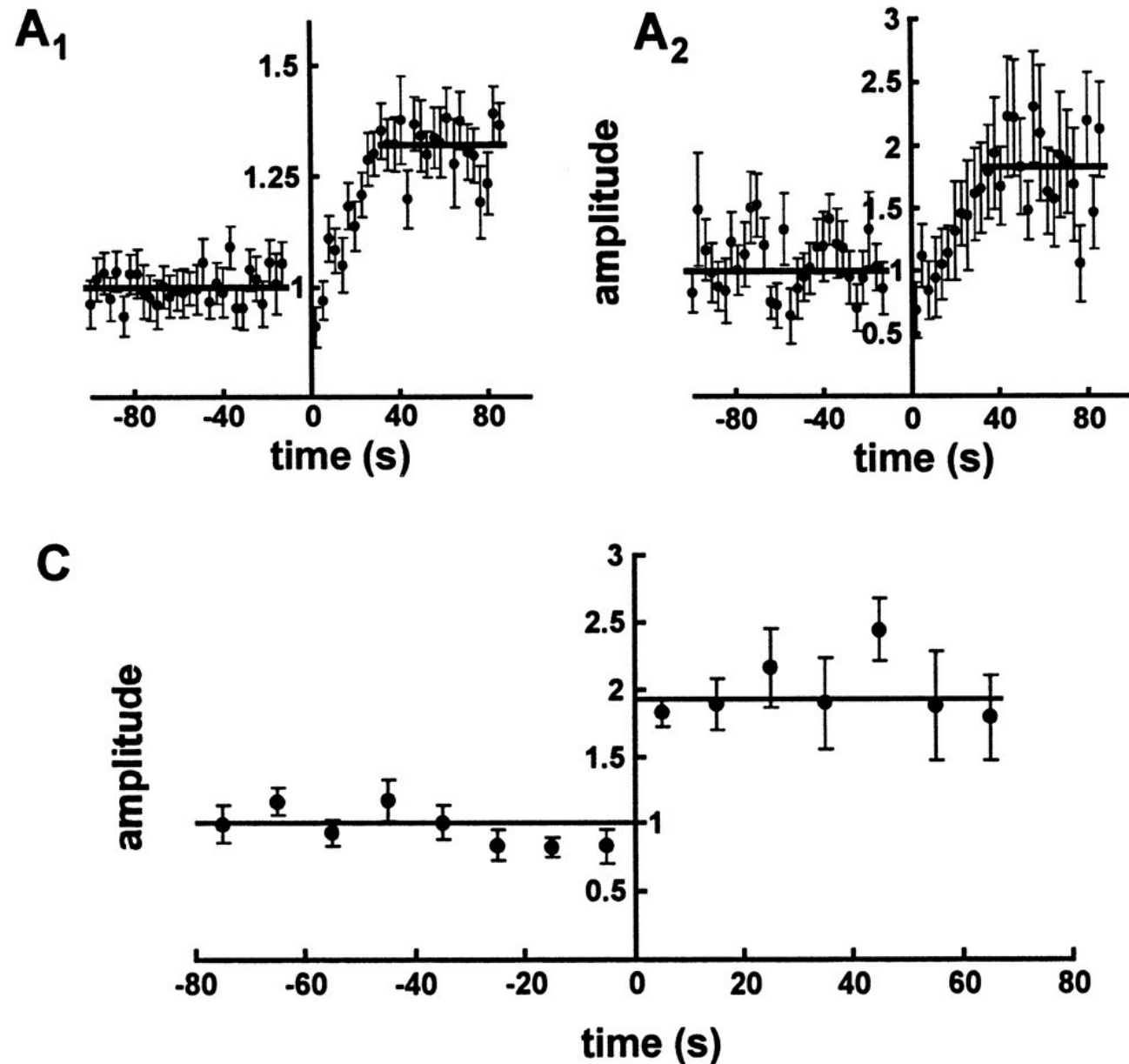


Tolle and Le Novère (2006) Particle-based Stochastic Simulation in Systems Biology. *Current Bioinformatics*, 1: 315-320

Tolle and Le Novère (2008) Brownian Diffusion of AMPA Receptors Is Sufficient to Explain Fast Onset of LTP. *Submitted*

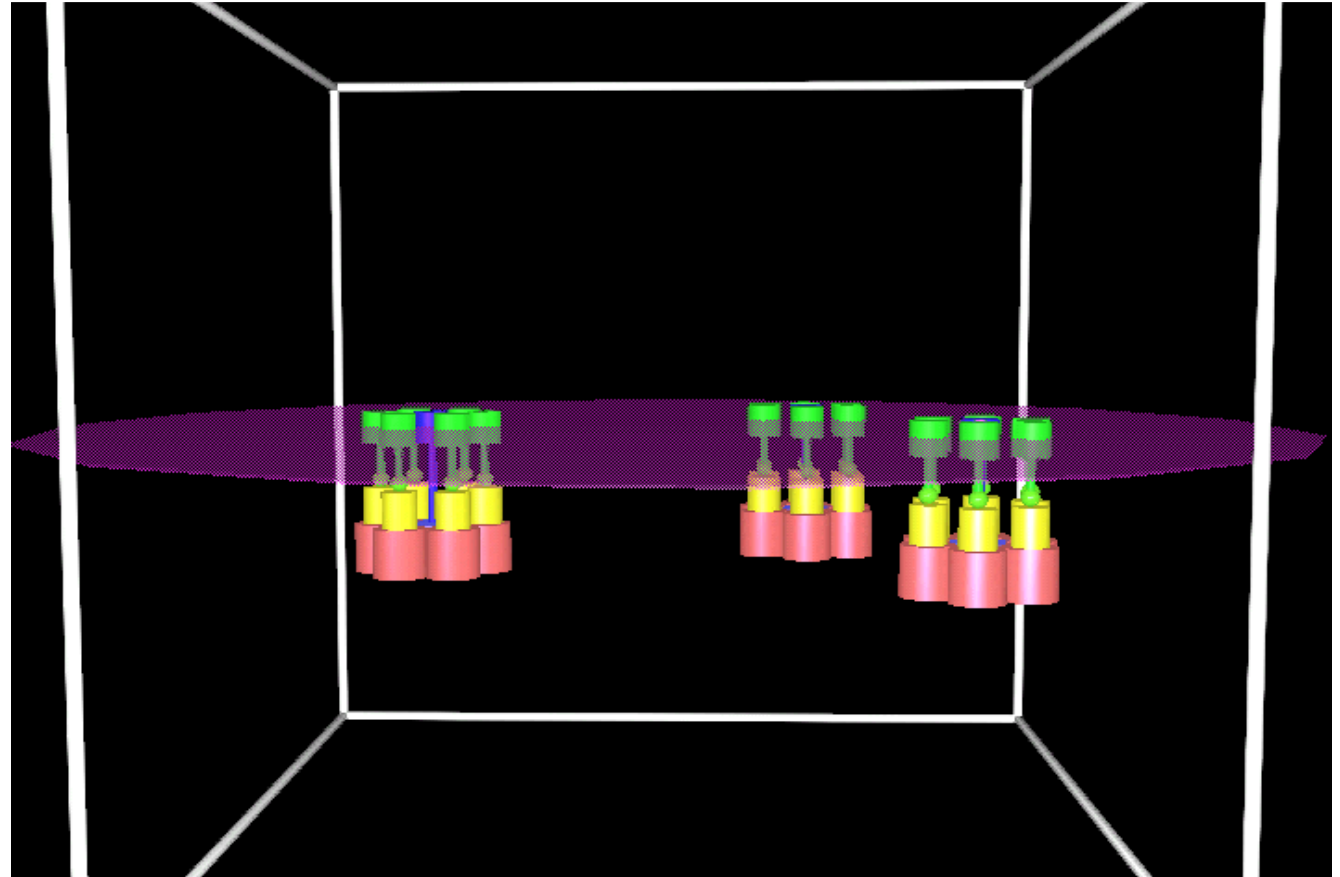
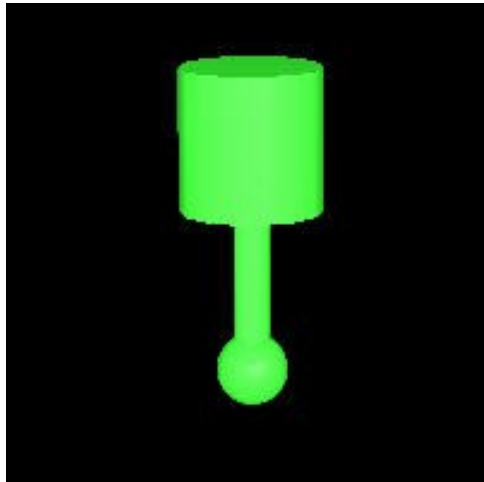
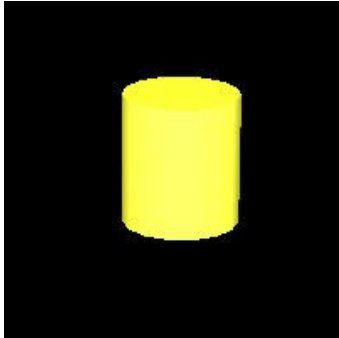






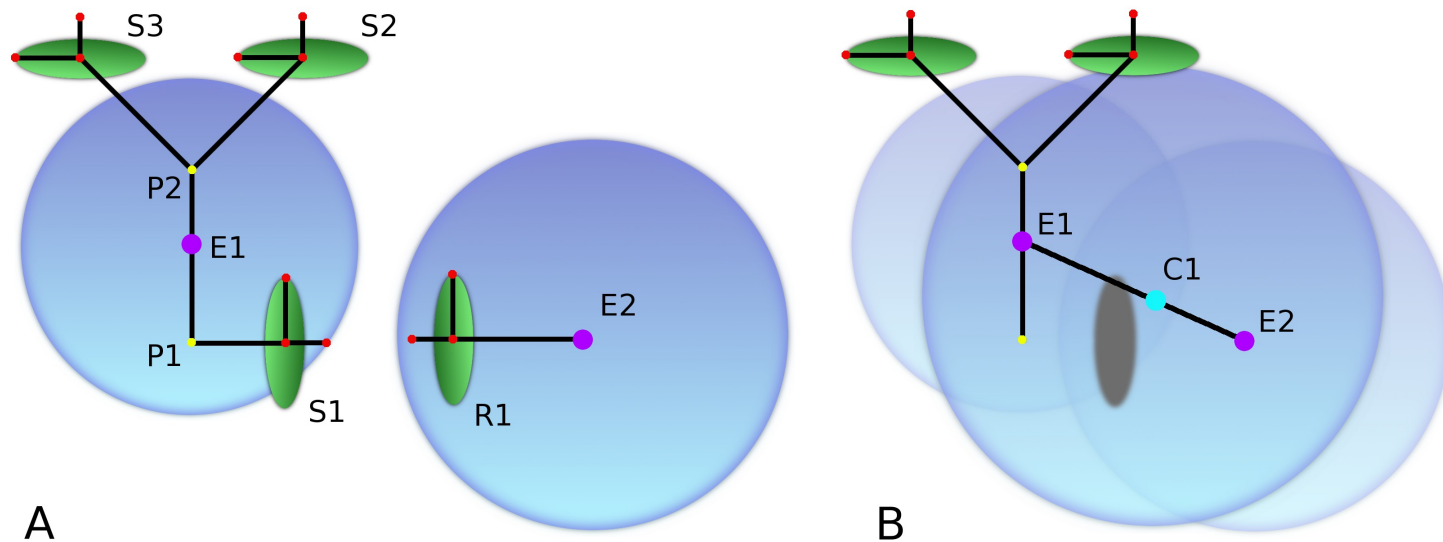
Petersen et al (1998)
PNAS 95: 4732-4737





- Particles carry binding surfaces
- Cluster and entity classes allows recording of Center Of Mass, radius, RMS displacement, state
- Clusters are dynamically created and destroyed – transient
- State variables attached to entities and reacting surfaces





- Different diffusion spaces:

- Static; Free diffusion; Membrane diffusion; Above membrane; Below membrane
- Membrane corrals, with different reflection properties

- Two types of motion:

- Translational $\overline{r^2} = 2D_T t = 2kb_T t$
- Rotational $\overline{\theta^2} = 2D_R t = 2kb_R t$

- random walk algorithm

$$p(x, t) = \frac{1}{\sqrt{4\pi Dt}} \exp\left(-\frac{x^2}{4Dt}\right) \quad \text{gaussian with} \quad \sigma^2 = 2Dt$$

- Translational $\Delta(x, y, z) = \sqrt{2D_T t} \times \text{gaussRand}$

- Rotational $\Delta\theta = \frac{\sqrt{2D_R t}}{r} \times \text{gaussRand}$

- Two types of diffusion equations:

- unrestricted brownian motion – Low Trans/Rot
- intra-membrane diffusion (Saffman and Delbrück 1975) – High Trans/Rot



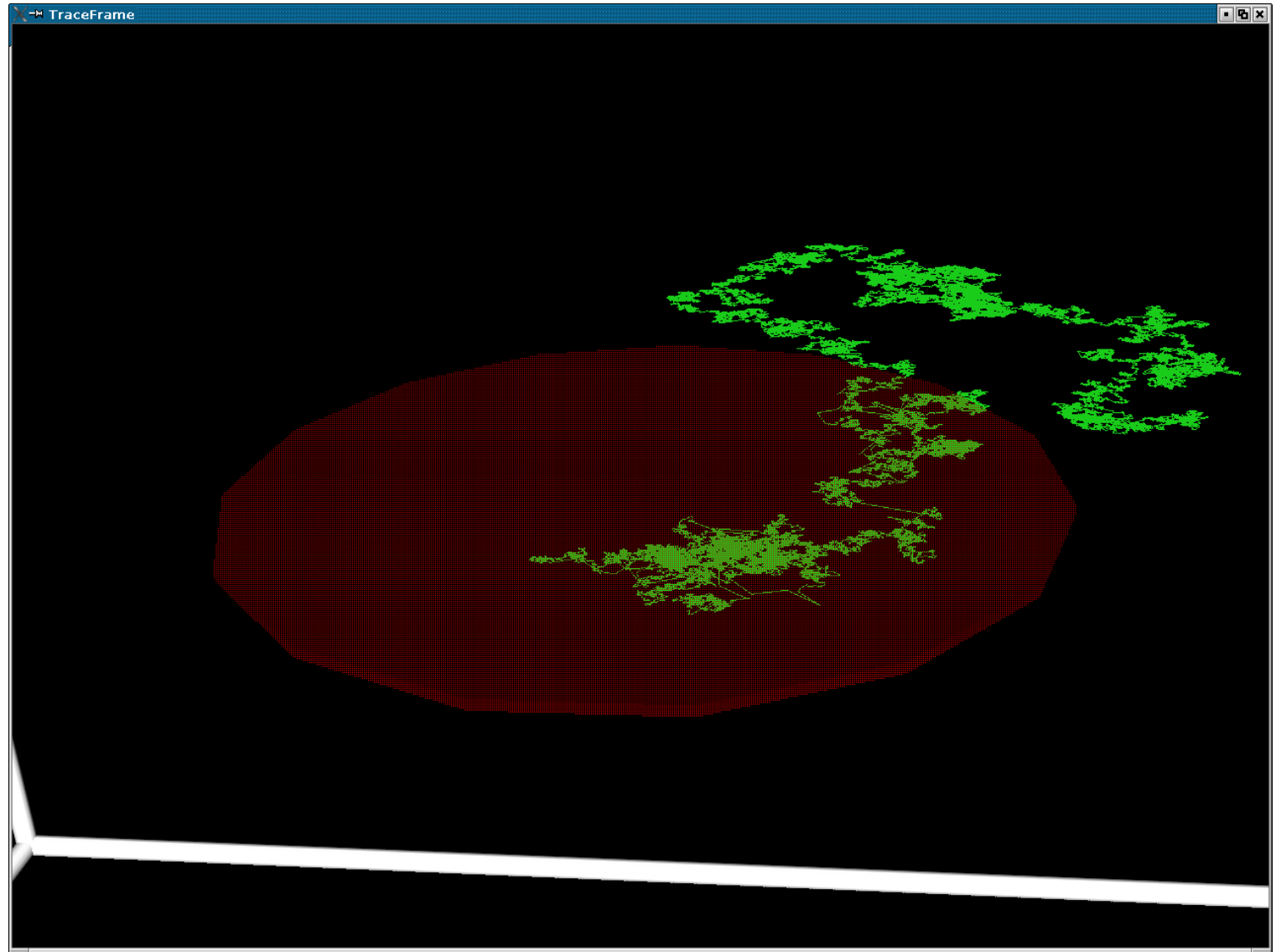
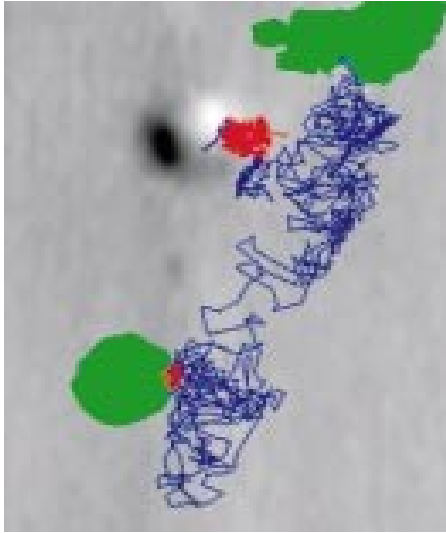
- Unrestricted brownian motion – Low Translation/Rotational

$$b_T = \frac{1}{4\pi\mu r} \quad b_R = \frac{1}{8\pi\mu r^3} \quad \frac{b_T}{b_R} = \frac{\xi}{4} r^2$$

- Intra-membrane diffusion (Saffman and Delbrück 1975)
High Translational/Rotational

$$b_T = \frac{1}{4\pi\mu h} \left(\log\left(\frac{\mu h}{\mu' r}\right) - \gamma \right) \quad b_R = \frac{1}{4\pi\mu r^3 h} \quad \frac{b_T}{b_R} = \left(\log\left(\frac{\mu h}{\mu' r}\right) - \gamma \right) \times r^2$$

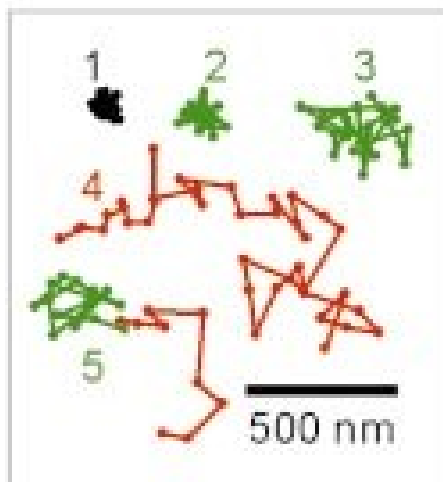




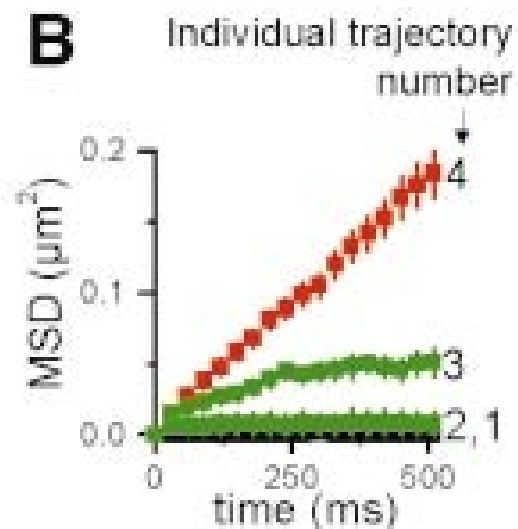
Borgdorff et al (2002)
Nature 417: 649-653



A



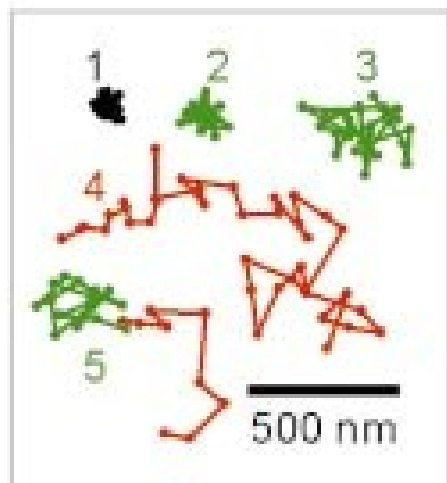
B



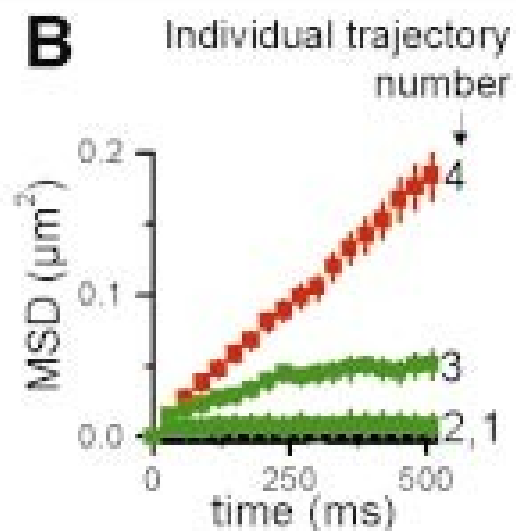
Tardin et al (2003)
EMBO J 22: 4656-4665



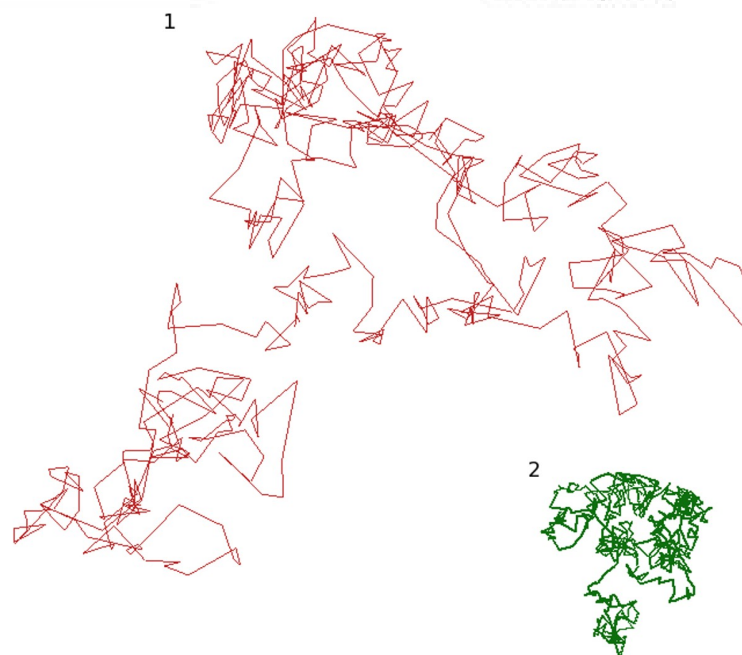
A



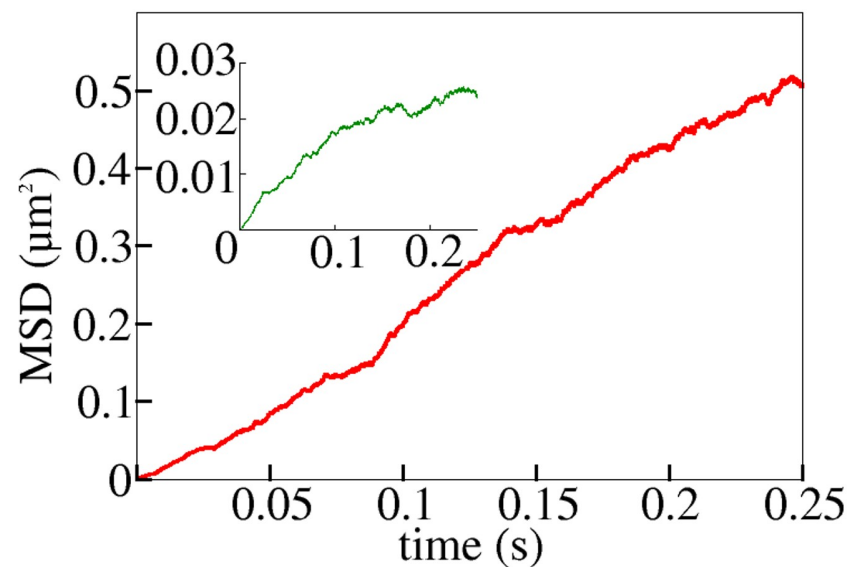
B



Tardin et al (2003)
EMBO J 22: 4656-4665



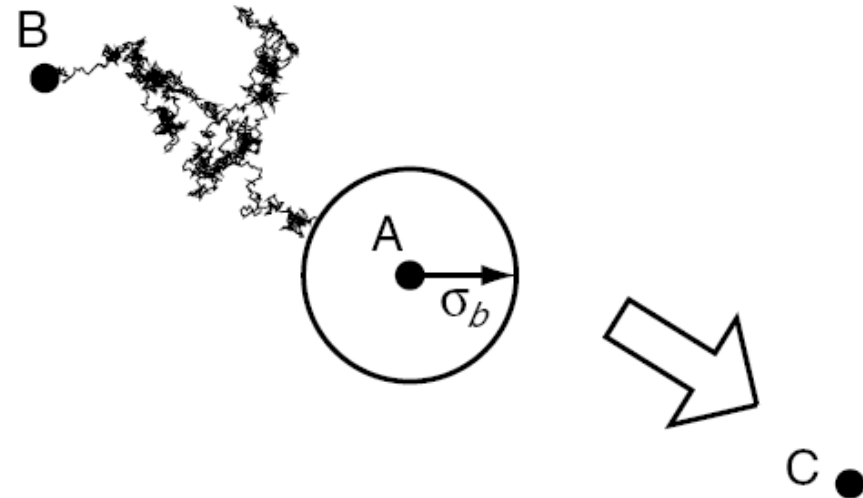
A



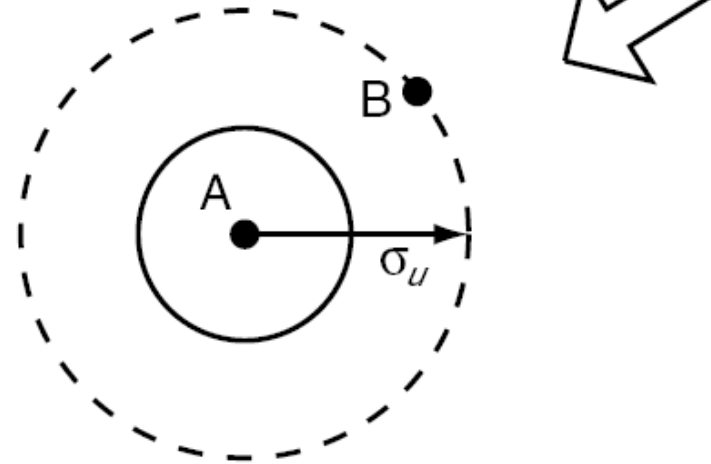
B



Forward reaction: $A + B \rightarrow C$

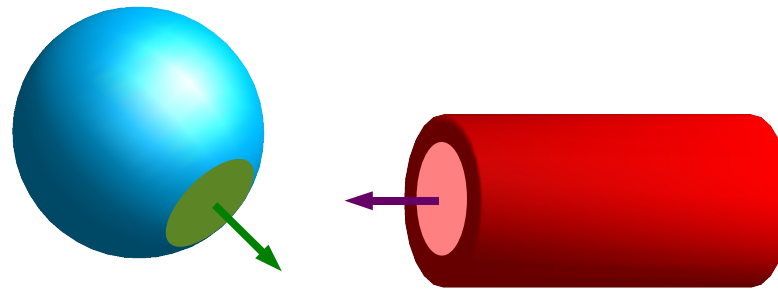


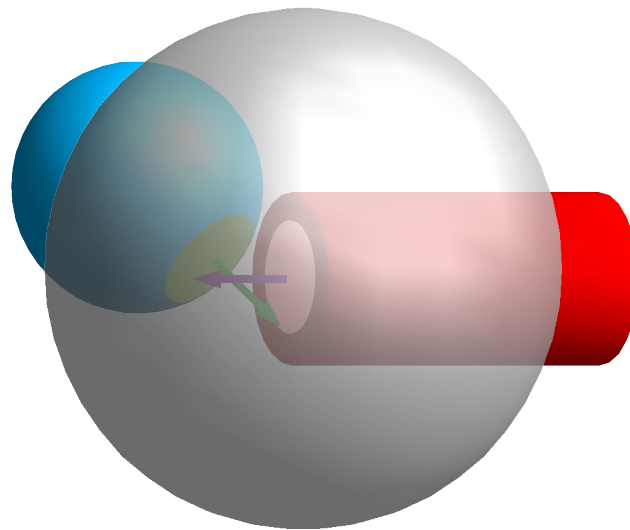
Back reaction: $C \rightarrow A + B$

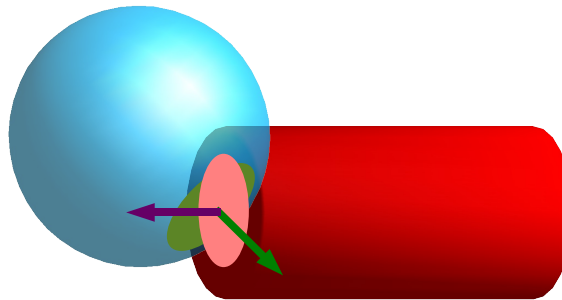


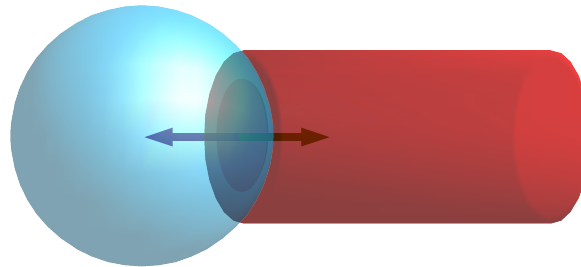
Andrews and Bray (2004)
Phys Biol 1: 137-151)

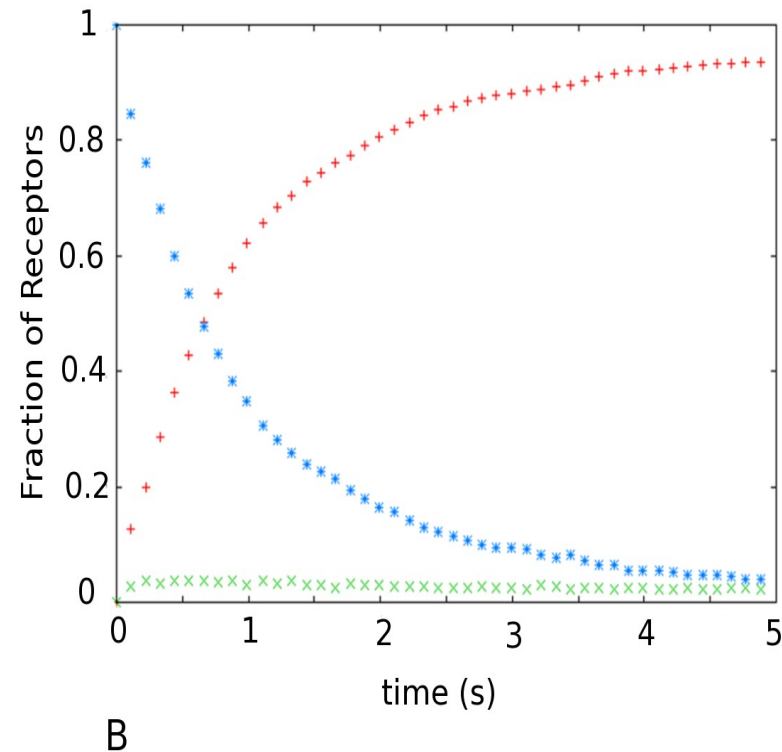
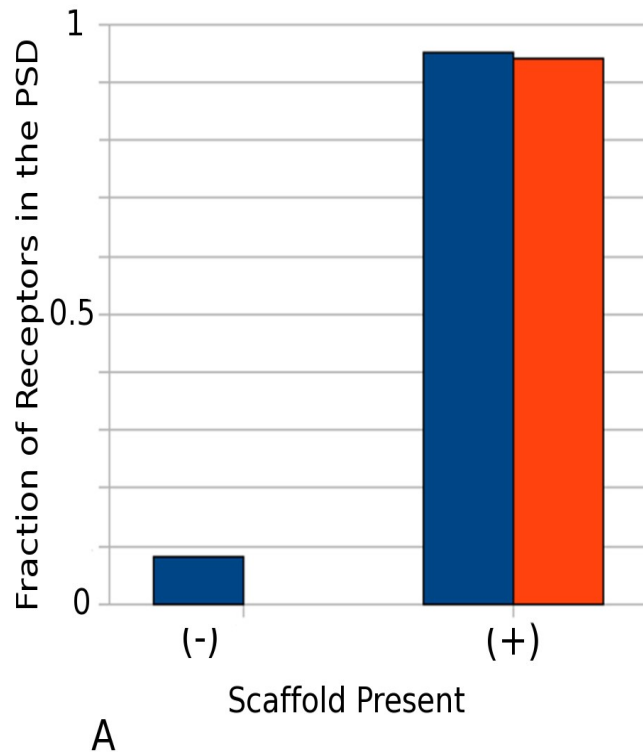






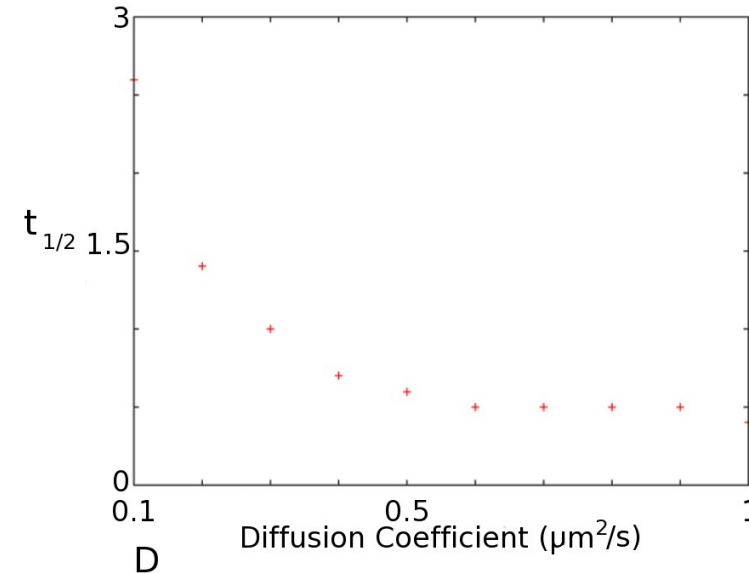
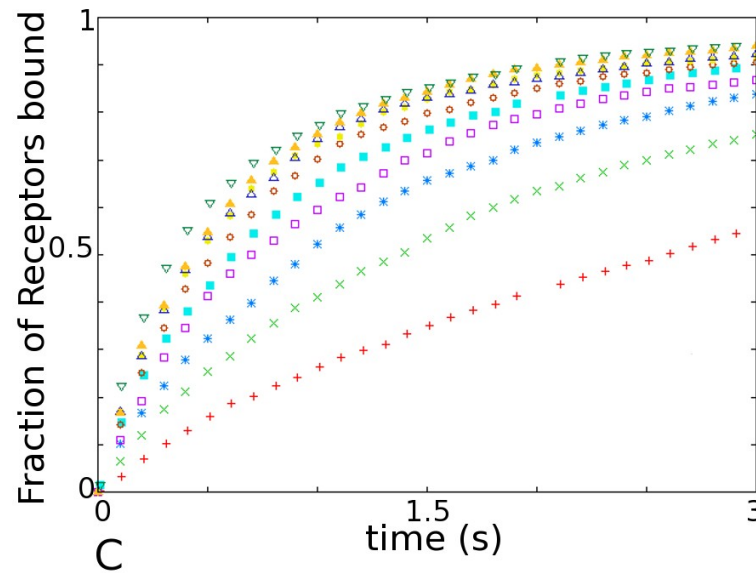
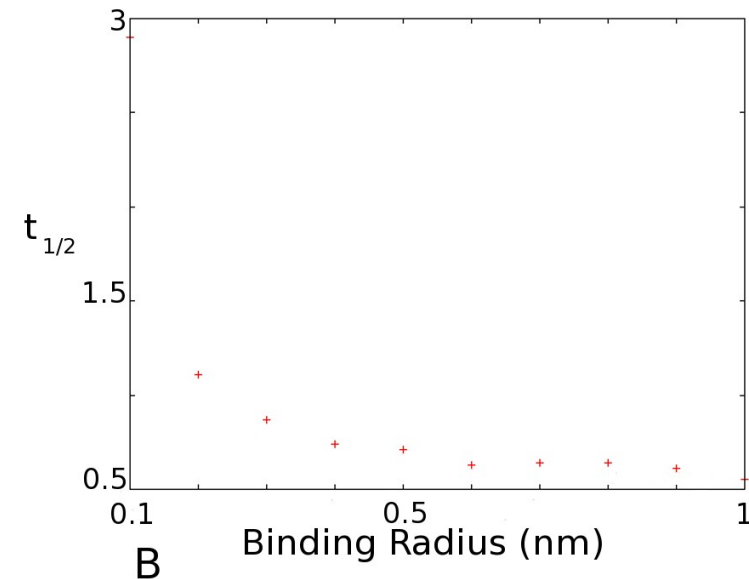
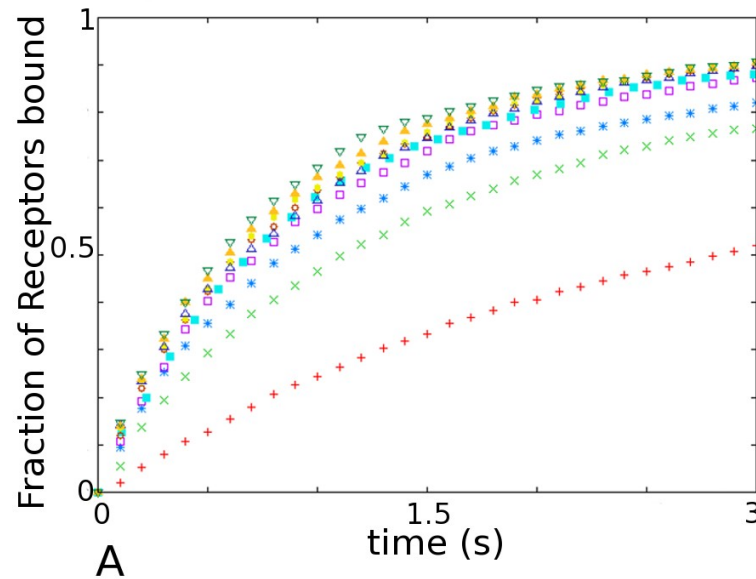


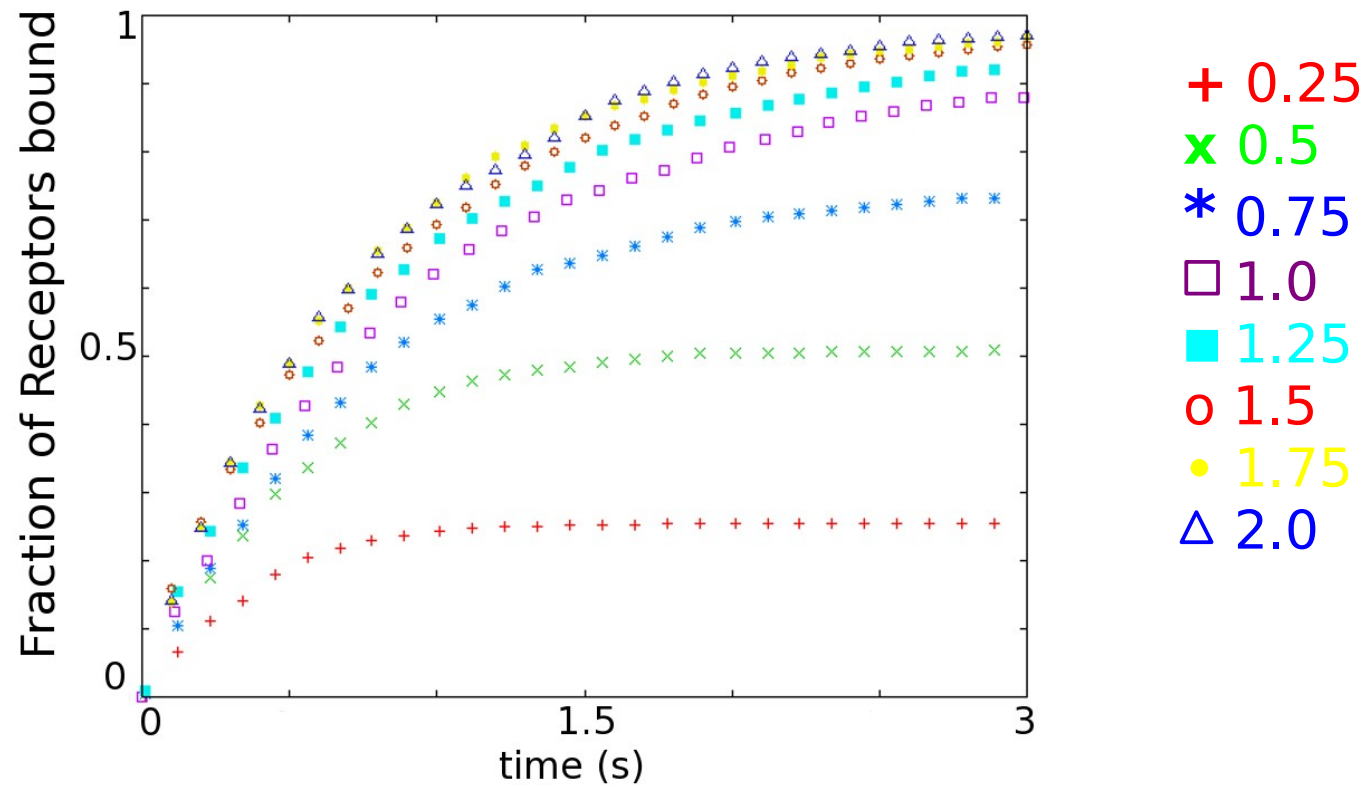


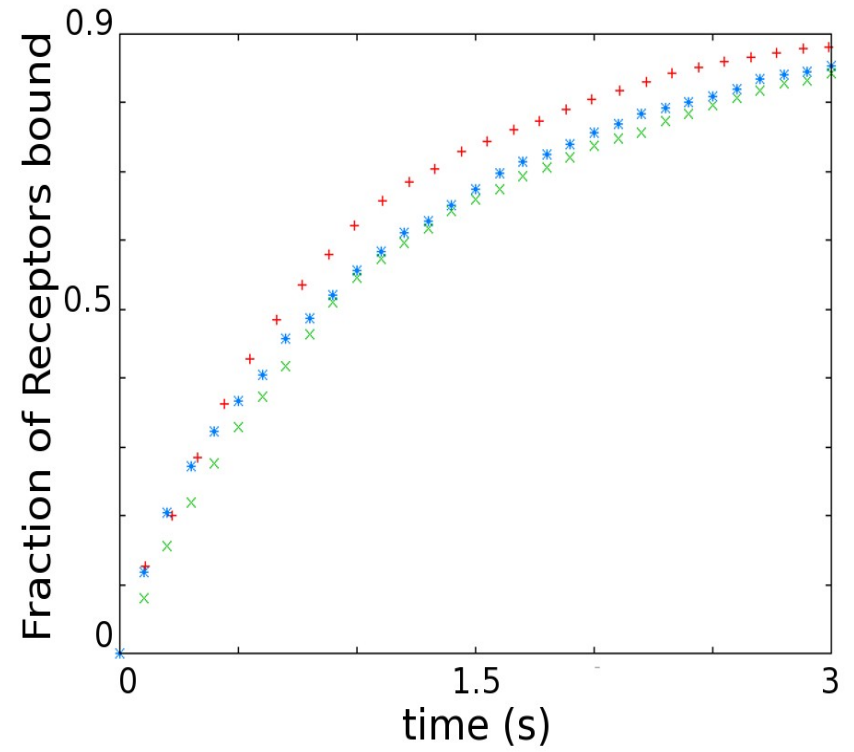
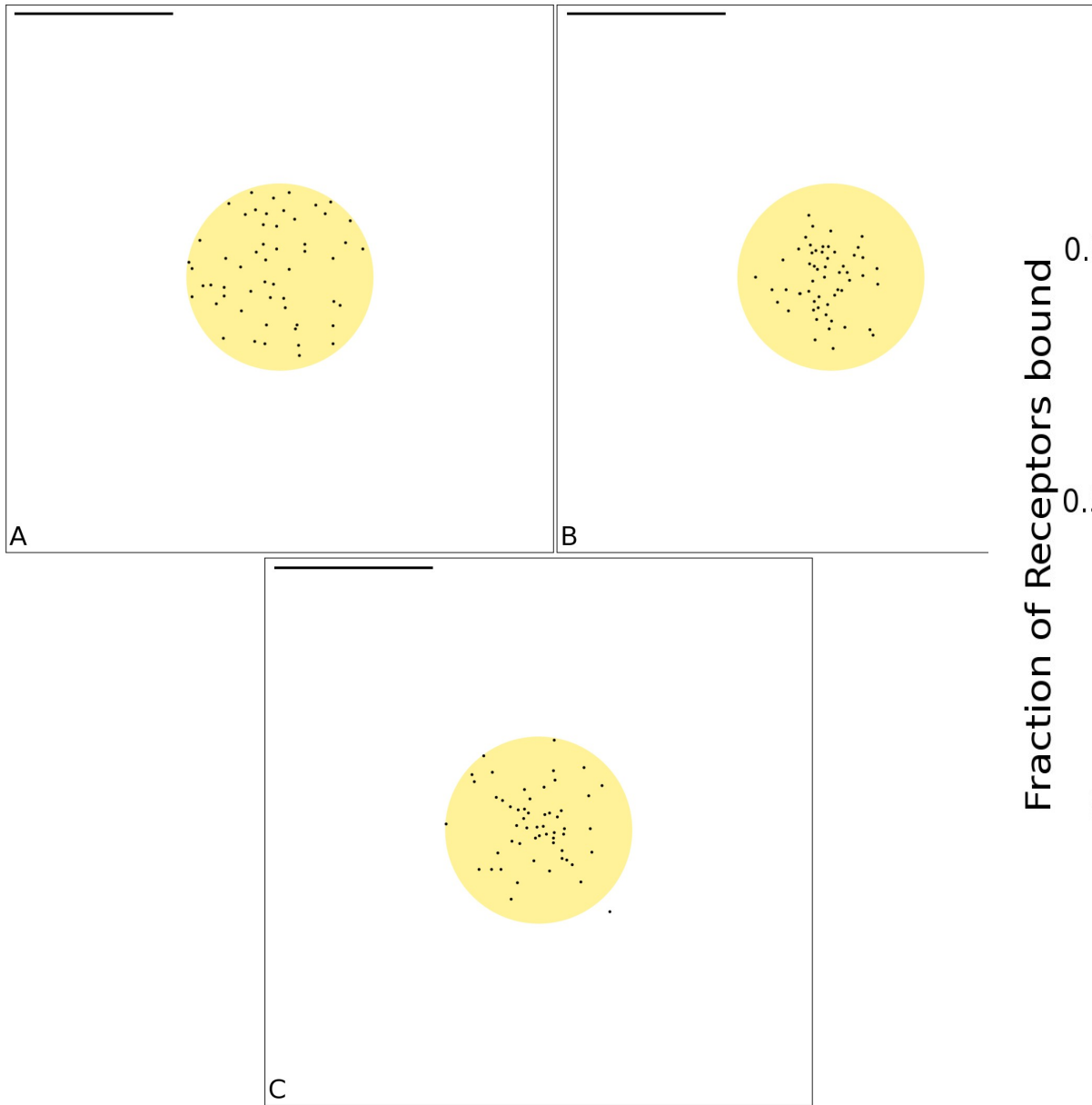


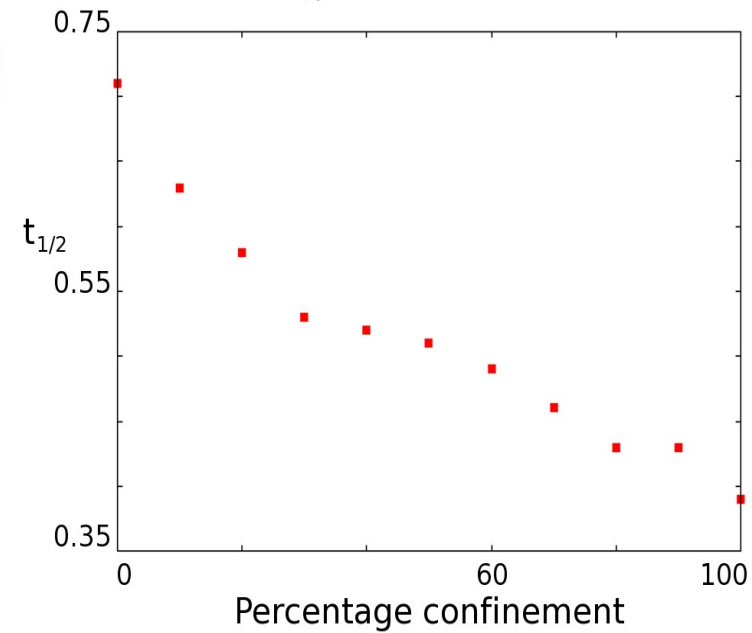
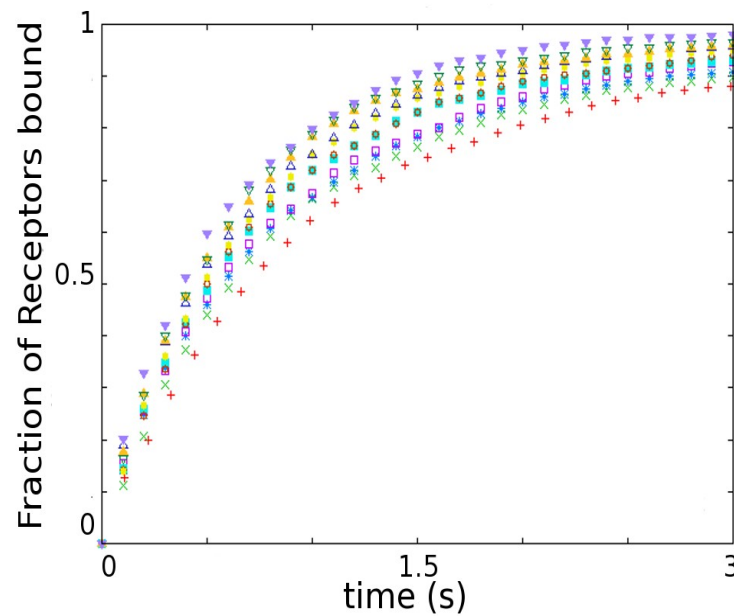
+ In PSD, bound
x In PSD, unbound
* In ESM

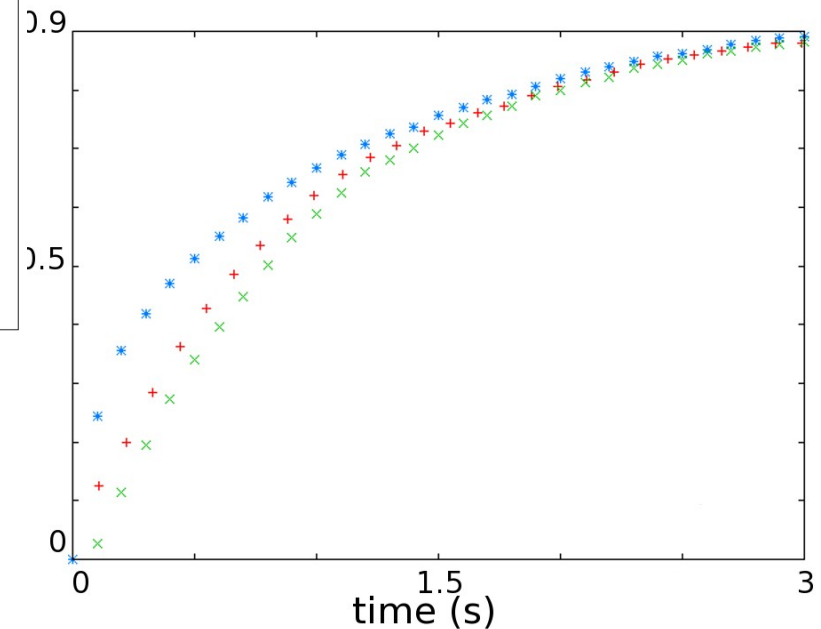
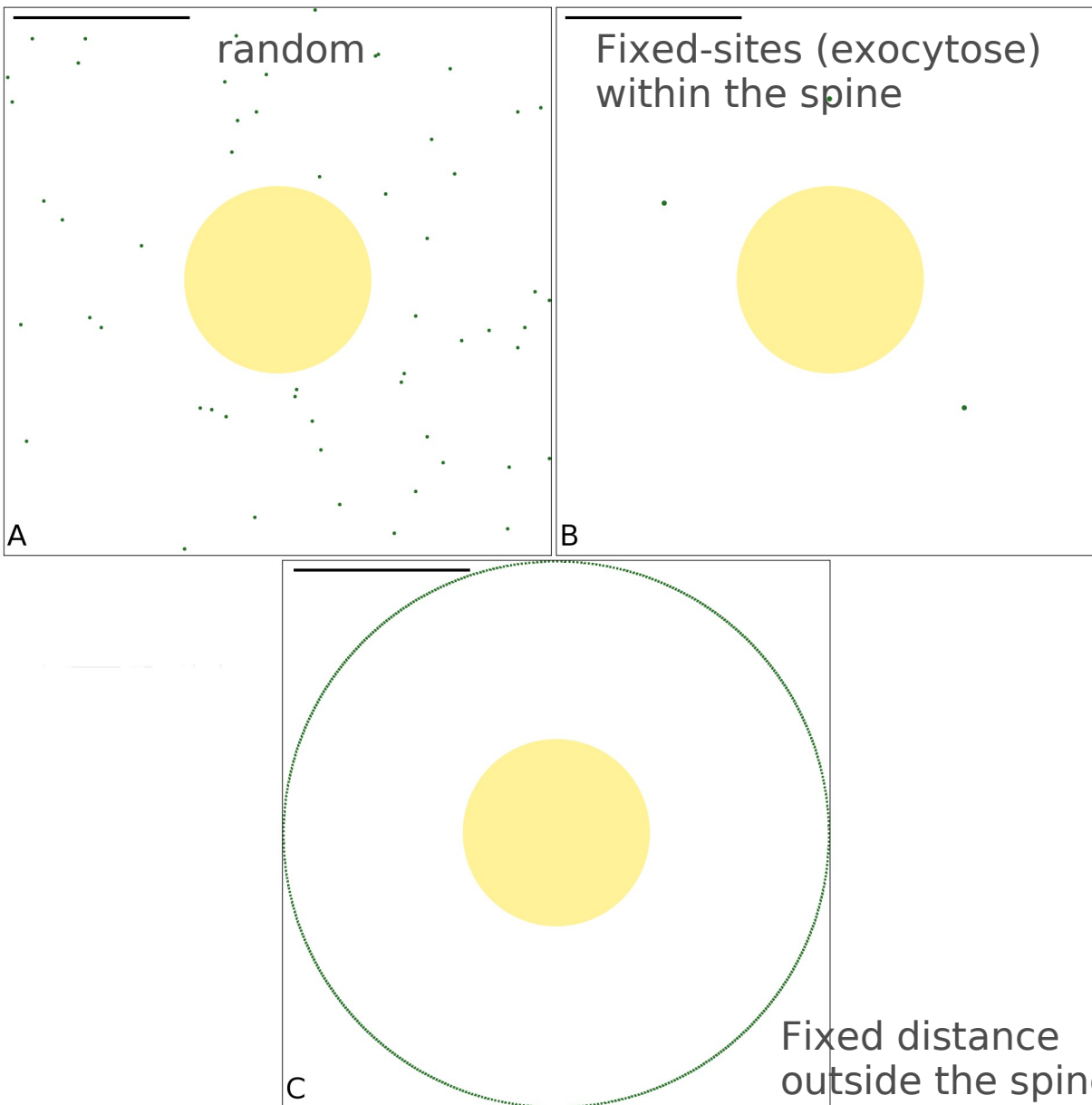












- If scaffolding proteins are present, 5 seconds are sufficient to cluster additional AMPA receptors to the post-synaptic density solely by “brownian” diffusion
 - Distribution of scaffolding proteins, and initial position of inserted receptors does not affect outcome
 - The crucial parameters are the ratio receptor/scaffold and the affinity of the scaffold proteins for the receptors
- => any process process modulating those characteristics will affect the plasticity of the synapse. This is the case of ... calmodulin-activated kinases ...



- Developers of APS
 - Fred Howell
 - Dan Mossop
- Steven Andrews



Dominic Tolle



■ Pros

- Able to accurately describe what is occurring in the cell (the spine)
- One can embed crowding, gradients, anisotropy of conditions etc.
- Agent-based approach avoid combinatorial explosion

■ Cons

- Heavy:
 - 1 second of post-synaptic density with 100 receptors ~ 1 month CPU
 - Voxelisation and computing over a cluster
1 second of synapse with 100 receptors ~ 1 day
 - 1 second of synapse generates ... 10 Terabytes of data (positions and orientation of each cluster at each timepoint)



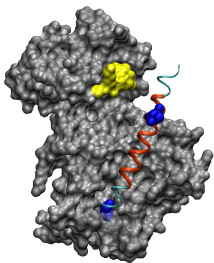
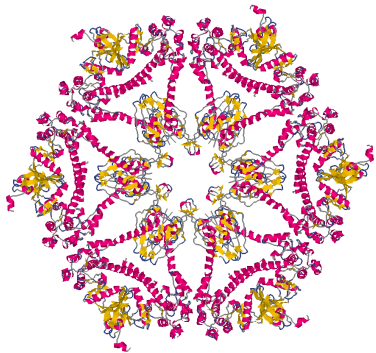
10^{-9}

10^{-6}

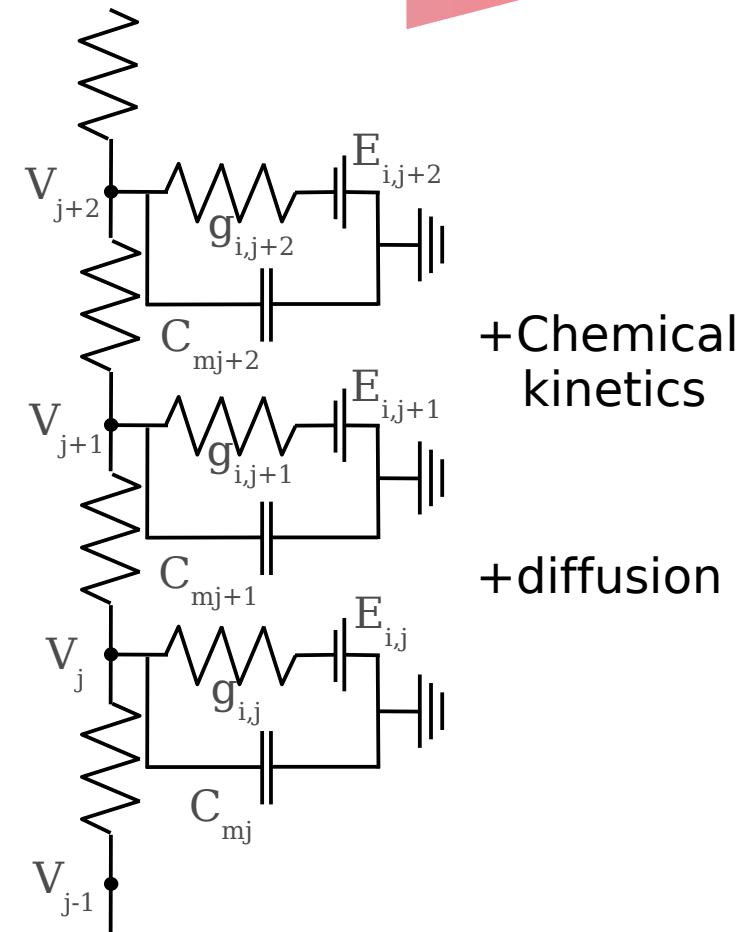
10^{-3}

m

spatial scale

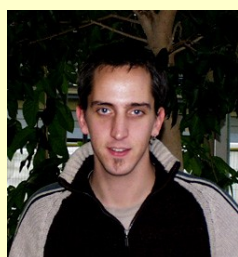


Scales presented today





Ranjita Dutta-Roy, SE



Dominic Tolle, EI, DE



Lu Li, CN



Nick Juty, UK



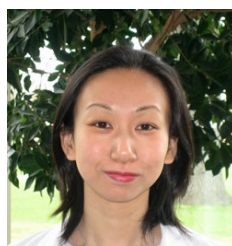
Camille Laibe, FR



Melanie Stefan, AU



Michele Mattioni, IT
Spatial simulation crew



Noriko Hiroi, JP

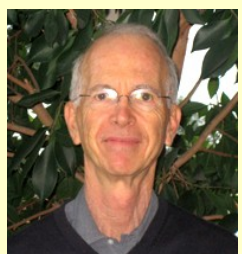
Signalling pathways crew



Christian Knüpfer, DE
Standard and ontology crew



Dagmar Köhn, DE

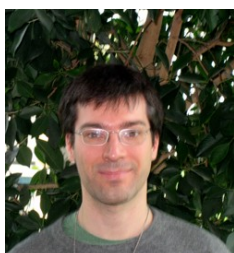


Stuart Edelstein, CH, US
Cooperativity crew

BioModels DB crew



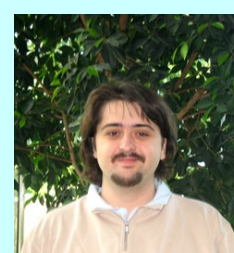
Viji Chelliah



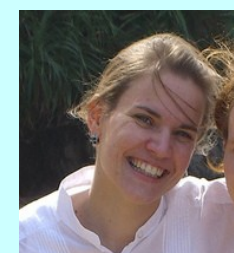
Lukas Endler



Chen Li



Duncan Berenguier, FR



SBML crew

Anika Oellrich, DE



Nicolas Rodriguez, FR



Kedar, IN

